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Jc912 U.S. PTO

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UTILITY PATENT
APPLICATION TRANSMITTAL

(Under 37 C.F.R. §1.53(b))

Docket No.: 237.00

Page 1 of 2

Title: Factor IX/Factor IXa Activating Antibodies and Antibody Derivatives

Express Mail Label No.: EL524823985US

Jc912 U.S. PTO
09/14/00

APPLICATION ELEMENTS

ADDRESS TO: Assistant Commissioner for Patents
Box Patent Application
Washington, D.C. 20231

INVENTORS: Friedrich Schefflinger, Randolph Kerschbaumer, Falko-Guenter Falkner, Friedrich Dörner, Hans-Peter Schwarz

1. ☒ Fee Transmittal Form (Submit an original, and a duplicate for fee processing)
2. ☒ Specification [Total Pages
3. ☒ Drawing(s) (35 USC §113) [Total Pages (Figs.1-36)
4. ☐ Signed Declaration [Total Pages

a. ☐ Newly executed (original or copy)

b. ☐ Copy from a prior application (37 CFR §1.63(d))

i. ☐ (Note Box 5 below)
DELETION OF INVENTOR(S)
Signed statement attached deleting inventor named in the prior application, see 37 CFR §§1.63(d)(2) and 1.33(b)

5. ☐ Incorporation By Reference (useable if Box 4b is checked.) entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein

6. ☐ Microfiche Computer Program

7. ☐ Nucleotide and/or Amino Acid Sequence Submission

8. ☐ Assignment Papers (cover sheet & document(s))

9. ☐ 37 CFR §3.73(b) Statement ☐ Power of Attorney

10. ☐ English Translation Document

11. ☐ Information Disclosure Statement (IDS)/PTO-1449 ☐ Copies of IDS Citations
12. ☐ Preliminary Amendment
13. ☒ Return Receipt Postcard (MPEP 503)
14. ☐ Small Entity Statement(s)
15. ☐ Certified Copy of Priority Documents (s)
- 16a. ☐ Cancel in the application original claims before calculating the filing fee
- 16b. ☐ Amend the specification by inserting before the first line the sentence: --This is a ___ Continuation, ___ Divisional, ___ Continuation-in-part, of application ___/___/___, filed ___/___/___
- 16c. ☐ I hereby verify that the attached papers are a true copy of the latest inventor-signed prior application, including a copy of the oath or declaration showing the original signature or an indication it was Signed. I hereby verify that the papers are a true copy of the latest Signed prior application number ___/___/___, filed ___/___/___ And further that all statements made herein of my own knowledge Are true; and further that these statements were made with the Knowledge that willful false statements and the like so made are Punishable by fine or imprisonment, or both, under Section 1001 Of Title 18 of the United States Code and that such willful false Statements may jeopardize the validity of the application or any Patent issuing thereon
- 16d. ☐ Other _____
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UTILITY PATENT
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Docket No.: 237.00 Page 2 of 2
Title: Factor IX/Factor IXa Activating Antibodies and Antibody
Derivatives
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CLAIMS	(1) FOR	(2) NUMBER	(3) NUMBER EXTRA	(4) RATE	(5) CALCULATIONS
	TOTAL CLAIMS (37 CFR 1.16(b))	24-20 =	4	X \$ 18 =	\$72
	INDEPENDENT CLAIMS (37 CFR 1.16(b))	3	0	\$ 78 =	\$0
	MULTIPLE DEPENDENT CLAIMS (if applicable) (37 CFR §1.16(d))			+ \$ 260 =	\$ 0
				BASIC FEE (37 CFR §1.16(a))	\$690
				TOTAL =	\$762

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- a. ☒ Fees required under 37 CFR §1.16
b. ☒ Fees required under 37 CFR §1.17
c. ☒ Fees required under 37 CFR §1.18

SIGNATURE OF APPLICANT, ATTORNEY, OR AGENT

NAME

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SIGNATURE

DATE

September 14, 2000

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CERTIFICATE OF MAILING UNDER 37 C.F.R. §1.10

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By: Diane M. Branham
Diane M. Branham

5 The present invention relates to actor IX/factor IXa-antibodies and antibody derivatives.

10 Blood clots (thrombi) are formed by a series of zymogen activations referred to as the coagulation cascade. In the course of this enzymatic cascade, the activated form of each of such zymogens (referred to as factors) catalyzes the activation of the next one. Thrombi are deposits of blood components on the surface of a blood vessel wall and mainly consist of aggregated blood platelets and insoluble, cross-linked fibrin.

15 Fibrin formation is effected by means of thrombin by limited proteolysis of fibrinogen. Thrombin is the final product of the coagulation cascade, (K.G. Mann, Blood, 1990, Vol. 76, pp.1-16).

20 Activation of factor X by the complex of activated factor IX (FIXa) and activated factor VIII (FVIIIa) is a key step in coagulation. The absence of the components of this complex or a disturbance of their function is associated with the blood coagulation disorder called hemophilia (J.E. Sadler & E.W. Davie: Hemophilia A, Hemophilia B and von Willebrand's disease, in G. Stamatoyanopoulos et al. (Eds.): The molecular basis of blood diseases. W.B. Saunders Co., Philadelphia, 1987, pp. 576-602). Hemophilia A denotes a (functional) absence of factor VIII activity, while Hemophilia B is characterized by the absence of factor IX activity.

30 At present, treatment of Hemophilia A is effected via a substitution therapy by administering factor VIII concentrates. However, approximately 20-30% of Hemophilia A patients develop factor VIII inhibitors (i.e. antibodies against factor VIII), whereby the effect of administered factor VIII preparations is inhibited. Treatment of factor VIII inhibitor patients

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is very difficult and involves risks, and so far there exist only a limited number of treatments for these patients.

In the case of patients having a low FVIII inhibitor level, it is possible, though expensive, to administer high doses of factor VIII to such patients and thus to neutralize the antibodies against factor VIII. The amount of factor VIII beyond that needed to neutralize the inhibitor antibodies then has hemostatic action. In many cases, desensitization can be effected, whereupon it is then possible again to apply standard factor VIII treatments. Such high dose factor VIII treatments require, however, large amounts of factor VIII, are time-consuming and may involve severe anaphylactic side reactions. Alternatively, the treatment may be carried out with porcine factor VIII molecules.

A further high-cost method involves removing factor VIII inhibitors through extra corporeal immunoadsorption on lectins which bind to immunoglobulins (protein A, protein G) or to immobilized factor VIII. Since the patient must be connected to an apheresis machine during this treatment, the treatment also constitutes a great burden on the patient. It is also not possible to treat an acute hemorrhage in this way.

At present, the therapy of choice is to administer activated prothrombin complex concentrates (APCC), such as FEIBA® and AUTOPLEX®, which are suitable for the treatment of acute hemorrhages even in patients having a high inhibitor titer (DE 31 27 318).

In the intravascular system of blood coagulation, the last step is the activation of factor X. This reaction is stimulated by the binding of factor VIIIa to factor IXa and the formation of a "tenase"-complex consisting of the factors IXa, VIIIa, X and phospholipid. Without the binding of FVIIIa, FIXa

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exhibits no or only a very slight enzymatic activity relative to FX.

Over the last several years, a number of possible binding sites for factor VIIa to factor IXa have been characterized, and it has been shown that antibodies or peptides which bind to these regions inhibit the activity of FIXa (Fay et al., J. Biol. Chem., 1994, Vol.269, pp.20522-20527, Lenting et al., J. Biol. Chem., 1996, Vol. 271, pp. 1935-1940, Jorquera et al., Circulation, 1992, Vol. 86, Abstract 2725). The inhibition of coagulation factors, such as factor IX, has also been achieved through the use of monoclonal antibodies with the aim of preventing thrombosis formation (WO 97/26010).

The opposite effect, i.e. an increase in the factor IXa mediated activation of factor X, has been described by Liles D.K. et al., (Blood, 1997, Vol. 90, suppl. 1, 2054) through the binding of a factor VIII peptide (amino acids 698-712) to factor IX. Yet, this effect only occurs in the absence of factor VIIa, while in the presence of factor VIIa the factor IXa/factor VIIa-mediated cleavage of factor X is inhibited by this peptide.

SUMMARY OF THE INVENTION

With a view to the possible risks and side effects which may occur in the treatment of hemophilia patients, there is a need for a therapy which allows for the effective treatment of FVIII inhibitor patients. Therefore, it is an object of the present invention to provide a preparation for the treatment of blood coagulation disorders which has particular advantages for factor VIII inhibitor patients.

According to the present invention, this object is achieved through the use of antibodies or antibody derivatives against factor IX/factor IXa which have factor VIIa-cofactor activity or factor IXa-activating

activity and lead to an increase in the procoagulant activity of factor IXa. Surprisingly, the action of these inventive factor IX/factor IXa-activating antibodies and antibody derivatives is not negatively affected by the presence of inhibitors, such as inhibitors against factor VIII/factor VIIIA, but instead the procoagulant activity of factor IXa in this case also is increased.

A further advantage of this invention is that the administration of the preparation according to the invention allows for rapid blood coagulation even in the absence of factor VIII or factor VIIIA, even in the case of FVIII inhibitor patients. Surprisingly, these agents are also effective in the presence of factor VIIIA.

The antibodies and antibody derivatives according to the present invention thus have a FVIII-cofactor-like activity which, in a FVIII assay (e.g. a COATEST® assay or Immunochrom test) after 2 hours of incubation exhibits a ratio of background (basic noise) to measured value of at least 3. Calculation of this ratio may, e.g., be effected according to the following scheme:

$$\frac{\text{Antibody measurement (OD 405) - blank value from reagent}}{\text{Mouse-IgG-measurement (OD 405) - blank value from reagent}} \geq 3$$

after two hours of incubation.

The antibodies according to the invention preferably have an in vivo half life of at least 5 days, more preferably at least 10 days, though it is more preferred to have a half life of at least 20 days.

A further aspect of this invention is a preparation comprising antibodies and/or antibody derivatives against factor IX/factor IXa and a pharmaceutically acceptable carrier substance. Furthermore, the preparation according to the invention may additionally comprise factor IX and/or factor IXa.

A further aspect of the invention is the use of the antibodies or antibody derivatives to increase the amidolytic activity of factor Ixa.

Fig. 1 shows the results of a screening of
5 supernatants from hybridoma cell cultures for FVIII-like activity. Pre-selected clones from fusion experiments, #193, #195 and #196, were tested in a chromogenic assay.

Fig. 2 shows the results of screening for IgG-mediated factor VIII-like activity in supernatants of a
10 hybridoma cell culture of a master plate.

Fig. 3 shows the subcloning of clone 193/C0, namely the results of the first cloning round.

Fig. 4 shows a comparison of the chromogenic FVIII-like activity and factor IX-ELISA-reactivity of
15 hybridoma cultures derived from the starting clone 193/C0.

Fig. 5 shows the results of the measurement of the chromogenic activity of some master clones and sub-clones.

Fig. 6A shows the FVIII-like activity of the anti-FIX/FIXa-antibodies 193/AD3 and 196/AF2 compared to human FVIII, TBS buffer and cell culture medium. After a lag phase, both antibodies gave rise to chromogenic substrate cleavage, as judged by the increasing optical
20 density.

Fig. 6B shows a comparison of the chromogenic activity of factor VIII, 196/AF1, 198/AC1/1 and mouse-IgG.

Fig. 7A shows a comparison of the kinetics of
30 Factor Xa generation by Factor VIII and 196/AF2 with and without the addition of a Factor Xa specific inhibitor.

Fig. 7B shows a comparison of the kinetics of the Factor Xa generation by Factor VIII, mouse-IgG and anti-factor IX/IXa-antibody 198/AM1 with and without the
35 addition of a factor Xa-specific inhibitor, Pefabloc Xa[®].

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Fig. 8A shows a measurement of the dependence of the factor VIII-like activity of purified anti-factor IX/IXa-antibody 198/AC1/1 in the presence and absence of phospholipids, FIXa/FX and calcium ions.

5 Fig. 8B shows a measurement of the dependence of FXa generation by anti-FIXa-antibody 196/AF1 in the presence of phospholipids, Ca^{2+} in FIXa/FX.

Fig. 8C shows the generation of FXa by unspecific mouse IgG antibody.

10 Fig. 9 is a graphical representation of the coagulation times of Factor VIII-deficient plasma in an APTT assay by using various concentrations of anti-factor IX/IXa-antibody 193/AD3.

15 Fig. 10A shows that in the presence of Factor IXa, antibody 193/AD3 leads to a reduction in the coagulation time of factor VIII-deficient plasma.

Fig. 10B shows a dose-dependent reduction of the clotting time by antibody 193/AD3 in the presence of factor IXa- and factor VIII-inhibitors.

20 Fig. 11 shows the chromogenic activity of antibodies 198/A1, 198/B1 and 198/AP1 in the presence and absence of human FIXa β .

Fig. 12 shows the primer sequences for the amplification of the genes of the variable heavy chain of mouse antibody.

25 Fig. 13 shows the primer sequences for the amplification of the genes of the variable light (kappa) chain of the mouse antibody.

30 Fig. 14 shows the DNA and derived protein sequence of the scFv from hybridoma cell line 193/AD3 (SEQ.ID.NOs. 81 and 82).

Fig. 15 shows the DNA and derived protein sequence of the scFv from hybridoma cell line 193/K2 (SEQ.ID.NOs. 83 and 84).

35 Fig. 16 shows the DNA and derived protein sequence of the scFv from hybridoma cell line 198/AB2 (subclone of 198/B1) (SEQ.ID.NOs. 85 and 86).

Fig. 17 shows the DNA and deduced protein sequence of scFv derived from the cell line 198/A1 (SEQ.ID.NOs. 87 and 88).

Fig. 18 demonstrates the chromogenic FVIII-like activity of peptide A1/3 in the presence of 2.9nM human FIXa. The scrambled version of peptide A1/3, peptide A1/5 does not give rise to any FXa generation.

Fig. 19 demonstrates the dependence of the chromogenic FVIII-like activity of peptide A1/3 on the presence of human FIXa. In the absence of human FIXa, peptide A1/3 does not give rise to any FXa generation. The buffer control, plain imidazole buffer is designated IZ.

Fig. 20 shows that the chirality of Arg-residues does not play a significant role for the chromogenic activity of peptides A1/3-rd and A1/3-Rd-srmb.

Fig. 21 shows that the addition of 2.4μM peptide B1/7 to the reaction mixture led to a measureable generation of Fxa.

Fig. 22 shows that the addition of a FX-specific inhibitor results in a significant reduction in the reaction. If there was no FIXa and FX is added to the reaction mixture, no FXa was synthesized.

Fig. 23 shows vector pBax-IgG1.

Fig. 24 shows the increase of the amidolytic activity of FIXa in the presence of antibody 198/B1 (Fig. 24A) and IgM antibody 198/AF1 (Fig. 24B).

Fig. 25 demonstrates the chromogenic FVIII-like activity of the antibody 198/A1 Fab fragment in the presence of 2.3nM human FIXa. As a positive control the intact antibody 198/A1 was used as well as 7.5pM FVIII. The buffer control (IZ) was used as a negative control.

Fig. 26 shows the nucleotide and amino acid sequence of the 198AB2 scFv-alkaline phosphatase fusion

protein (ORF of the expression vector pDAP2-198AB2#100, (SEQ.ID.NOs. 89 and 90).

The genes for the VL and the VH domains of antibody 198/AB2 (198/AB2 is an identical subclone of 198/B1) were derived from the corresponding hybridoma cells as described in example 10. The PCR product of the VH-gene was digested SfiI - AscI and the PCR-product of the VL-gene was digested AscI and NotI. VH and VL genes were linked via the AscI site and inserted into SfiI - NotI digested vector pDAP2 (Kerschbaumer R.J. et al, Immunotechnology 2, 145-150, 1996; GeneBank accession No.:U35316). PelB leader: leader sequence of *Erwinia carotovora* Pectate Lyase B, His tag, Histidine tag for metal ion chromatography.

Fig. 27 demonstrates the chromogenic FVIII-like activity of two antibody 198/B1 (subclone AB2) scFv fragment-alkaline phosphatase fusion proteins (198AB2#1 and 198AB2#100) in the presence of 2.3nM human FIXa. As a positive control 7.5pM FVIII was used.

Fig. 28 shows the amino acid and nucleotide sequence of pZip198AB2#102 (SEQ.ID.NOs. 91 and 92).

Fig. 29 shows the nucleotide and amino acid sequence of the mAB#8860 scFv-alkaline phosphatase fusion protein (vector pDAP2-8860scFv#11, (SEQ.ID.NOs. 93 and 94). The genes for the VL and the VH domains of antibody #8860 were derived from the corresponding hybridoma cells as described in example 10. The PCR product of the VH-gene was digested SfiI - AscI and the PCR-product of the VL-gene was digested AscI and NotI. VH and VL genes were linked via the AscI site and inserted into SfiI - NotI digested vector pDAP2 (Kerschbaumer R.J. et al, Immunotechnology 2, 145-150, 1996; GeneBank accession No.:U35316).

Fig. 30 shows the nucleotide and amino acid sequence of the mAB #8860 scFv-leucine zipper fusion

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protein (miniantibody; vector p8860-Zip#1.2,
(SEQ.ID.NOs. 95 and 96). The gene of the scFv fragment
was derived from mAB #8860 and was swapped from vector
pDAP2-8860scFv#11 into SfiI-NotI digested plasmid pZip1
5 (Kerschbaumer R.J. et al., Analytical Biochemistry 249,
219-227, 1997; GeneBank accession No.: U94951)

Fig. 31 demonstrates the chromogenic FVIII-like
activity of the 198/B1 (subclone AB2) miniantibody
198AB-Zip#102 in the presence of 2.3nM human FIXa. As a
10 positive control 4.8pM FVIII was used whereas a
unrelated miniantibody (8860-Zip#1.2) and plain reaction
buffer (IZ) served as negative controls.

Fig. 32 shows a schematic representation of the
plasmid pMycHis6.

15 Fig. 33 shows the nucleotide and amino acid
sequence of the part of the plasmid pMycHis6 differing
from vector pCOCK (SEQ.ID.NOs. 97 and 98). Vector
pMycHis6 was constructed by cleaving vector pCOCK
(Engelhardt et al., 1994, Biotechniques, 17:44-46) with
20 NotI and EcoRI and insertion of the oligonucleotides:
mychis6-co: 5'ggccgcagaacaaaaactcatctcagaagaggatct
gaatggggcggcacatcaccatcaccatcactaataag 3' (SEQ ID.No.
79) and mycchis-ic:
5'aattcttattagtgatggtgatggtgatgtgccgccccattcagatcctcttct
25 gagatgagtttttgttctgc (SEQ.ID.No. 80).

Fig. 34 shows the nucleotide and amino acid
sequence of 198AB2 scFv (linked to the c-myc-tag and the
His6- tag): ORF of the expression vector pMycHis6-
198AB2#102. Vector pMycHis6 was constructed by cleaving
30 vector pCOCK (Engelhardt O. et al, BioTechniques 17, 44-
46, 1994) NotI - EcoRI and inserting the following
annealed oligonucleotides:

(5'-GGCCGCAGAACAACAACTCATCTCAGAAGAGGATCTGAATGGG
GCGGCACATCACCATCACCATCACTAATAAG - 3' (SEQ.ID.No. 103)
35 and
5'- TTATTAGTGATGGTGATGGT
GATGTGCCGCCCCATTGAGATCCTCTTCTGAGATGAGTTTTTGTCTGC-

3' (SEQ.ID.NO. 104)). The resultant vector, named pMyHis6, was cleaved SfiI - NotI and the gene of scFv 198AB2 was swapped into this vector from vector pDAP2-198AB2#100.

5 Fig. 35 shows the nucleotide and amino acid sequence of the mAB #8860 scFv linked to the c-myc-tag and the His6- tag (vector p8860-M/H#4c, SEQ.ID.NOs. 101 and 102). Plasmid pMyHis6 was cleaved with SfiI and NotI and the DNA sequence coding for the scFv 8860#11
10 protein was inserted from pDAP2-8860scFv#11 (see Fig.29) yielding plasmid p8860-M/H#4c.

Fig. 36 demonstrates the chromogenic FVIII-like activity of the 198/B1 (subclone AB2) scFv fragment (MyHis-198AB2#102) in the presence of 2.3nM human FIXa.
15 As a positive control 4.8pM FVIII was used whereas a unrelated scFv (8860-M/H#4c) and plain reaction buffer (IZ) served as negative controls.

Antibodies and Antibody Derivatives

20 The present invention also comprises the nucleic acids encoding the inventive antibodies and antibody derivatives, expression vectors, hybridoma cell lines, and methods for producing the same.

Antibodies are immunoglobulin molecules having a specific amino acid sequence which only bind to antigens
25 that induce their synthesis (or its immunogen, respectively) or to antigens (or immunogens) which are very similar to the former. Each immunoglobulin molecule consists of two types of polypeptide chains. Each molecule consists of large, identical heavy chains (H
30 chains) and two light, also identical chains (L chains). The polypeptides are connected by disulfide bridges and non-covalent bonds. In vivo, the heavy and light chains are formed on different ribosomes, assembled in the cell, and secreted as intact immunoglobulins (Roitt I.
35 et al., in: Immunology, second ed., 1989).

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The inventive antibodies and antibody derivatives and organic compounds derived there from comprise human and animal monoclonal antibodies or fragments thereof, single chain antibodies and fragments thereof and
5 miniantibodies, bispecific antibodies, diabodies, triabodies, or di-, oligo- or multimers thereof. Also included are peptidomimetics or peptides derived from the antibodies according to the invention, e.g. they
10 comprise one or several CDR regions, preferably the CDR3 region.

Further included are human monoclonal antibodies and peptide sequences which, based on a structure activity connection, are produced through an artificial modeling process (Greer J. et al., J. Med. Chem., 1994, Vol. 37, pp. 1035-1054).
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The term factor IX/IXa activating antibodies and antibody derivatives may also include proteins produced by expression of an altered, immunoglobulin-encoding region in a host cell, e.g. "technically modified
20 antibodies" such as synthetic antibodies, chimeric or humanized antibodies, or mixtures thereof, or antibody fragments which partially or completely lack the constant region, e.g. Fv, Fab, Fab' or F(ab)'₂ etc. In these technically modified antibodies, e.g., a part or
25 parts of the light and/or heavy chain may be substituted. Such molecules may, e.g., comprise antibodies consisting of a humanized heavy chain and an unmodified light chain (or chimeric light chain), or vice versa. The terms Fv, Fc, Fd, Fab, Fab' or F(ab)₂
30 are used as described in the prior art (Harlow E. and Lane D., in "Antibodies, A Laboratory Manual", Cold Spring Harbor Laboratory, 1988).

The present invention also comprises the use of Fab fragments or F(ab)₂ fragments which are derived from
35 monoclonal antibodies (mAb), which are directed against factor IX/factor IXa and cause an increase of the procoagulant activity of factor IXa.

Preferably, the heterologous framework regions and constant regions are selected from the human immunoglobulin classes and isotypes, such as IgG (subtypes 1 to 4), IgM, IgA and IgE. In the course of the immune response, a class switch of the immunoglobulins may occur, e.g. a switch from IgM to IgG; therein, the constant regions are exchanged, e.g. from μ to γ . A class switch may also be caused in a directed manner by means of genetic engineering methods ("directed class switch recombination"), as is known from the prior art (Esser C. and Radbruch A., Annu. Rev. Immunol., 1990, Vol. 8, pp. 717-735). However, the antibodies and antibody derivatives according to the present invention need not comprise exclusively human sequences of the immunoglobulin proteins.

In one particular embodiment, a humanized antibody comprises complement determining regions (CDRs) from murine monoclonal antibodies which are inserted in the framework regions of selected human antibody sequences. However, human CDR regions can also be used. Preferably, the variable regions in the human light and heavy chains are technically altered by one or more CDR exchanges. It is also possible to use all six CDRs or varying combinations of less than six CDRs.

The humanized antibody according to the present invention preferably has the structure of a human antibody or of a fragment thereof and comprises the combination of characteristics necessary for a therapeutic application, e.g., the treatment of coagulation disorders in patients, preferably factor VIII inhibitor patients.

A chimeric antibody differs from a humanized antibody in that it comprises the entire variable regions including the framework regions of the heavy and light chains of non-human origin in combination with the constant regions of both chains from human immuno-

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globulin. A chimeric antibody consisting of murine and human sequences may, for example, be produced.

According to the present invention, the antibodies and antibody derivatives may also be single chain antibodies or miniantibodies (scFv fragments, which, e.g., are linked to proline-rich sequences and oligomerisation domains, e.g. Pluckthun A. and Pack P., *Immuno-technology*, 1997, Vol. 3, pp. 83-105) or single chain Fv (sFv) which incorporate the entire antibody binding region in one single polypeptide chain. For instance, single chain antibodies may be formed by linking the V-genes to an oligonucleotide which has been constructed as a linker sequence and connects the C terminus of the first V region with the N terminus of the second V region, e.g. in the arrangement VH-Linker-VL or VL-Linker-V_H; both, V_H and V_L thus may represent the N-terminal domain (Huston JS et al., *Int. Rev. Immunol.*, 1993, Vol. 10, pp. 195-217; Raag R. and Whitlow M., *FASEB J.*, 1995, Vol. 9, pp. 73-80). The protein which can be used as linker sequence may, e.g., have a length of up to 150 Å, preferably up to 80 Å, and more preferably up to 40 Å. Linker sequences containing glycine and serine are particularly preferred for their flexibility, or glutamine and lysine, respectively, for their solubility. The choice of the amino acid is effected according to the criteria of immunogenicity and stability, also depending on whether or not these single chain antibodies are to be suitable for physiological or industrial applications (e.g. immunoaffinity chromatography). The single chain antibodies may also be present as aggregates, e.g. as trimers, oligomers or multimers. The linker sequence may, however, also be missing, and the connection of the V_H and V_L chains may occur directly.

Bispecific antibodies are macromolecular, heterobifunctional cross-linkers having two different binding specificities within one single molecule. In

this group belong, e.g., bispecific (bs) IgGs, bs IgM-IgAs, bs IgA-dimers, bs (Fab')₂, bs(scFv)₂, diabodies, and bs bis Fab Fc (Cao Y. and Suresh M.R., Bioconjugate Chem., 1998, Vol. 9, pp. 635-644).

5 By peptidomimetics, protein components of low molecular weight are understood which imitate the structure of a natural peptide component, or of templates which induce a specific structure formation in an adjacent peptide sequence (Kemp DS, Trends
10 Biotechnol., 1990, pp. 249-255). The peptidomimetics may, e.g., be derived from the CDR3 domains. Methodical mutational analysis of a given peptide sequence, i.e. by alanine or glutamic acid scanning mutational analysis, allows for the identification of peptide residues
15 critical for procoagulant activity. Another possibility to improve the activity of a certain peptide sequence is the use of peptide libraries combined with high throughput screening.

The term antibodies and antibody derivatives may
20 also comprise agents which have been obtained by analysis of data relating to structure-activity relationships. These compounds may also be used as peptidomimetics (Grassy G. et al., Nature Biotechnol., 1998, Vol. 16, pp. 748-752; Greer J. et al., J. Med.
25 Chem., 1994, Vol. 37, pp. 1035-1054).

Examples of hybridoma cells expressing the antibodies or antibody derivatives according to the invention were deposited on 9 September 1999 under the numbers 99090924 (#198/A1), 99090925 (#198/B1) and
30 99090926 (#198/BB1) and on December 16, 1999 under the numbers 99121614 (#193/A0), 99121615 (#196/C4), 99121616 (#198/D1), 99121617 (198/T2), 99121618 (#198/G2), 99121619 (#198/AC1) and 99121620 (#198/U2) according to the Budapest Treaty.

35 **Methods of Production:**

The antibodies of the present invention can be prepared by methods known from the prior art, e.g. by

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321, pp.522-525; Phage Display of Peptides and Proteins, A Laboratory Manual, 1996, Eds. Kay et al., pp. 127-139; US 4,873,316; Vaughan T.J. et al., Nature Biotechnology, 1998, pp. 535-539; Persic L. et al., Gene, 1997, pp. 9-18; Ames R.S. et al., J.Immunol.Methods, 1995, pp. 177-186).

The expression of recombinantly produced antibodies may be effected by means of conventional expression vectors, such as bacterial vectors, such as pBr322 and its derivatives, pSKF or eukaryotic vectors, such as pMSG and SV40 vectors. Those sequences which encode the antibody may be provided with regulatory sequences which regulate the replication, expression and secretion from the host cell. These regulatory sequences comprise promoters, e.g. CMV or SV40, and signal sequences.

The expression vectors may also comprise selection and amplification markers, such as the dihydrofolate reductase gene (DHFR), hygromycin-B- phosphotransferase, thymidine-kinase etc.

The components of the vectors used, such as selection markers, replicons, enhancers etc., may either be commercially obtained or prepared by means of conventional methods. The vectors may be constructed for the expression in various cell cultures, e.g. for mammalian cells such as CHO, COS, fibroblasts, insect cells, yeast or bacteria, such as E. coli. Preferably, those cells are used which allow for an optimal glycosylation of the expressed protein. Particularly preferred is the vector pBax (cf. Fig. 17) which is expressed in CHO cells or in SK-Hep.

The production of Fab fragments or F(ab)₂ fragments may be effected according to methods known from the prior art, e.g. by cleaving a mAb with proteolytic enzymes, such as papain and/or pepsin, or by recombinant methods. These Fab and F(ab)₂ fragments may also be prepared by means of a phage display gene library (Winter et al., 1994, Ann. Rev. Immunol., 12:433-455).

The antibody derivatives may also be prepared by means of methods known from the prior art, e.g. by molecular modeling, e.g. from Grassy G. et al., Nature Biotechnol., 1998, Vol. 16, pp. 748-752, or Greer J. et al., J. Med. Chem., Vol. 37, pp. 1035-1054, or Rees A. et al., in: "Protein Structure Prediction: A practical approach", ed. Sternberg M.J.E., IRL press, 1996, chapt. 7-10, pp. 141-261.

The purification of the inventive antibodies and antibody derivatives may also be carried out by methods described in the prior art, e.g., by ammonium sulfate precipitation, affinity purification (protein G-Sepharose), ion exchange chromatography, or gel chromatography. The following methods may be used as the test methods to show that the antibodies and antibody derivatives of the present invention bind to factor IX/factor IXa, increase the procoagulant activity of factor IXa or have factor VIII-like activity.: the one step coagulation test (Mikaelsson and Oswaldson, Scand. J. Haematol., Suppl., 33, pp. 79-86, 1984) or the chromogenic tests, such as COATEST VIII:C® (Chromogenix) or Immunochem (IMMUNO). In principle, all the methods used for determining factor VIII activity may be used. As the control blank value for the measurements, e.g., unspecific mouse-IgG antibody may be used.

The present antibodies and antibody derivatives are suitable for therapeutic use in the treatment of coagulation disorders, e.g. in the case of hemophilia A, for factor VIII inhibitor patients etc. Administration may be effected by any method suitable to effectively administer the therapeutic agent to the patient, e.g. by oral, subcutaneous, intramuscular, intravenous or intranasal administration.

Therapeutic agents according to the invention may be produced as preparations which comprise a sufficient amount of antibodies or of antibody derivatives as the active agent in a pharmaceutically acceptable carrier

substance. These agents may be present either in liquid or in powderized form. Moreover, the preparations according to the invention may also comprise mixtures of different antibodies, the derivatives thereof and/or organic compounds derived therefrom, as well as mixtures consisting of antibodies and factor IX and/or factor IXa. Factor IXa may be present as factor IXa α and/or factor IXa β . An example of an aqueous carrier substance is, e.g., saline. The solutions are sterile, sterilisation being effected by conventional methods.

The antibodies or antibody derivatives according to the invention may be present in lyophilized form for storage and be suspended in a suitable solvent before administration. This method has proven generally advantageous for conventional immunoglobulins, and known lyophilisation and reconstitution methods may be applied in this case.

Moreover, the antibodies and antibody derivatives according to the invention may also be used for industrial applications, e.g. for the purification of factor IX/factor IXa by means of affinity chromatography, or as a component of detection methods (e.g. ELISA assays), or as an agent for identification of and interaction with functional domains of a target protein. The present invention will be described in more detail by way of the following examples and drawing figures, to which, however, it shall not be restricted.

Examples

Example 1: Immunization of immunocompetent mice and generation of anti-FIX/IXa antibody secreting hybridoma cells

Groups of 1-3 normal immunocompetent 5-8 week old Balb/c mice were immunized with 100 μ g antigen (100 μ l doses) via the intraperitoneal (i.p.) route. In a typical experiment, mice were inoculated with either recombinant human coagulation factor (F) IX (BenefixTM),

human activated FIX α (Enzyme Research Laboratories, Lot: FIX α 1190L) or human FIX $\alpha\beta$ (Enzyme Research Laboratories, Lot: HFIX $\alpha\beta$ 1332 AL,) adjuvanted with Al(OH)₃ or KFA.

- 5 Individual mice were boosted at various times with 100 μ g antigen (100 μ l doses, i.p) and sacrificed two days later. Spleen cells were removed and fused to P3 X63-Ag8 6.5.3 myeloma cells essentially as described by Lane et al., 1985 (J. Immunol. Methods, Vol. 81, pp. 223-228).
 10 Each fusion experiment was individually numbered, i.e. #193, 195, 196 or 198.

- Hybridoma cells were grown in 96 well plates on a macrophage feeder layer (app. 10⁵ cells/ml) and selected in HAT-medium (RPMI-1640 medium supplemented with
 15 antibiotics, 10% FCS, Na-pyruvate, L-glutamine, 2-mercaptoethanol and HAT (HAT 100x: 1.0x10⁻²M hypoxanthine in H₂O (136.1 mg/100ml H₂O), 4.0x10⁻⁵M aminopterin in H₂O (1.76 mg/100ml H₂O) and 1.6x10⁻³M thymidine in H₂O (38.7 mg/100ml H₂O). Medium was first
 20 changed after 6 days and thereafter twice a week. After 2-3 weeks HAT-medium was changed to HT-medium (RPMI-1640 supplemented with antibiotics, 10%FCS, Na-pyruvate, L-glutamine, 2-mercaptoethanol and HT) and later on (after additional 1-2 weeks) to normal growth medium (RPMI-1640
 25 medium supplemented with 10%FCS, Na-pyruvate, L-glutamine and 2-mercaptoethanol) (see: HYBRIDOMA TECHNIQUES, EMBO, SKMB Course 1980, Basel).

- In another set of experiments FIX deficient C57B16 mice (Lin et al., 1997, Blood, 90:3962) were used for
 30 immunization and subsequent hybridoma production. Since FIX knockout (k.o.) mice do not express endogenous FIX, the anti (a)-FIX antibody spectrum achievable is supposed to be different compared to normal Balb/c mice (due to lack of tolerance).

Example 2: Assaying for FVIII-like activity in supernatants of anti-FIX/FIXa antibody secreting hybridoma cells

In order to assay the FVIII-like activity of anti-FIXa antibodies secreted by hybridoma cells, the commercially available test-kit COATEST VIII:C/4[®] (Chromogenix) was employed. The assay was done essentially as described by the manufacturer with the following modifications:

To allow high throughput screening, the assay was downscaled to microtiter plate format. Briefly, 25µl aliquots of hybridoma supernatants were transferred to microtiter plate (Costar, #3598) wells and warmed to 37°C. Chromogenic substrate (S-2222), synthetic thrombin inhibitor (I-2581), factor (F) IXa and FX were reconstituted in sterile water and FIXa/FX was mixed with phospholipids according to the supplier's protocol. Per reaction, 50µl of the phospholipid/FIXa/FX solution were combined with 25µl CaCl₂ (25mM) and 50µl of the substrate/inhibitor cocktail. To start the reaction, 125µl of the premix were added to the hybridoma supernatant in the microtiter plates and incubated at 37°C. Absorbency at 405nm and 490nm of the samples was read at various times (30min to 12h) against a reagent blank (MLW, cell culture medium instead of hybridoma supernatant) in a Labsystems iEMS Reader MF[™] microtiter plate reader. FVIII-like activity of the samples was calculated by comparing the absorbency of the samples against the absorbency of a diluted FVIII reference standard (IMMUNO AG # 5T4AR00) using GENESIS[™] software.

The results of a screening for FVIII-like activity in hybridoma cell culture supernatants are shown in Fig. 1. Pre-selected clones derived from fusion experiments #193, #195 and #196 (see above) were examined in a chromogenic FVIII assay as described. Clones 193/M1, 193/N1 and 193/P1 are subclones derived from the master

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clone 193/C0 (see below). Master clone 195/10 was derived from fusion experiment #195 and clones 196/A0, 196/B0 and 196/C0 were derived from fusion experiment #196. In a typical screening experiment, approximately 1000 clones (in 96 wells) from a single fusion experiment were pre-screened for FVIII-like activity. Subsequently, selected clones were grown on a larger scale (3-5 ml supernatant) and re-analyzed in a chromogenic assay. As a negative control cell culture medium was assayed on each plate (MLW).

Wells either exhibiting high FVIII-like activity or substantial FVIII-like activity were subjected to subcloning procedures. The selection and subcloning process is exemplified for the screening and subcloning of an IgG producing cell line (i.e. 193/C0) but has been done exactly the same way for an IgM (i.e. 196/C0, see below, Fig. 5) producing clone.

The selection process was done by initially plating all hybridoma cell clones derived from a single fusion experiment on ten 96 well plates thereby creating the so called "master plates". Singular positions (wells) on a master plate usually contained more than one hybridoma cell clone (usually 3 to 15 different clones). Subsequently, the antibody secreted by only several thousand cells was tested. These cells grew under conditions suboptimal for antibody production, which is known to be best in dying cells. So the expected specific anti-FIX antibody concentration in the supernatant may be in the range of 10^{-12} to 10^{-14} M. This explains why incubation periods had to be extended compared to standard FVIII assays.

Results of a screening for an IgG mediated FVIII-like activity in hybridoma cell culture supernatants of a master plate are shown in Fig. 2. Supernatants were examined in a chromogenic FVIII assay. Shown are the results derived from the fifth master plate of fusion experiment number #193 (Balb/c mice immunized with

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FIX α). Absorbance was read after 4 hours of incubation at 37°C. Position ES was identified as exhibiting FVIII like activity significantly higher than the blank (MLW). This cell pool was designated 193/C0 and was further subcloned (Figure 3). As each well of the master plate contains more than one hybridoma cell clone, cells of a single positive well were expanded and plated at a calculated cell density of 2 - 0.2 cells/well on a 96 well plate. Again, the supernatants were tested for FVIII-like activity and positive positions were subjected to another round of subcloning. Typically three to four rounds of subcloning were performed with each clone displaying FVIII-like activity to obtain homogenous cell populations. Here the results of the chromogenic assay of the 193/C0 subclones are shown. Absorbance was read after a 4 hour incubation period at 37°C. Positions A6 and D5 exhibited substantial FVIII-like activity and were named 193/M1 and 193/P1, respectively. These two clones were subjected to another round of subcloning. As a negative control plain cell culture medium was assayed on each plate (MLW(H1)).

A comparison of chromogenic FVIII-like activity and FIX-ELISA reactivity of small scale (3 ml) hybridoma cultures is shown in Fig. 4. Before a decision was made whether a master clone (or subclone) was to be further subcloned, clones were grown at a 3-5 ml scale and the supernatants were checked again. This graph shows the FIX specific ELISA results and the FVIII-like chromogenic activity of the master clone 193/C0 and all its subclones which were identified as positives and rechecked. Blanks (absorbency of the chromogenic reagent itself) were subtracted in the case of the ELISA as well as the chromogenic assay readings depicted here. Clone 193/M1 was subcloned and yielded clones 193/V2, 193/M2 and 193/U2. The other clones of the 2nd round came from 193/P1, 193/AB2 and 193/P2 were subcloned. 193/AF3, 193/AB3 and 193/AE3 are subclones of 193/AB2. The other

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clones of the 3rd round came from 193/P2. Finally 193/AF3 (→193/AF4), AE3 (→193/AE4, 193/AL4, 193/AN4 and 193/AO4) and 193/AD3 (→193/AG4, 193/AH4, 193/AD4, 193/AI4, 193/AK4) were subcloned.

- 5 From each fusion experiment, several (5-15) master clones (selected from the master plate) were identified and subjected to subcloning. After 3 rounds of sub-cloning, most of the cell lines were homogenous as demonstrated by ELISA and chromogenic activity analysis
10 (see Fig. 4) as well as by cDNA sequence analysis. A specific master clone and all its subclones produce the same FIX/FIXa binding antibody. However, there are huge differences in the antibody protein sequences of clones derived from different master clones (see Example 11).
15 Most hybridoma cell lines express antibodies from the IgG subclass (i.e. clones #193, #198, like 198/A1, 198/B1, 198/BB1). However, we were also able to select some clones expressing IgM antibodies.

- The chromogenic activity of hybridoma supernatant
20 of some important master clones and subclones was determined. Absorbance was measured after a 1h 30 min and 3h 30 min incubation period at 37°C (Fig. 5). In contrast to all the clones from the 193rd fusion, clone 196/C0 and its subclone 196/AP2 produced a FIX/FIXa-
25 specific IgM antibody that gave a strong chromogenic activity even after a short period of incubation.

- The following cell lines have been deposited with the European Collection of Cell Cultures (ECACC) in accordance with the Budapest Treaty: 98/B1 (ECACC No.
30 99090925); 198/A1 (ECACC No. 99090924); 198/BB1 (ECACC No. 99090926); 193/AO (ECACC No. 99121614); 196/C4 (ECACC No. 99121615); 198/D1 (ECACC No. 99121616); 198/T2 (ECACC No. 99121617); 198/G2 (ECACC No. 99121618); 198/AC1 (ECACC No. 99121619); and 198/U2
35 (ECACC No. 99121620).

To do a more in depth analysis of the biochemical properties of certain antibodies, homogenous hybridoma

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Example 3: Factor IX/FIXa_(α,β) binding properties of antibodies exhibiting FIX/FIXa activating activity

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Purification of mouse-IgG from hybridoma cell culture supernatants

Hybridoma supernatant (100-500 ml) was supplemented with 200 mM Tris/HCl buffer (pH 7.0) and solid NaCl to

give final concentrations of 20 mM Tris and 3M NaCl, respectively. The supernatant was then clarified by centrifugation at 5500 x g for 10 minutes. A 1 ml protein G affinity chromatography column (Protein G Sepharose Fast Flow, Amersham-Pharmacia) was washed with 15 ml 20 mM Tris/Cl pH 7.0 and afterwards equilibrated with 10 ml of 20 mM Tris/Cl buffer pH 7.0 containing 3M NaCl. The hybridoma supernatant containing 3M NaCl was then loaded onto the column by gravity. The column was washed with 15 ml of 20 mM Tris/Cl buffer, pH 7.0, containing 3M NaCl. Bound IgG was further eluted with 12 ml glycine/HCl buffer pH 2.8 and 1 ml fractions were collected. 100µl of 1M Tris pH 9.0 were added to each fraction for neutralization. Fractions containing the IgG were identified by mixing 50µl with 150µl of a staining solution (BioRad concentrate, 1:5 diluted with water) in wells of a microplate. Positive fractions were pooled, concentrated to 1 ml in an ultrafiltration concentrator device (Centricon Plus 20, Amicon) according to the manufacturer. The concentrate was diluted with 19 ml TBS (20 mM Tris/Cl buffer pH 7.0 containing 150mM NaCl) and again concentrated to 1 ml. The diluting-concentrating step was repeated for two more times in order to bring IgG into TBS.

Purification of mouse-IgM from hybridoma cell supernatants

100-500 ml of hybridoma cell culture supernatant were concentrated to 5-10 ml either with an ultra-filtration concentrator device (Centricon Plus 20, Amicon) according to the manufacturer or by ammonium sulfate precipitation (40% saturation, 0°C) and redissolving the precipitate with 5-10 ml of TBS. In either case the concentrate was dialyzed against 20mM Tris Cl buffer pH 7.4 containing 1.25M NaCl and further concentrated to 1 ml in a Centricon Plus 20, (Amicon) ultrafiltration device. IgM was purified from this

concentrate with the ImmunoPure IgM Purification Kit (Pierce) according to the manufacturer. Fractions collected during elution from the maltose binding protein-column were tested for IgM, pooled, concentrated and brought into TBS as described for IgG.

Determination of IgG concentrations in purified preparations

Total IgG content 280nm - extinction of appropriate dilutions were measured. E280 = 1.4 corresponds to 1 mg/ml protein.

Factor IXa specific IgG (quantitative ELISA)

Wells of a microplate (Nunc Maxisorp) were incubated with 2µg/ml factor IXa diluted in TBS (25mM Tris/HCl pH 7.5 containing 150mM NaCl) overnight at 4°C. Wells were washed four times with TBST (25mM Tris/HCl pH 7.5 containing 150mM NaCl and 0.1% (v/v) Tween 20). As a standard monoclonal AB the HIX1 anti-FIX (accurate) was used. Standard and samples were diluted in TBST containing 2%(w/v) BSA. The standard dilution series and appropriate dilutions of the samples were incubated on the ELISA-plate for 2 hours at room temperature. Plates were washed 4 times with TBST and incubated (2h, RT) with 100µl/well of a 1:25000 dilution (in TBST/1%BSA) of an anti-mouse IgG (Fc-specific) peroxidase conjugated antibody (Sigma, #A-0168) FIXa. Wells were washed 5 times with TBST and finally stained with 100µl freshly prepared staining solution (10ml 50mM sodium citrate, pH 5 supplemented with 100µl OPD (60mg OPD/ml) and 10µl 30% H₂O₂). The reaction was stopped by the addition of 50ml H₂SO₄ and after 30 minutes the optical density was recorded at 492nm and 620nm in a Labsystems iEMS Reader MFTM microtiter plate reader employing GENESIS™ software.

Example 4: Anti-FIX/FIXa antibodies exhibiting FVIII-like activity in a chromogenic FVIII assay

Several anti-FIX/FIXa antibody producing hybridoma clones were subcloned up to four times and the resulting monoclonal hybridoma cell line used to produce monoclonal antibody containing supernatant. IgG isotype antibodies derived from these supernatants were purified over affinity columns and dialyzed against TBS (see above). IgM antibodies were used as unpurified supernatant fractions. The following experiments were done with two sets of representative antibodies: 193/AD3 and 198/AC1/1 (IgG isotype, the antibody 198/AC1/1 is a preparation from the parent 198/AC1 hybridoma clone, i.e. that a (frozen) vial containing 198/AC1 cells is cultivated and antibodies are produced. The supernatant is then used for these experiments.) and 196/AF2 and 196/AF1 (IgM isotype) (Fig. 6A and Fig. 6B). Briefly, 25µl aliquots of monoclonal antibody containing sample (unpurified hybridoma supernatant or, where indicated, a certain amount of FIX specific antibody) were transferred to microtiter plate wells and warmed to 37°C. Chromogenic substrate (S-2222), synthetic thrombin inhibitor (I-2581), factor (F) IXa and FX were reconstituted in sterile water and FIXa/FX was mixed with phospholipids according to the supplier's protocol. Per reaction, 50µl of the phospholipid/FIXa/FX solution were combined with 25µl CaCl₂ (25mM) and 50µl of the substrate/inhibitor cocktail. To start the reaction, 125µl of the premix were added to the monoclonal antibody solution in the microtiter plates and incubated at 37°C. Absorbance at 405nm and 490nm of the samples was read at various times (5min to 6h) against a reagent blank (cell culture medium instead of hybridoma supernatant) in a Labsystems iEMS Reader MF™ microtiter plate reader using GENESIS™ software.

The time course of FVIII-like activity exhibited by monoclonal antibodies 193/AD3 (IgG isotype) and 196/AF2 (IgM isotype) compared to human FVIII (12 and 16mU/ml),

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TBS and to cell culture medium is shown in Fig. 6A. After a lag phase, both antibodies give rise to chromogenic substrate cleavage, as judged by the increasing optical density measurable at 405nm wavelength.

The time course of FVIII-like activity exhibited by monoclonal antibodies 198/AC1/1 (IgG isotype, 10µg/ml) and 196/AF1 (IgM isotype, unpurified supernatant) compared to human FVIII (16mU/ml) and 10µg/ml of mouse IgG is shown in Fig. 6B. After a lag phase, both antibodies give rise to chromogenic substrate cleavage, as judged by the increasing optical density measurable at 405nm wavelength.

Example 5: FVIII-like activity exhibited by anti-FIX/FIXa-antibodies generates factor Xa and is phospholipid, FIXa/FX and Ca²⁺ dependent.

Factor VIII activity is usually determined with a chromogenic assay and/or an APTT-based clotting assay. Both types of assays rely on FVIIIa/FIXa-mediated factor Xa generation. In the case of a chromogenic FVIII assay, the factor Xa produced will subsequently react with a chromogenic substrate, which can be monitored spectroscopically, e.g., in an ELISA reader. In an APTT based clotting assay free factor Xa will assemble with FVa on a phospholipid surface in the so-called prothrombinase complex and activate prothrombin to thrombin. Thrombin in turn gives rise to fibrin generation and finally to clot formation. Central to the two assay systems is generation of factor Xa by the FVIIIa/FIXa complex.

To demonstrate that the FVIII-like activity exhibited by anti-FIX/FIXa-antibodies indeed generates factor Xa, the following experiment was carried out. Several 25µl aliquots of unpurified hybridoma supernatant 196/AF2 (IgM isotype) were transferred to microtiter plate wells and warmed to 37°C. As a positive

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control, 16mU of Recombinate™ were diluted into hybridoma medium (196 HM 007/99) and treated exactly the same way as the hybridoma supernatant. As a negative control, plain hybridoma medium was used. Chromogenic substrate (S-2222), synthetic thrombin inhibitor (I-2581), factor IXa and FX were reconstituted in sterile water and FIXa/FX was mixed with phospholipids according to the supplier's protocol. Pefabloc Xa®, a factor Xa specific proteinase inhibitor (Pentapharm, LTD), was reconstituted with water to a final concentration of 1mM/l. Per reaction, 50µl of the phospholipid/FIXa/FX solution were combined with 25µl CaCl₂ (25mM) and 50µl of the substrate/thrombin-inhibitor cocktail. To start the reaction, 125µl of the premix were added to the samples in the microtiter plates and incubated at 37°C. Where indicated, 35µM Pefabloc Xa® were added. Absorbance at 405nm and 490nm was read at various times (every 5 minutes to 6h) against a reagent blank (cell culture medium) in a Labsystems iEMS Reader MF™ microtiter plate reader employing the GENESIS™ software.

The results of the factor IXa stimulation by the FVIII-like activity exhibited by the IgM anti- FIX/FIXa-antibody 196/AF2 in generating factor Xa as judged by the readily measurable cleavage of the chromogenic substrate S-2222 (compare "16mU FVIII" and "196/AF2") is shown in Fig. 7A. Factor Xa activity is effectively blocked by the FXa specific inhibitor "Pefabloc Xa®" (compare "196/AF2" versus "196/AF2 35µM Pefabloc Xa®") indicating that indeed FXa was generated.

The same experiment was performed using purified IgG preparations of clone 198/AM1 (Fig. 7B). Purified IgG was diluted in TBS to a final concentration of 0,4mg/ml and 25µl (i.e. a total of 10µg), transferred to microtiter plate wells and warmed to 37°C. As a positive

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control, 6mU plasma- derived FVIII was used. 10µg
 unspecific mouse IgG (Sigma, I-5381) served as a
 negative control. The assay was performed as described
 above.

- 5 Further experiments show the factor IXa stimulation
 by the FVIII-like activity exhibited by the IgG anti-
 FIX/FIXa-antibody 198/AM1 generates factor Xa as judged
 by the readily measurable cleavage of the chromogenic
 substrate S-2222 (Fig. 7B). Again factor VIII and
 10 antibody 198/AM1 generate FXa which is effectively
 blocked by the FXa specific inhibitor "Pefabloc Xa®". As
 a negative control, unspecific mouse IgG (Sigma, I5381)
 was assayed.

- 15 In another set of experiments, the dependence of
 the FVIII-like activity of either purified anti-
 FIX/FIXa-antibodies (IgM, Fig.8A) or of unpurified
 antibodies derived from cell culture supernatants (IgG,
 Fig. 8B) on the presence of phospholipids (PL), FIXa/FX
 and Ca^{2+} was demonstrated. Mouse IgG was used as a
 20 control (Fig. 8C). Factor VIII-like activity was assayed
 essentially as described above. When indicated, either
 the FIXa/FX mixture, the PL or Ca^{2+} was omitted from the
 reaction. Absorbency at 405nm and 490nm of the samples
 was read at various times against a reagent blank
 25 (buffer instead of purified antibody) in a Labsystems
 iEMS Reader MF™ microtiter plate reader. The results
 are shown in Fig. 8A, Fig. 8B and Fig. 8C.

- The dependence of the FVIII-like activity of
 purified anti-FIXa-antibody 198/AC1/1 (IgG isotype,
 30 concentration used throughout the assay was 10µg/ml) on
 the presence of phospholipids (PL), FIXa/FX and Ca^{2+} is
 further shown in Fig. 8A. As is easily recognizable,
 only the complete assay, including antibody, PL, Ca^{2+} ,
 and FIXa/FX gives rise to a reasonable FXa generation.
 35 The dependence of the FVIII-like activity of cell
 culture supernatant containing unpurified IgM isotype

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anti-FIX/FIXa-antibody (196/AF1) on the presence of phospholipids, FIXa/FX and Ca^{2+} is shown in Fig. 8B.

Again, as already shown for the purified IgG preparation (Fig. 8A), antibody 198/AC1/1, only the
 5 complete assay, including PL, Ca^{2+} , FIXa/FX, will give a reasonable amount of FXa generation. To demonstrate the specificity of the reaction, total IgG prepared from normal mouse plasma was assayed under the same conditions as above. The results are shown in Fig. 8C.
 10 No FVIII-like activity could be detected. There is, as expected, no activity detectable in the absence of phospholipids, FIXa/FX and Ca^{2+} . All experiments were done in a microtiter plate and the OD405 was scanned every 5 minutes for 6h.

15 **Example 6: Certain anti-FIX/FIXa-antibodies are procoagulant in the presence of FIXa**

During normal hemostasis, FIX becomes initially activated either by the tissue factor (TF)/factor VIIa pathway or later on by activated factor XI (FXIa).
 20 Subsequent to its activation, FIXa associates on the platelet surface in a membrane bound complex with activated FVIII. Factor IXa by itself has little or no enzymatic activity towards FX, but becomes highly active in the presence of FVIIIa. To demonstrate that certain
 25 anti-FIX/FIXa antibodies have FVIII-like activity and hence are procoagulant in a FVIII deficient human plasma, the following experiment was carried out. Different amounts of antibody 193/AD3 or mouse IgG (as a control) were used in a standard aPTT based one stage
 30 clotting assay. Briefly, 100 μl of antibody-containing samples were incubated with 100 μl of FVIII deficient plasma (DP) and with 100 μl of DAPTTIN (PTT Reagent for determining activated Thromboplastin Time; IMMUNO AG) reagent, in a KC10A clotting analyzer. Where indicated,
 35 a total amount of 50ng activated FIX was included in the reaction mixture. After a 4 minute incubation, the

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reaction was started by the addition of 100 μ l CaCl₂ (25mM). The results are shown in Table 1 and Fig. 9.

clotting time (sec)			
5	<u>μg AB</u>	<u>193/AD3</u>	<u>mouse IgG</u>
		50ng FIXa	50ngFIXa
10	9	101.6	102.5
	4.5	95.6	103.2
	2.25	93.1	103.2
	1.8	93.7	101.9
	1.35	91.4	103.4
	0.9	94.4	102.2
	0.45	98.1	101.9
	0.34	97.1	103.9
15	0.23	99.3	103.7

Table 1: Clotting times of FVIII deficient plasma in an APTT based clotting assay employing various amounts of procoagulant (193/AD3) and control antibody (mouse IgG) in the presence of 50ng activated FIX (0.01U FIX). The molar ratio of antibody in the reaction and activated FIX is 10:1. The molar ratio between antibody and total FIX (FIX and FIXa, assuming that human FVIII deficient plasma contains 1U (5 μ g) FIX) varies between 6:1 (9 μ g antibody in reaction) and 1:6 (0.23 μ g antibody in reaction). At the optimal shortening of the clotting time, the molar ratio between antibody and total FIX is 1:1. The clotting time without the addition of FIXa is in the range of 120 seconds.

Fig. 9 is a graphical representation of the clotting times of FVIII deficient plasma in an aPTT based clotting assay employing various amounts of procoagulant (193/AD3) and control (mouse IgG) antibody in the presence of 50ng activated FIX. There is a clear dose-dependent reduction of the clotting time in samples supplemented with antibody 193/AD3. These results imply

that antibody 193/AD3 is procoagulant in the presence of FIXa.

Example 7: Anti-FIX/FIXa-antibodies are procoagulant in the presence of FVIII inhibitors and FIXa

A severe complication of the standard FVIII substitution therapy is the development of alloantibodies directed against FVIII, leading to FVIII neutralization and a condition where the patient's blood will not clot.

To demonstrate that certain anti-FIXa-antibodies have FVIII-like activity even in the presence of FVIII inhibitors, the following experiment was carried out. Different amounts of antibody 193/AD3 or, as a control, mouse IgG were used in a standard APTT based one-stage clotting assay. Briefly, 100µl antibody samples were incubated with either 100µl of FVIII deficient plasma (Fig.10A) or FVIII inhibitor plasma (inhibitor potency 400BU/ml), Fig.10B) as well as with 100µl of DAPTTIN reagent, in a KC10A clotting analyzer. In addition, a total amount of 50ng activated FIXa was included in the reaction mixture. After a 4 minute incubation, the reaction was started by the addition of 100µl CaCl₂ (25mM). To ensure equal conditions, the experiments employing FVIII deficient plasma and FVIII inhibitor plasma were done side by side. The results are shown in Fig. 10A and 10B. As already shown in Example 6, there is a clear dose-dependent reduction of the clotting time in samples supplemented with antibody 193/AD3 in the presence of FVIII inhibitors.

Example 8: Anti-FIX/FIXa-antibodies are procoagulant in the presence of defective FVIII and FIXa

To demonstrate that certain anti-FIXa-antibodies have FVIII-like activity in the presence of defective FVIII, the following experiment may be carried out.

Increasing amounts of antibody 193/AD3 or, as a control, mouse IgG are used in a standard aPTT-based one stage clotting assay. In this clotting assay, a hemophilia A patient's plasma having very low clotting activity due to the presence of defective FVIII (DF8) is used. Briefly, 100µl antibody samples are incubated with either 100µl of DF8 plasma or FVIII deficient plasma as well as with 100µl of DAPTTIN reagent, in a KC10A clotting analyzer. In addition, a total amount of 50ng activated FIXa is included in the reaction mixture. After a short incubation, the reaction will be started by the addition of 100µl CaCl₂ (25mM). To ensure equal conditions, the experiment employing FVIII deficient plasma and DF8 plasma is done side by side.

Example 9: Anti-FIX/FIXa-antibodies with procoagulant activity in the presence of FIXa distinguish between human and bovine FIXa

FIX/FIXa specific monoclonal antibodies selected from the 198th fusion experiment were purified from the respective hybridoma supernatant and quantified as described in Example 3. These antibodies were analyzed in a modified one-stage clotting assay (as described in Example 6) and some showed procoagulant activity.

The chromogenic activity of these antibody preparations was measured in the following FXa generation kinetic assay: 10µg of monoclonal antibody (in 25µl) were transferred to microtiter plate wells and warmed to 37°C. Chromogenic substrate (S-2222), synthetic thrombin inhibitor (I-2581), factor IXa and FX were reconstituted in sterile water and FIXa/FX (both bovine) were mixed with phospholipids according to the supplier's protocol. Per reaction, 50µl of the phospholipid/FIXa/FX solution were combined with 25µl CaCl₂ (25mM) and 50µl of the substrate/inhibitor

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cocktail. To start the reaction, 125 μ l of the premix were added to the monoclonal antibody solution in the microtiter plates and incubated at 37°C. Absorbance at 405nm and 490nm of the samples was read at various times (5min to 2h) against a reagent blank (25ml TBS instead of monoclonal antibodies) in a Labsystems iEMS Reader MFTM microtiter plate reader using GENESISTM software. In parallel, the same reactions were performed except that 50ng human FIXa were added per reaction. Those antibodies that showed procoagulant activity had no chromogenic activity in the case of bovine FIX, but displayed high activity when human FIXa was present.

Fig. 11 shows the time course of the FVIII-like activity exhibited by the monoclonal antibodies 198/A1, 198/B1 and 198/AP1 with (+) and without (-) addition of 50ng human FIXa β . Non-specific polyclonal mouse IgG was used as a control. 198/A1 and 198/B1 show procoagulant activity (similar as 193/AD3 in example 6) whereas 198/AP1 does not. Antibody 198/BB1 had the same activity pattern (data not shown).

Further monoclonal antibodies selected from the 198th fusion experiment include 198/D1 (ECACC NO. 99121616), 198/T2 (ECACC No. 99121617), 198/G2 (ECACC No. 99121118), 198/U2 (ECACC No. 99121620).

Example 10: Structure and procoagulant activity of antibody derivatives derived from anti-FIX/FIXa-antibodies; Subcloning antibody variable domains from hybridoma cell lines 193/AD3, 193/K2, 198/A1 and 198/B1 (clone AB2)

Cloning procedure: Messenger RNA was prepared from 1x10⁶ hybridoma cells of the respective cell line (either 193/AD3, 193/K2, 198/A1 or 198/B1 (clone AB2)) employing the "QickPrep[®] Micro mRNA Purification Kit" (Pharmacia) according to the manufacturer's instructions. The corresponding cDNA was produced by retro transcription of mRNA using the "Ready-To-Go-You-

Prime-First-Strand Beads kit" (Pharmacia) according to the manufacturer's instructions. Heavy and light chain encoding sequences were converted to the corresponding cDNA employing a set of primers. To reverse transcribe heavy chain-specific mRNA (VH), an equimolar mixture of the oligonucleotides MOCG1-2FOR (5' CTC AAT TTT CTT GTC CAC CTT GGT GC 3') (SEQ.ID.NO. 1), MOCG3FOR (5' CTC GAT TCT CTT GAT CAA CTC AGT CT 3') (SEQ.ID.NO. 2) and MOCMFOR (5' TGG AAT GGG CAC ATG CAG ATC TCT 3') (SEQ.ID.NO. 3) was used (RTmix1). In the same reaction tube, light chain-specific cDNA (VL) was synthesized using primer MOCKFOR - (5' CTC ATT CCT GTT GAA GCT CTT GAC 3') (SEQ.ID.NO. 4).

The coding sequences for VH were amplified by PCR using the primer-sets depicted in Fig. 12 and the specific cDNA, derived from the reverse transcription mixture (RTmix1) described above, as the template. VK-chain genes were amplified using the primer sets depicted in Fig. 13 and also employing Rtmix1 as a template. The VH-PCR product was cleaved SfiI-AscI and inserted into SfiI-AscI digested vector pDAP2 (GeneBank accession no.: U35316). The pDAP2-VH constructs obtained thereby were named pDAP2-193AD3/VH, pDAP2-198A1/VH, pDAP2-198AB2/VH (derived from antibody 198/B1) and pDAP2-193/K2/VH, respectively. The plasmids were subsequently cleaved with AscI-NotI and the corresponding AscI-NotI digested VK-gene PCR product was inserted. The resultant vectors were designated pDAP2-193/AD3scFv, pDAP2-198/A1scFv, pDAP2-198/AB2scFv (derived from antibody 198/B1) and pDAP2-193/K2scFv and code for the VH-gene and the VL-gene of the monoclonal antibodies 193/AD3, 198/A1, 198/AB2 (derived from antibody 198/B1) and 193/K2. Heavy and light chains are linked by the coding sequence for an artificial, flexible linker (G₄SGGRASG₄S; Engelhardt et al., 1994) and enables expression of the scFv variant of the respective antibody.

In Fig. 14, the DNA and the deduced protein sequence of the scFv derived from the hybridoma cell line 193/AD3 are depicted. Nucleotides 1 to 357 code for the heavy chain variable domain, nucleotides 358 to 402
 5 code for the artificial flexible linker and nucleotides 403 to 726 code for the light chain variable region. The protein sequence of the CDR3 region of the heavy chain has the sequence YGNSPKGFAY (SEQ.ID.NO. 5) and is given in bold letters. The artificial linker sequence
 10 (G₄SGGRASG₄S) is shown.

In Fig. 15, the DNA and the deduced protein sequence of the scFv derived from the hybridoma cell line 193/K2 is shown. Nucleotides 1 to 363 code for the heavy chain variable domain, nucleotides 364 to 408 code
 15 for the artificial flexible linker, and nucleotides 409 to 747 code for the light chain variable region. The protein sequence of the CDR3 of the heavy chain has the sequence DGGHGYGSSFDY (SEQ.ID.NO. 6), and is given in bold letters. The artificial linker sequence
 20 (G₄SGGRASG₄S) is show.

In Fig. 16, the DNA and the deduced protein sequence of the scFv derived from the hybridoma cell line 198/AB2 (derived from antibody 198/B1) are depicted. Nucleotides 1 to 366 code for the heavy chain
 25 variable domain, nucleotides 367 to 411 code for the artificial flexible linker, and nucleotides 412-747 code for the light chain variable region. The protein sequence of the CDR3 region of the heavy chain has the sequence EGGGFTVNWYFDV (SEQ.ID.NO. 7) and is given in
 30 bold letters. The artificial linker sequence (G₄SGGRASG₄S) is also shown.

In Fig. 17, the DNA and the deduced protein sequence of the scFv derived from the hybridoma cell line 198/A1 are depicted. Nucleotides 1 to 366 code for
 35 the heavy chain variable domain, nucleotides 367 to 411 code for an artificial flexible linker, and nucleotides 412-747 code for the light chain variable region. The

protein sequence of the CDR3 region of the heavy chain has the sequence EGGGYVNWYFDV (SEQ.ID.NO.8) and is given in bold letters. The artificial linker sequence (G₄SGGRASG₄S) is also shown.

5 **Example 11: Procoagulant activity of peptides
derived from CDR3 regions of anti-FIX/FIXa-antibodies**

10 In principle, the antibody molecule can be envisioned as a biological device for the presentation of a combinatorial array of peptide elements in three dimensional space (see Gao et al., 1999, PNAS, 96:6025). Therefore, an antibody (or an antibody derivative, e.g. scFv, Fab, etc.) can be used either as a tool for the detection of functionally important domains of a specific target protein, or on the other hand, for the
15 delineation of amino acid sequences specifically mediating certain interactions, i.e. activating or enhancing the activity of FIXa towards the physiological substrate FX. The latter process has led to the evaluation of a number of heavy chain CDR3 region
20 (CDR3_H) derived peptide sequences as FIXa enhancing agents.

 Enhancing the procoagulant activity of peptides which exhibit such activity may be accomplished through sequence variation within the peptide regions critical
25 for mediating the FIXa activity enhancement. As a possible step towards peptide sequences with enhanced procoagulant activity, the binding site of an antibody, i.e. 198/A1 or 198/B1, on the FIXa molecule is mapped by employing sequence comparison analyses, competitive
30 binding assays, Western blot analyses and competitive ELISA analyses. Since the crystal structure of FIX is known, molecular modeling is subsequently used to improve the fitting of i.e. 198/B1 derived peptides in the 198/B1 binding site on human FIXa.

35 On the other hand, methodical mutational analysis of a given peptide sequence such as 198/A1 or 198/B1

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CDR3_H derived peptide sequences by, e.g., "alanine scanning mutational analysis" allows for the identification of peptide residues critical for procoagulant activity. Another way to improve the activity of a certain peptide sequence is the use of peptide libraries combined with high throughput screening.

The antigen binding site of an antibody is derived from the juxtaposition of the six "complement determining regions (CDR's)" at the N-terminal end of the VL-HL dimer (or Fv region). The contribution of a single CDR to the antibody specificity for a given antigen may vary considerably, but in general it is thought that the CDR3 region of the heavy chain (CDR3_H) is of special influence, i.e. the particular protein sequence of CDR3_H region may be highly important for antigen recognition. The length of CDR3_H regions has been reported to vary considerably and is in the range of 4-25 amino acids (Borrebaeck, p.16).

An example of a methodical mutational analysis of peptide sequences is given below. To improve the solubility/procoagulant efficacy of peptides derived from the CD3-region of anti FIX/FIXa antibodies, the N-terminal as well as the C-terminal amino acid sequences were changed. In addition, a series of mutated peptides was constructed and analyzed.

The principle of such a study is exemplified by a series of peptides derived from CDR3_H region of antibodies 198/A1 and 198/B1. The original peptide A1 (see table 2) is derived from the CDR3_H region of antibody 198/A1 and peptide B1 is derived from the CDR3_H region of antibody 198/B1, respectively (see example 10, Fig. 16 and 17). The term "scrambled version" means that a peptide has the same amino acids but in random order.

Peptide	Sequence	Amino-acids	MW (D)	pI	Remark
A1	EGGGYYVNWYFDV (SEQ.ID.No. 9)	(13aa)	1569	7.2	Decreased solubility
A1/1	VYGFGWGYEVNDY (SEQ.ID.No. 10)	(13aa)	1569	7.1	Scrambled version of A1,
A1/2	EEEEGGGGYYVNWYFDEEE (SEQ.ID.No. 11)	(18aa)	2244	5.8	Acidic pI, soluble,
A1/3	RRREGGGGGYYVNWYFDRRR (SEQ.ID.No. 12)	(18aa)	2407	9.9	Basic pI, soluble,
A1/4	EYGEYGEVNEYDEFWE (SEQ.ID.No. 13)	(18aa)	2244	5.8	Scrambled version of A1/2
A1/5	VRYNRYRWGYRGRFGDE (SEQ.ID.No. 14)	(18aa)	2407	9.9	Scrambled version of A1/3
A1/3-scr3	RRRGEYGVYWNGDFYRRR (SEQ.ID.No. 15)	(18aa)	2407	9.9	Scrambled version of A1/3
A1/3-Rd	RdRdRdEGGGYYVNWYFDRdRdRd (SEQ.ID.No. 16)	(18aa)	2407	9.9	Peptide A1/3 but substitute D-Arg for L-Arg
A1/3-Rd-srmb	RdRdRdGEYGVYWNGDFYRdRdRd (SEQ.ID.No. 17)	(18aa)	2407	9.9	Scrambled version of A1/3-Rd

Table 2

List of a series of antibody 198/A1 derived peptides. Listed are the length of the peptide (aa, amino acids #), the calculated molecular weight (MW, in Dalton (D) and the statistical isoelectric point (pI). D-Arg is abbreviated as Rd.

In a first series of experiments we improved the solubility of the original CDR_{3H} peptide sequence (A1; EGGGYVNWYFDV) by removing the C-terminal Val residue and adding several charged residues at the N- as well as the C-terminal end of the peptide. The resulting peptides, A1/2 (acidic pI), A1/3 (basic pI) and their respective scrambled versions A1/4, A1/5 and A1/3scr3 were readily soluble in a variety of buffer systems at physiological pH.

To analyze the FVIII-like (FIXa activating) activity of the peptides, an assay system based on a commercial available FVIII assay was developed (see examples 2 and 4). The basic principle is, that without a cofactor, FIXa will have very limited activity towards its natural substrate FX. Only in the presence of a substance having FIXa activation properties, i.e. FVIII or a substance exhibiting FVIII-like activity, a substantial amount of FXa is produced by cleavage of FX through the FIXa/activator complex. The amount of FXa generated is monitored by cleavage of a chromogenic substrate. The principle of the revised chromogenic assay is described for two representative peptides: A1/3 and A1/5 (Table 2). Briefly, 25µl aliquots of peptide stock solution (in imidazole buffer (IZ) 50mM imidazole, 100mM NaCl, pH7.2) were transferred to microtiter plate wells and warmed to 37°C. Chromogenic FXa substrate (S-2222), synthetic thrombin inhibitor (I-2581), bovine FIXa and bovine FX were reconstituted in sterile water and FIXa/FX mixed with phospholipids according to the supplier's protocol. Since the peptides do not react with bovine FIXa, (which comes as a mixture with bovine FX in the Test Kit) 2,9nM (in most cases 2.3nM) human FIXa (ERL) were added (see Example 11, Fig 19). Per reaction, 50µl of the phospholipid /FIXa/FX solution were combined with 25µl CaCl₂ (25mM) and 50µl of the substrate/inhibitor cocktail. To start the reaction, 125µl of the premix were added to the peptide solution in the microtiter plate and incubated at 37°C. Absorbance at 405nm and 490nm of the samples was read at various times (5 min to 2h) against a reagent blank in a Labsystems iEMS Reader MF™ microtiter plate reader using GENESIS™ software.

The result of this experiment are shown in Example 11, Fig 18. Peptide A1/3 induced a readily measurable

FXa generation in the presence of 2.9nM human FIXa, whereas the scrambled version A1/5 was inactive. In addition, the acidic peptide A1/2 as well as the scrambled versions A1/4 and A1/3-scr3 did not give any significant chromogenic activity when tested under comparable conditions (data not shown). To prove that the peptide A1/3 like the parental antibody 198/A1 does not react with bovine FIXa and FX the experiment shown in Fig. 19 was done. The peptide A1/3 was incubated as described above with (A1/3 (24 μ M), +hFIXa) and without (A1/3 (24 μ M), w/o hFIXa) 2.3nM human FIXa (hFIXa). In a control experiment we added plain dilution buffer (IZ) supplemented with 2.3nM hFIXa to the reaction mixture. As shown in Fig. 19, the reaction takes place only in the presence of human FIXa.

Fig. 18 demonstrates the chromogenic FVIII-like activity of peptide A1/3 in the presence of 2.9nM human FIXa (hFIXa). The scrambled version of peptide A1/3, peptide A1/5 does not give rise to any FXa generation. Fig. 19 demonstrates the dependence of the chromogenic FVIII-like activity of peptide A1/3 on the presence of human FIXa (hFIXa). In the absence of human FIXa, peptide A1/3 does not give rise to any FXa generation. The buffer control, plain imidazole buffer is designated IZ.

The peptides were also analyzed for their potential to reduce the clotting time in a FVIII deficient plasma. The aPTT based one stage clotting assay was essentially done as described (see example 6). Clotting times (time from starting the reaction to the "clot"-formation were compared either against FVIII, a buffer control (IZ) or a control peptide (scrambled version). The results of two typical clotting experiments done with two different aPTT reagents (DAPTTIN and Pathromtin SL) are shown in table 3A and table 3B.

Exp. 1	peptide conc.	w/o FIXa sec	w/o FIXa sec	average sec	2.2nM FIXa sec	2.2nM FIXa sec	average sec
IZ	0	107.7	106.8	107	93.1	94.5	94
A1/3	15 μ M	78.2	77.1	78	59.3	59.9	60
	12.5 μ M	80.2	80.6	80	60.2	58.9	60
	7.5 μ M	97.8	97.9	98	73.1	72.7	73
	2.5 μ M	105.2	104.8	105	91.1	91	91
A1/3- scr3	15 μ M	122.5	122	122	106.1	105.5	106
	12.5 μ M	116	117.6	117	103.1	104.5	104
	7.5 μ M	114.2	113.9	114	100.8	100.6	101
	2.5 μ M	107.8	107.4	108	96.3	95.2	96
Exp. 2	peptide conc.	w/o FIXa sec	w/o FIXa sec	average (sec)	2.2nM FIXa sec	2.2nM FIXa sec	average (sec)
IZ	0	111	109.7	110	94.7	95.5	95
A1/3	12.5 μ M	83.6	85.5	85	56.7	56.7	57
	10 μ M	79.1	78.5	79	63.1	62.5	63
	7.5 μ M	100.1	100.5	100	71.6	73.9	73
	5 μ M	103.4	104.8	104	77	76	77
	2.5 μ M	110.1	108.9	110	88	88.8	88
	1.25 μ M	108.7	109.3	109	90.7	90.8	91

- Table 3A. Clotting activity of peptides A1/3 and A1/3-scr (scrambled version of A1/3) in FVIII deficient plasma either in the presence or in the absence (w/o) of 2.2nM human FIXa. Shown are two independent representative experiments (Exp. 1 and Exp. 2). All clotting experiments have been done in duplicate. Given are the clotting times for the individual experiments and the average clotting time in seconds (sec).
- Experiments shown in table 3A have been done employing the aPTT reagent DAPTTIN (Baxter Hyland Immuno). Compared to the buffer control (IZ, imidazole buffer) the peptide A1/3 gave rise to a dose dependent reduction in the clotting time. The reduction in the clotting time became much more pronounced by the addition of 2.2nM activated human FIX to the reaction mix. The scrambled version of peptide A1/3, A1/3-scr3 did not show any reduction of the clotting time. In fact, at concentrations above 2.5 μ M, the scrambled peptide became

inhibitory and therefore prolonged the clotting time. Peptides A1/1, A1/2, A1/4 and A1/5 did not give any reduction in the clotting time indicating that they lack procoagulant activity (data not shown).

5

	Final conc.	w/o FIXa sec	w/o FIXa sec	average sec	2.2nM FIXa sec	2.2nM FIXa sec	average sec
IZ	0	131.8	132.1	132	107.9	108.7	108
FVIII	12.5mU/ml	68.9	69	69	52.9	53.6	53
	6.25mU/ml	77.8	77.9	78	58.6	58.9	59
A1/3							
	15µM	152.8	149.3	151	75.4	75.2	75
	10µM	135.7	134.6	135	76.2	79.8	78
	5µM	152.6	155.6	154	86.6	90.2	88
	1µM	138.3	138.8	139	103.7	105.9	105

Table 3B. Clotting activity of peptide A1/3 in FVIII deficient plasma when Pathromtin SL (DADE Behring) is used as an aPTT reagent. The experiments were done in duplicate, either in the presence or in the absence (w/o) of 2.2nM human FIXa. Given are the clotting times for the individual experiments and the average clotting time in seconds (sec). Factor VIII and imidazole buffer (IZ) were included as positive and negative control respectively.

In contrast to the experiments shown in table 3A the experiments shown in table 3B have been done employing the aPTT reagent Pathromtin SL. In the presence of FIXa, the peptide A1/3 gave rise to a dose dependent reduction in the clotting time whereas in the absence of FIXa no reduction of the clotting time was detectable.

In another series of experiments we set out to improve the plasma stability (protection from, e.g., proteolytic degradation) of peptide A1/3. One approach was to substitute the N- and C-terminal L-Arg residues

with D-Arg residues (exemplified by peptides A1/3-rd and A1/3-Rd-srmb). Peptides A1/3-rd and A1/3-Rd-srmb (scrambled version of the peptide) were then analyzed in a chromogenic as well as in the aPTT based clotting assay. These experiments revealed that exchanging the terminal L-Arg residues for D-Arg residues did not change the FVIII-like activity as measured in the chromogenic assay, indicating that chirality of the Arg-residues does not play a major role in chromogenic activity (Fig. 20). In addition, the aPTT based one-stage clotting activity, although somewhat reduced, was still easily detectable (Table 4).

	Peptide conc.	w/o FIXa sec	w/o FIXa, sec	average sec	2.2nM FIXa sec	2.2nM FIXa sec	average sec
IZ	0	110	109.1	110	96	96	96
A1/3	15µM	77.8	78	78	56.1	55.5	56
	12.5µM	99.4	100.5	100	65	68	67
	10µM	104.4	104.5	104	72	73.2	73
	7.5µM	105.2	105.2	105	80.7	80.5	81
	5µM	108.4	107.7	108	89.7	88.3	89
	2.5µM	107.9	107.6	108	93.6	93.3	93
	1.25µM	106.7	107	107	94.4	95	95
A1/3-Rd	15µM	96.4	95.4	96	76.1	74.4	75
	12.5µM	98	98.6	98	72.3	73.7	73
	10µM	93.5	95.8	95	74.2	77.2	76
	7.5µM	97.6	98.1	98	80.9	82.2	82
	5µM	99.2	99.1	99	86	85.1	86
	2.5µM	102.7	103.4	103	94.4	94.7	95
	1.25µM	107.5	107.7	108	96.6	96	96
A1/3-Rd srmb	15µM	121.9	121.3	122	112.7	112.4	113
	12.5µM	117.2	118	118	108.1	107.8	108
	10µM	115.8	115.3	116	107.2	107.8	108
	7.5µM	114.6	113.6	114	107.6	106.6	107
	5µM	113.1	112.4	113	108.5	108.2	108
	2.5µM	111.9	111.9	112	105	104.2	105
	1.25µM	107.2	107.1	107	101.1	105.3	103

Table 4. One stage clotting activity of peptides A1/3, A1/3-Rd and A1/3-Rd-srmb (sequences see table 2). IZ, buffer control.

Fig. 20 demonstrates the unchanged chromogenic activity of peptide A1/3-Rd. Peptides at a final concentration of 12 μ M or the buffer control (IZ) were incubated in the presence of 2.3nM human FIXa (+). The chromogenic activity of peptide A1/3 and A1/3-Rd was found to be virtually unchanged and gave almost identical results in the chromogenic assay. The scrambled version of peptide A1/3, A1/5 as well as the buffer gave no significant FXa generation.

In the next series of experiments we set out to determine the individual role of any amino acid of the peptide core sequence by substituting each residue for the amino acid Alanine (Table 5).

Peptide	Sequence	Amino acid #	MW (D)	pI	Remark
A1/3	RRREGGGYYVNWYFDRRR (SEQ.ID.No. 18)	(18aa)	2407	9.9	Basic pI, soluble,
A1/3-13	RRR A GGGYVNWYFDRRR (SEQ.ID.No. 19)	(18aa)	2349	10.4	E ₁ -A ₁
A1/3-1	RRRE A GGYYVNWYFDRRR (SEQ.ID.No. 20)	(18aa)	2421	9.9	G ₂ -A ₂
A1/3-2	RRREG A GGYYVNWYFDRRR (SEQ.ID.No. 21)	(18aa)	2421	9.9	G ₃ -A ₃
A1/3-3	RRREGG A YYVNWYFDRRR (SEQ.ID.No. 22)	(18aa)	2421	9.9	G ₄ -A ₄
A1/3-4	RRREGGG A YVNWYFDRRR (SEQ.ID.No. 23)	(18aa)	2315	9.9	Y ₅ -A ₅
A1/3-5	RRREGGGY A VNWYFDRRR (SEQ.ID.No. 24)	(18aa)	2315	9.9	Y ₆ -A ₆
A1/3-6	RRREGGGYY A NWYFDRRR (SEQ.ID.No. 25)	(18aa)	2379	9.9	V ₇ -A ₇
A1/3-7	RRREGGGYYV A WYFDRRR (SEQ.ID.No. 26)	(18aa)	2364	9.9	N ₈ -A ₈
A1/3-8	RRREGGGYYVN A YFDRRR (SEQ.ID.No. 27)	(18aa)	2292	9.9	W ₉ -A ₉
A1/3-9	RRREGGGYYVNW A FDRRR (SEQ.ID.No. 28)	(18aa)	2315	9.9	Y ₁₀ -A ₁₀
A1/3-10	RRREGGGYYVNWY A DRRR (SEQ.ID.No. 29)	(18aa)	2331	9.9	F ₁₁ -A ₁₁
A1/3-11	RRREGGGYYVNWYF A RRR (SEQ.ID.No. 30)	(18aa)	2363	10.5	D ₁₂ -A ₁₂
A1/3-12srm	RRRYVYNGWGYFEG A RRR (SEQ.ID.No. 31)	(18aa)	2363	10.4	Scrambled version

Table 5. Listed are the peptides designed to elucidate

the role of any single amino acid within the peptide core sequence ($E_1G_2G_3G_4Y_5Y_6V_7N_8W_9Y_{10}F_{11}D_{12}$). The lower case numbers describe the position of the amino acid within the peptide. Alanine, an uncharged small amino acid, was substituted for each amino acid ("Alanine scan"). Also listed are the lengths of the peptides (amino acids #), the calculated molecular weights (MW, in Dalton (D) and the statistical isoelectric points (pI).

Each of the peptides was dissolved individually in imidazole buffer (50mM imidazole, 100mM NaCl, pH7.2) and subsequently diluted in clotting buffer (50mM imidazole, 100mM NaCl, 1% human albumin, pH7.4) to the desired final concentration. The peptides were analyzed for their chromogenic activity as well as for their potential to reduce the clotting time in a FVIII deficient plasma. The one-stage clotting assay was essentially done as described (see example 6). Clotting times (time from starting the reaction to the "clot"-formation were compared either against a buffer control or a control peptide (scrambled version).

Some of the results of the "Alanine scan" are given for the peptides A1/3-2 and A1/3-3. The change of G_3 - A_3 as exemplified in the peptide A1/3-2 yields high chromogenic activity and a strong reduction of the one-stage clotting time (34 seconds at a concentration of $12.5\mu\text{M}$) in the presence of 2.2nM human FIXa. Peptide A1/3-3 (G_4 - A_4) exhibits an optimum of chromogenic activity around a final concentration of $12\mu\text{M}$ with decreased activity at either higher or lower concentrations. The peptide is somewhat inhibitory in a one-stage clotting assay at higher concentrations ($12.5\mu\text{M}$) in the absence of FIXa but becomes strongly active in the presence of 2.2nM FIXa (31 seconds, $12.5\mu\text{M}$).

In the next series of experiments we set out to determine the individual role of any amino acid of the peptide core sequence by substituting each core residue

for the amino acid glutamic acid (E) (see Table 6).

Peptide	Sequence	Amino-Acids	MW (D)	pI	Remark
A1/3	RRREGGGYYVNWYFDRRR	(18aa)	2407	9.9	Basic pI, soluble,
A1/3-22	RRREEGGGYYVNWYFDRRR (SEQ.ID.No. 32)	(18aa)	2479	9.5	G ₂ -E ₂
A1/3-23	RRREGEGGYVNWYFDRRR (SEQ.ID.No. 33)	(18aa)	2479	9.5	G ₃ -E ₃
A1/3-24	RRREGGEYYVNWYFDRRR (SEQ.ID.No. 34)	(18aa)	2479	9.5	G ₄ -E ₄
A1/3-26	RRREGGGGEYVNWYFDRRR (SEQ.ID.No. 35)	(18aa)	2373	9.4	Y ₅ -E ₅
A1/3-27	RRREGGGYEVNWYFDRRR (SEQ.ID.No. 36)	(18aa)	2373	9.4	Y ₆ -E ₆
A1/3-28	RRREGGGYYENWYFDRRR (SEQ.ID.No. 37)	(18aa)	2437	9.5	V ₇ -E ₇
A1/3-29	RRREGGGYYVEWYFDRRR (SEQ.ID.No. 38)	(18aa)	2422	9.5	N ₈ -E ₈
A1/3-30	RRREGGGYYVNEYFDRRR (SEQ.ID.No. 39)	(18aa)	2350	9.5	W ₉ -E ₉
A1/3-31	RRREGGGYYVNWEFDRRR (SEQ.ID.No. 40)	(18aa)	2373	9.4	Y ₁₀ -E ₁₀
A1/3-32	RRREGGGYYVNWYEDRRR (SEQ.ID.No. 41)	(18aa)	2389	9.5	F ₁₁ -E ₁₁
A1/3-33	RRREGGGYYVNWYFERRR (SEQ.ID.No. 42)	(18aa)	2421	9.9	D ₁₂ -E ₁₂
A1/3-34srm	RRRGEYGEYWNNGDFYRRR (SEQ.ID.No. 43)	(18aa)	2437	9.5	Scrambled version

Table 6. Listed are the peptides designed to elucidate the role of any single amino acid within the peptide core sequence (E₁G₂G₃G₄Y₅Y₆V₇N₈W₉Y₁₀F₁₁D₁₂). The lower case numbers describe the position of the amino acid within the peptide. Glutamic acid, a negatively charged large amino acid, was substituted for each amino acid of the core sequence (''Glutamic acid scan''). Also listed are the lengths of the peptide (amino acids #), the calculated molecular weights (MW, in Dalton (D) and the statistical isoelectric points (pI).

Each of the peptides was solved individually in imidazole buffer (50mM imidazole, 100mM NaCl, pH7.2) and subsequently diluted in clotting buffer (50mM imidazole, 100mM NaCl, 1% human albumin, pH7.4) to the desired

final concentration. The peptides derived from the
 "Glutamic acid scan" series were analyzed for their
 chromogenic FVIII-like activity as well as for their
 potential to reduce the clotting time in a FVIII
 deficient plasma. The one-stage clotting assay was
 essentially done as described (see example 6).

The peptide A1/3-24 showed some interesting
 properties. The molecule exhibited high chromogenic
 FVIII-like activity at concentrations between 6.5 μ M-12 μ M
 but lost activity at higher concentrations (up to 24 μ M).
 The peptide had no procoagulant activity in the absence
 of human FIXa but was strongly active in the presence of
 2.2nM hFIXa.

In a second series of experiments we set out to
 improve the procoagulant activity of the antibody 198/B1
 CDR3H derived peptide sequence B1. In a first step we
 improved the solubility of the original peptide sequence
 (B1; EGGGFTVNWYFDV) by removing the C-terminal Val
 residue and adding several charged residues at the N- as
 well as the C-terminal end of the peptide. The resulting
 peptides B1/4, B1/6 (acidic pI), B1/7 (basic pI) and
 their scrambled versions B1/5, B1/7scr3 are readily
 soluble in a variety of buffer systems at physiological
 pH.

Peptide	Sequence	Amino- acids	MW (D)	pI	Remark
B1	EGGGFTVNWYFDV (SEQ.ID.No. 44)	(13aa)	1491	6.0	Decreased solubility
B1/4	REGGGFTVNWYFDR (SEQ.ID.No. 45)	(14aa)	1704	7.9	Soluble,
B1/5	FGVGYRGETRNFWD (SEQ.ID.No. 46)	(14aa)	1704	8.0	Scrambled version, soluble
B1/6	EEEEGGGFTVNWYFDEEE (SEQ.ID.No. 47)	(18aa)	2166	5.0	Acidic pI soluble
B1/7	RRREGGGFTVNWYFDRRR (SEQ.ID.No. 48)	(18aa)	2329	9.9	Basic pI soluble
B1/7scr3	RRRFGVGYGETNFDWRRR (SEQ.ID.No. 49)	(18aa)	2329	9.9	Basic pI, soluble, scrambled version

Table 7 is a list of a series of antibody 198/B1 derived

Peptides B1/4 and B1/5 were soluble in 50mM Tris,
5 100mM NaCl, pH=6.5. Both peptides were analyzed in a
chromogenic FVIII assay. Peptide B1/4 but not the
scrambled version B1/5 was found to have some
chromogenic activity (data not shown).

The FIXa activating activity (FVIII cofactor-like activity) from peptide B1/7 was first measured in the chromogenic assay described above.

Fig. 21 demonstrates the chromogenic activity of

peptide B1/7. The peptide at a final concentration of $2.4\mu\text{M}$ or the buffer control (IZ) were incubated in the presence of 2.3nM human FIXa.

In Fig. 22 peptide B1/7 at a final concentration of $2.4\mu\text{M}$ or the buffer control (IZ) were incubated in the presence of 2.3nM human FIXa (as indicated either as '+2.3nM hFIXa' or '+'). The chromogenic activity of peptide B1/7 was found to be dependent on the presence of FIXa and FX since no reaction is detectable when FIXa and FX are left out of the reaction (w/o FIXa/FX). To prove that the peptide B1/7 mediates indeed FXa generation, the FXa specific protease inhibitor Pefabloc Xa was added to the reaction mix ($35\mu\text{M}$ Pefabloc Xa). In a second set of experiments, the procoagulant effect of peptides B1/6, B1/7 and B1/7scr3 were tested in a aPTT based one-step coagulation assay. The experiments were done essentially as described in Example 6. The results are shown in tables 8 and 9.

Peptide	$12.5\mu\text{M}$ (-)	$1.25\mu\text{M}$ (-)	$0.125\mu\text{M}$ (-)	12.5nM (-)	Buffer (-)	remarks
B1/6	115	110	111	111	110	
B1/7	157	112	109	110	110	
B1/7scr3	115	105	106	105	107	

Table 8: FVIII deficient plasma was incubated either with peptides B1/6, B1/7scr3 or B1/7 in the absence of activated human FIX. As a negative control, plain buffer was added to the deficient plasma. The clotting times for the various combinations are given. Under these conditions, peptide B1/7 at its highest concentration ($12.5\mu\text{M}$) becomes inhibitory to the coagulation process as indicated by the extended clotting time of 157 seconds.

Peptide	12.5 μ M (+)	1.25 μ M (+)	0.125 μ M (+)	12.5nM (+)	Buffer (+)	remarks
B1/6	103	100	101	100	100	
B1/7	83	92	99	99	100	
B1/7scr3	102	94	94	94	94	

Table 9: FVIII deficient plasma was incubated either with peptides B1/6, B1/7scr3 or B1/7 in the presence of activated human FIX. As a negative control, plain buffer was added to the deficient plasma. The clotting times for the various combinations are given. In the presence of FIXa, peptide B1/7 becomes procoagulant as indicated by the reduced clotting time (83 seconds compared to 102 seconds for the scrambled peptide and 100 seconds for the buffer control).

Example 12: Procoagulant activity of peptide derivatives obtained from CDR3 regions of anti-FIX/FIXa-antibodies in FVIII inhibitor plasma

To assay for the procoagulant activity of peptide A1/3 in FVIII inhibitor plasma the following experiment was carried out. We performed a standard aPTT based one stage clotting assay, but instead of FVIII deficient plasma we employed FVIII inhibitor plasma. The inhibitory potency of the plasma was 8.1 Bethesda Units per ml.

		w/o FIXa	w/o FIXa			FIXa	FIXa	
	Peptide conc.	sec	sec	Average sec		sec	sec	average sec
IZ	0	104.8	103.6	104		94.2	94.1	94
A1/3	12.5 μ M	85.8	85.3	86		61	60.2	61
	10 μ M	88.4	87.9	88		61.3	61.8	62
	7.5 μ M	93.7	92.7	93		68.8	70.9	70
	5 μ M	101.5	101.1	101		81	82	82
	2.5 μ M	106.1	105.3	106		90.2	90.5	90
	1.25 μ M	104.5	104.3	104		91.3	91.4	91

Table 10: Various amounts of peptide A1/3 (12.5 μ M-1.25 μ M) were added to FVIII inhibitor plasma (either in

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Example 13: Conversion of the 196/C4 IgM into IgG1

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The coding sequence of a human kappa light chain constant domain is amplified from the cDNA by PCR using specific primers.

The gene of a human gamma 1 chain constant region (CH1-hinge-CH2-CH3) is amplified from the cDNA by PCR using specific primers.

The PCR product of the light chain constant domain is digested with XbaI and NheI and inserted into digested pSI. The resultant vector is cleaved with EcoRI and XbaI and annealed oligonucleotides are inserted, resulting in vector pSI-Ckappa. The annealed

oligonucleotides provide for the leader and the SacI-XbaI sites for insertion of the kappa chain variable region. The PCR product of the human gamma 1 chain constant region is digested with SpeI and BamHI and
 5 inserted into digested pSI. The resultant vector is cleaved with SpeI and NotI and annealed oligonucleotides are inserted resulting in vector pSI-Cgamma. The annealed oligonucleotides provide for the leader and the XhoI-BstEI sites for insertion of the heavy chain
 10 variable region. Vector pEF/Bsd is digested NheI and SfiI, blunt ended by Klenow treatment and the whole expression cassette of pSI-Ckappa, excised with BglII and BamHI, is inserted (after Klenow treatment). The resultant vector is digested with EcoRI and HindIII and
 15 treated with Klenow. The whole expression cassette of pSI-Cgamma is excised with BglII and BamHI and is inserted (after Klenow treatment). The resultant vector is named pBax-IgG1.

The light chain variable region can be inserted in
 20 between the SacI-XbaI sites, yielding the complete coding-sequence of a kappa light chain. The heavy chain variable region can be cloned in between the XhoI-BstEI sites, resulting in a complete IgG1 heavy chain gene. Both open reading frames are expressed under the control
 25 of the SV40-promoter and harbour the coding sequence of a signal peptide at the 5' end of the genes for secretion of the heavy and light chains into the endoplasmatic reticulum. Transfection into COS cells allows the expression of an IgG1 with the same binding
 30 properties as the parental IgM.

Construction of the plasmid pBax-196/C4 is further accomplished by amplifying the VH of the 196/C4 scFv (subcloned as described in Experiment 10) by PCR using
 specific primers. The PCR product is digested with XhoI
 35 and BstEII and inserted into XhoI and BstEII digested pBax IgG1. The VL of the 196/C4 scFv is amplified by PCR using specific primers. The PCR product is digested with

SacI and XbaI and inserted into SacI and XbaI-digested pBax IgG1-VH. The resultant vector (pBax-196/C4) is transfected into COS cells by electroporation, and hybrid IgG1 molecules (murine variable region and human constant region) with the same specificity as the parental IgM is expressed.

Example 14: Activation of FIXa amyolytic activity by anti-FIXa antibodies:

Briefly, 20 μ l factor IXa (containing 200mU FIXa (Stago)) were incubated at 37°C, with 200 μ l of reaction buffer (50mM TrisHCl pH7.4, 100mM NaCl, 5mM CaCl₂ and 40% Ethyleneglycol), 25 μ l of FIXa substrate (CH₃SO₂-D-CHG-Gly-Arg-pNA, AcOH, 10 μ M/ml, Pentapharm LTD) in the absence or presence of various amounts of anti-FIX antibodies 198/B1 (IgG isotype) or 196/AF1 (IgM isotype). Specific cleavage of FIXa substrate was monitored at 405nm in an ELISA reader.

The presence of the anti-FIX antibodies enhanced the amyolytic activity of FIXa at least 2 fold. Fig. 24 shows the increase of the amidolytic activity of FIXa in the presence of antibody 198/B1 (Fig. 24A) and antibody 198/AF1 (Fig. 24B).

Example 15: FVIII-like activity exhibited by Fab fragments derived from anti- FIX/FIXa-antibodies.

Fab fragments of anti-FIX/FIXa antibodies were prepared and purified according to standard protocols. Briefly, 1ml antibody 198/A1 (4mg/ml in 50mM imidazole, 100mM NaCl, pH7.4) was incubated overnight with 87 μ l fragmentation buffer (1M Na Acetate, 10mM EDTA 67.5mg/ml L-cysteine) and 0.25mg papain (immobilized on agarose beads), at 37°C. The preparation was filtered to remove the papain. L-histidine was added (final concentration 50mM) and afterwards the pH was adjusted to 7.0. Finally, solid NaCl is added to give a final concentration of 1M.

Subsequently, the 198/A1 Fab fragment was purified

by binding to protein L: We used ImmunoPure Immobilized
 PROTEIN L Plus (Pierce) in a PHARMACIA XK 16/20 Column
 (gel-volume: 2ml) Buffers for chromatography were: 1)
 equilibration-buffer : 50mM L-histidine pH 7.0; 1M
 5 NaCl; 0,1% (w/v) NaN_3 ; 2) wash-buffer: 50mM L-Histidine
 pH 7.0; 0.1% (w/v) NaN_3 ; 3) elution-buffer: 100 mM
 glycine pH 2.5; 0.1% (w/v) NaN_3 ; and 4) neutralization
 buffer: 2M Tris/Cl pH 8,0;

10 Chromatography was essentially done by following
 steps 1 to 7 described in table 11. In order to
 neutralize the low pH of the elution buffer "Fraction-
 tubes" were pre-loaded with 0.2 ml 2M Tris pH 8.0.

	STEP	BUFFER	Flow rate	Vol.	CV	Fractions
1.	column-wash	elution-buffer	2.0 ml/min	10 ml	5	waste
2.	equilibration	equi-buffer	2.0 ml/min	10 ml	5	waste
3.	sample-load	sample	1.0 ml/min	x ml	x	flow-through
4.	wash 1	equi-buffer	1.0 ml/min	20 ml	10	flow-through
5.	wash 2	wash-buffer	1.0 ml/min	10 ml	5	flow-through
6.	elution	elution-buffer	1.0 ml/min	15 ml	7.5	1,0 ml fractions-
7.	neutralization	wash-buffer	2.0 ml/min	10 ml	5	waste

15 Table 11

The final 198/A1 Fab preparation was dialyzed
 against 50mM imidazole, 100mM NaCl, pH7.4 and analyzed
 in a chromogenic FVIII assay as described above (Fig.
 20 25). Compared to an intact antibody, the 198/A1 Fab
 fragment has somewhat less activity; however, the Fab
 fragment still gives rise to FIX dependent FXa
 generation.

Fig. 25 demonstrates the chromogenic FVIII-like
 25 activity of the antibody 198/A1 Fab fragment in the
 presence of 2.3nM human FIXa. As a positive control we
 used the intact antibody 198/A1 as well as 7.5pM FVIII.
 Buffer control (IZ) instead of 198/A1 Fab fragment or
 FVIII was used as a negative control.

30 **Example 16: FVIII-like activity exhibited by fusion**

proteins between scFv fragments of anti-FIX/FIXa antibodies and E. coli alkaline phosphatase.

The single chain Fv fragment (see example 10) of antibody 198/B1 (subclone AB2) was fused to the N-terminus of E. coli alkaline phosphatase employing the pDAP2 vector system (Kerschbaumer et al., 1996). Two identical clones were isolated and designated pDAP2-198AB2#1 and pDAP2-198AB2#100 (Fig. 26). The resulting fusion proteins were expressed in E. coli, purified by metal affinity chromatography (Kerschbaumer et al., 1997) and analysed in a standard chromogenic assay (Fig. 27).

Fig. 27 demonstrates the chromogenic FVIII-like activity of two antibody 198/B1 (subclone AB2) scFv fragment-alkaline phosphatase fusion proteins (198AB2#1 and 198AB2#100) in the presence of 2.3nM human FIXa. As a positive control we used 7.5pM FVIII.

Example 17: FVIII-like activity exhibited by a bivalent miniantibody.

In order to obtain a bivalent miniantibody, the scFv fragment of antibody 198/B1 (subclone AB2) was fused to a amphipatic helical structure employing the pZip1 vector system (Kerschbaumer et al. (*Analytical Biochemistry* 249, 219-227, 1997)). Briefly, the gene of the 198/B1 scFv fragment was isolated from the plasmid pDAP-198AB2#100 (example 16) by digestion with SfiI and NotI. The DNA fragment was gel purified and inserted in the SfiI/NotI digested vector pZip1. The resulting plasmid was sequenced and designated pZip-198AB2#102 (Fig.28). In parallel, we constructed a miniantibody version from an irrelevant monoclonal antibody termed #8860. In a first step, the single chain Fv fragment of antibody #8860 was assembled in the vector pDAP2. The cloning was done essentially as described in example 10. The construct was named pDAP2-8860scFv#11 (Fig. 29).

Subcloning of the scFv fragment contained within pDAP2-8860scFv#11 into plasmid pZip1 (see above) yielded the miniantibody construct p8860-Zip#1.2 (Fig. 30). Since antibody #8860 does not react with FIX/FIXa (as judged by Western Blot and ELISA analysis) it represents an appropriate negative control. Subsequently, the miniantibody proteins were expressed in *E. coli* and purified from bacterial supernatants by binding to Protein L according to the following protocol:

For affinity chromatography we used ImmunoPure Immobilized PROTEIN L Plus (Pierce) in a PHARMACIA XK 16/20 Columns having a gel-volume of 4ml

Buffers employed were: 1) equilibration-buffer : 50mM L-Histidine pH 7.0, 1M NaCl, 0.1% (w/v) NaN_3 , wash-buffer: 50mM L-histidine pH 7.0, 0.1% (w/v) NaN_3 , elution-buffer: 100 mM glycine pH 2.5, 0.1% (w/v) NaN_3 , and neutralization buffer: 2M Tris/Cl pH 8.0

Samples were prepared as follows: The bacterial culture supernatant was obtained by centrifugation of the bacterial expression culture (11,000 x g, 4°C, 10 minutes). 470 g of ammonium-sulphate was added to 1 liter of supernatant and the solution stirred on ice for 1 hour to precipitate the protein. The precipitate was pelleted at 14,000 x g for 35 minutes at 2°C and re-dissolved in 100 ml 20mM Tris pH 7.0. Subsequently the concentrate was dialyzed against 20mM Tris pH 7.0, L-histidine was added to a final concentration of 50mM and the pH was adjusted to 7.0. Finally, solid NaCl was added to give a final concentrations of 1M. Before loading on the column, a sample was first centrifuged at 16,000 x g for 15 min at room temperature and then filtered through a 0.45µm sterile filter.

Chromatography was essentially done by following steps 1 to 7 described in table 12. In order to neutralize the low pH of the elution buffer "Fraction-tubes" were pre-loaded with 0.2 ml 2M Tris pH 8.0.

	STEP	BUFFER	Flow rate	Vol.	CV	Fractions
1.	column-wash	elution-buffer	2.0 ml/min	20 ml	5	waste
2.	equilibration	equi-buffer	2.0 ml/min	20 ml	5	waste
3.	sample-load	sample	1.0 ml/min	x ml	x	flow-through
4.	wash 1	equi-buffer	1.0 ml/min	40 ml	10	flow-through
5.	wash 2	wash-buffer	1.0 ml/min	20 ml	5	flow-through
6.	elution	elution-buffer	1.0 ml/min	30 ml	7.5	1.0 ml fractions-
7.	neutralization	wash-buffer	2.0 ml/min	20 ml	5	waste

Table 12. The final 198/B1 (subclone AB2) miniantibody preparation (designated 198AB-Zip#102) and the negative control 8860-Zip#1.2 were dialyzed against 50mM imidazole, 100mM NaCl, pH7.4 and analyzed in a chromogenic FVIII assay as described above (Fig. 31).

As can be seen in Fig. 31, the miniantibody construct 198AB-Zip#102 gives rise to substantial FXa generation (compare to FVIII) whereas the negative control miniantibody 8860-Zip#1.2 does not.

Fig. 31 demonstrates the chromogenic FVIII-like activity of the 198/B1 (subclone AB2) miniantibody 198AB-Zip#102 in the presence of 2.3nM human FIXa. As a positive control we used 4.8pM FVIII whereas an unrelated miniantibody (8860-Zip#1.2) and plain reaction buffer (IZ) served as negative controls.

Example 18: FVIII-like activity exhibited by anti-FIXa/FIX antibody scFv fragments

The single chain Fv fragment of antibody 198/B1 (subclone AB2) as well as the scFv fragment of antibody #8860 were expressed employing the pMycHis6 vector system. Vector pMycHis6 (Fig. 32 & 33) was constructed by cleaving vector pCOCK (Engelhardt et al., 1994, Biotechniques, 17:44-46) with NotI and EcoRI and insertion of the following oligonucleotides: mychis6-co: 5'ggccgcagaacaaaaactcatctcagaagaggatctgaatggggcggcacatcaccatcaccatcactaataag 3' (SEQ.ID.NO. 79) and mycchis-ic:5'aattcttattagtgatggtgatggtgatgtgccgccccattcagatcctctcttgagatgagtttttgttctgc 3' (SEQ.ID.NO. 80)

Fig. 32 shows a schematic representation of the plasmid pMyHis6. The c-myc-tag sequence is used to detect the scFv fragment in an ELISA or a Western Blot analysis (Evan et al., Mol.Cell.Biol., 1985, 5(12), pp. 3610-6).

5 The His6-tag sequence was included to facilitate the purification of scFv fragments by metal ion chromatography (Hochuli et al., 1988. Biotechnology, 6:1321-1325). The plasmid contains the lacZ gene promoter (PlacZ) the PelB-leader sequence (see legend
10 Fig. 26) an E. coli origin of replication (colE1ori) and a M13 phage origin of replication (M13ori). To allow for specific selection, the plasmid also carries the gene for the enzyme β -lactamase (AmpR) mediating resistance against the antibiotic ampicillin.

15 The gene of the 198/B1 (clone AB2)-scFv was rescued from plasmid pDAP2-198AB2#100 (example 16) by digestion with SfiI and NotI and inserted into SfiI/NotI cleaved pMyHis6. The resultant plasmid was designated pMyHis-198AB2#102. Fig. 34 shows the nucleotide and amino acid
20 sequence of 198AB2 scFv (linked to the c-myc-tag and the His6- tag):the resulting ORF of the expression vector is named pMyHis6-198AB2#102. Vector pMyHis6 was constructed by cleaving vector pCOCK (Engelhardt O. et al, BioTechniques 17, 44-46, 1994) NotI - EcoRI and
25 inserting the following annealed oligonucleotides:
(5'-GGCCGCGAACAACAACTCATCTCAGAAGAGGATCTGAATGGG
GCGGCACATCACCATCACCATCACTAATAAG - 3' (SEQ.ID.No. 103)
and 5'- TTATTAGTGATGGTGATGGT
GATGTGCCGCCCATTCAGATCCTCTTCTGAGATGAGTTTTTGTTCCTGC-
30 3' (SEQ.ID.NO. 104)). The resultant vector, named pMyHis6, was cleaved SfiI - NotI and the gene of scFv 198AB2 was swapped into this vector from vector pDAP2-198AB2#100.

35 In analogy to the 198AB2 construct, the #8860 scFv fragment was cloned from a plasmid designated pDAP2-8860scFv clone 11. The pure scFv protein of #8860 was designated 8860-M/H#4c (plasmid p8860-M/H#4c, Fig. 35).

The scFv proteins were expressed in *E. coli* and affinity purified from bacterial supernatants on Protein L columns (see example 17). The final MycHis-198AB2#102 and 8860-M/H#4c preparations were dialyzed against 50mM imidazole, 100mM NaCl, pH7.4 and analyzed in a chromogenic FVIII assay as described above (Fig. 36).

As can be seen in Fig. 36, the scFv construct MycHis-198AB2#102 gave rise to a substantial FXa generation whereas the negative controls 8860-M/H#4c and plain reaction buffer (IZ) did not.

Fig. 36 demonstrates the chromogenic FVIII-like activity of the 198/B1 (subclone AB2) scFv fragment (MycHis-198AB2#102) in the presence of 2.3nM human FIXa. As a positive control we used 4.8pM FVIII whereas a unrelated scFv (8860-M/H#4c) and plain reaction buffer (IZ) served as negative controls.

Claims:

1. An antibody or antibody derivative against factor IX/factor IXa which increases the procoagulant activity of FIXa.
2. An antibody or antibody derivative according to claim 1, wherein said antibody or antibody derivative increases the procoagulant activity of FIXa in the presence of FVIII inhibitors.
3. An antibody according to any one of claim 1 wherein said antibody is selected from the group consisting of IgG, IgM, IgA and IgE antibodies.
4. An antibody or antibody derivative according to claim 1, wherein said antibody or antibody derivative is selected from the group consisting of monoclonal antibodies, antibody fragments, chimeric antibodies, humanized antibodies, single chain antibodies, bispecific antibodies, diabodies, and di-, oligo- or multimers thereof.
5. An antibody derivative according to claim 1, wherein said antibody derivative comprises a complement determining region (CDR) peptide.
6. An antibody derivative according to claim 5, wherein said CDR peptide is a CDR3 peptide.
7. An antibody derivative according to claim 6, wherein said CDR3 peptide comprises an amino acid sequence selected from the group consisting of:
Tyr-Gly-Asn-Ser-Pro-Lys-Gly-Phe-Ala-Tyr;
Cys-X-X-Tyr-Gly-Asn-Ser-Pro-Lys-Gly-Phe-Ala-Tyr-X-X-Cys,
wherein
X may be any desired amino acid;

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14. A hybridoma cell line expressing an antibody or antibody derivative against factor IX/factor IXa according to claim 1.

15. A hybridoma cell line according to claim 14,
wherein said cell line is selected from the group
consisting of #196/AF1, #196/AF2, #193/AD3, #193/K2-1,
#198/AC1/1, #198/AM1, #198/A1, #198/B1, #198/AP1,
5 198/A1, 198/B1, 198/BB1, 198/A1, 198/B1, 198/BB1.
16. An antibody or antibody derivative according to
claim 1, which is expressed by a hybridoma cell line
according to claim 14.
10
17. A DNA molecule, wherein said DNA molecule encodes
an antibody or an antibody derivative according to claim
1.
18. A pharmaceutical preparation comprising an
antibody or antibody derivative according to claim 1 and
a pharmaceutically acceptable carrier.
15
19. A preparation according to claim 18, additionally
20 comprising factor IXa α and/or factor IXa β .
20. A method for treating patients afflicted with
blood coagulation disorders comprising administering a
pharmaceutically effective amount of the preparation of
25 claim 18 to said patients.
21. The method of claim 20, wherein said blood
coagulation disorders are selected from the group
comprising hemophilia A and hemorrhagic diathesis.
30
22. The method of claim 21, additionally comprising
the step of selecting hemophilia inhibitor patients.
23. A method of obtaining an antibody or antibody
35 derivative which interacts with factor IX/factor IXa
and increases the procoagulant activity of Factor IXa,
comprising the steps of:

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- immunizing an immunocompetent mouse with an antigen selected from the group consisting of FIX, FIX α , FIX α β or fragments thereof,
- isolating spleen cells of the immunized mouse,
- 5 - producing hybridoma clones,
- screening the hybridoma cell supernatants for an increase in the procoagulant activity of Factor Ixa, isolating and purifying the antibodies or antibody derivatives from hybridoma cell supernatants which
- 10 exhibit an increase in the procoagulant activity of factor IXa.

24. Use of an antibody or antibody derivative according to claim 1 for increasing the amidolytic activity of
- 15 factor IXa.

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ABSTRACT

An antibody or antibody derivative against factor IX/activated factor IX (FIXa) which increases the procoagulant activity of FIXa.

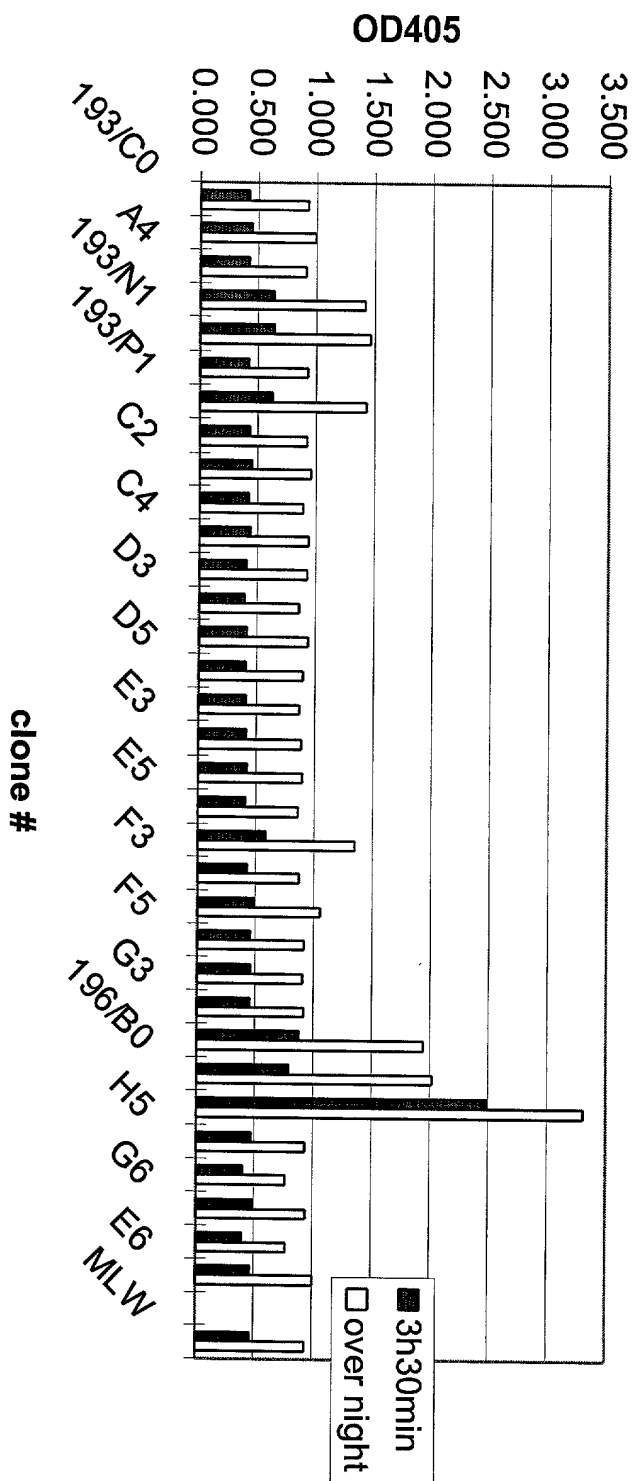


Fig. 1

09619209400

OD405

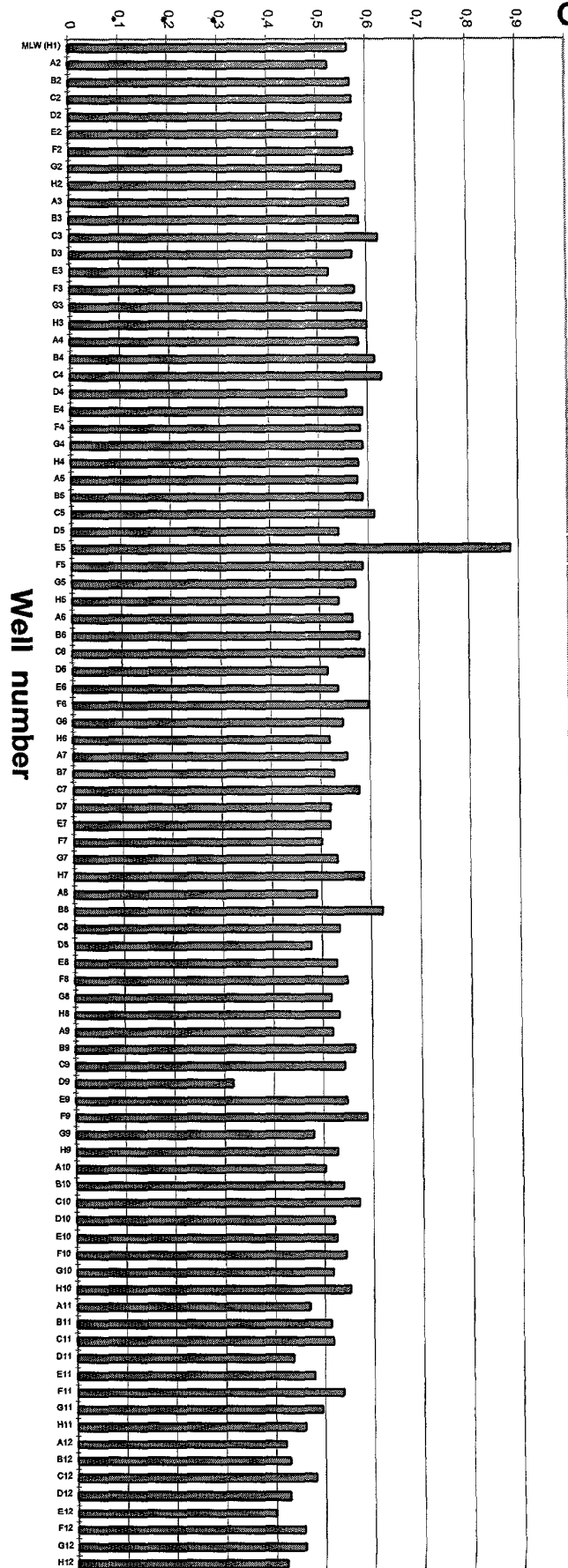


Fig. 2

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OD405

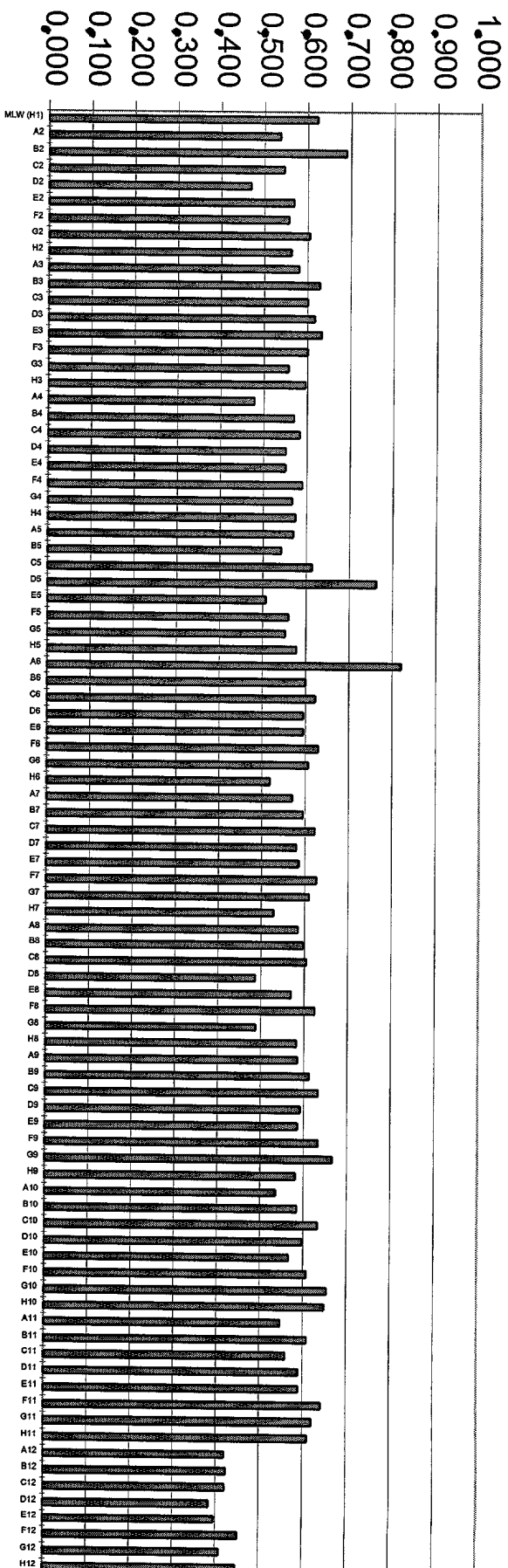


Fig. 3

Well number

09661992.09.4.00

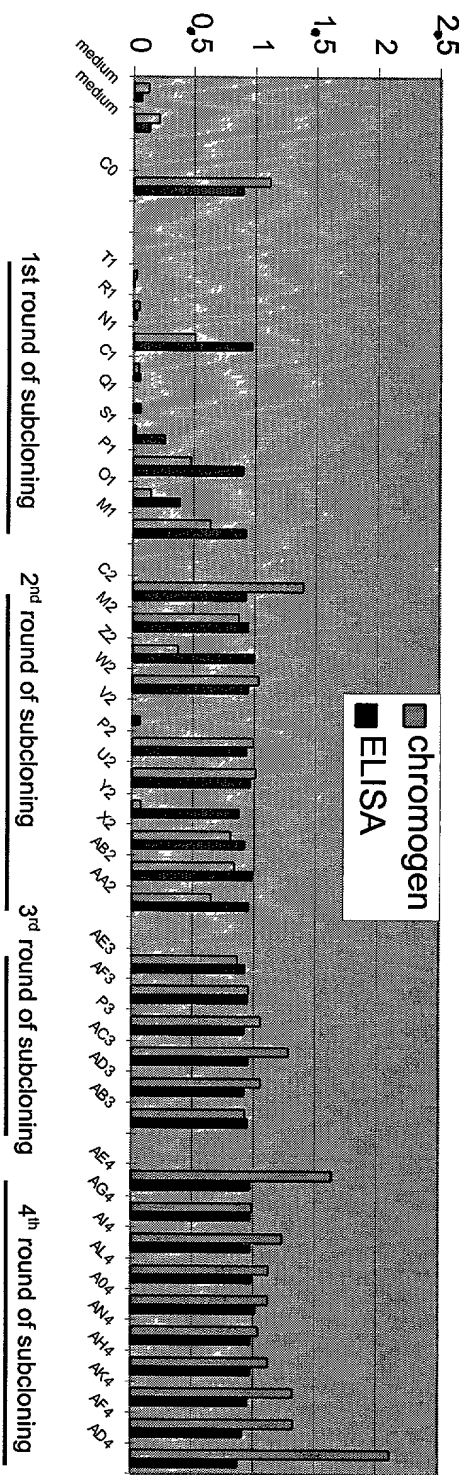


Fig. 4

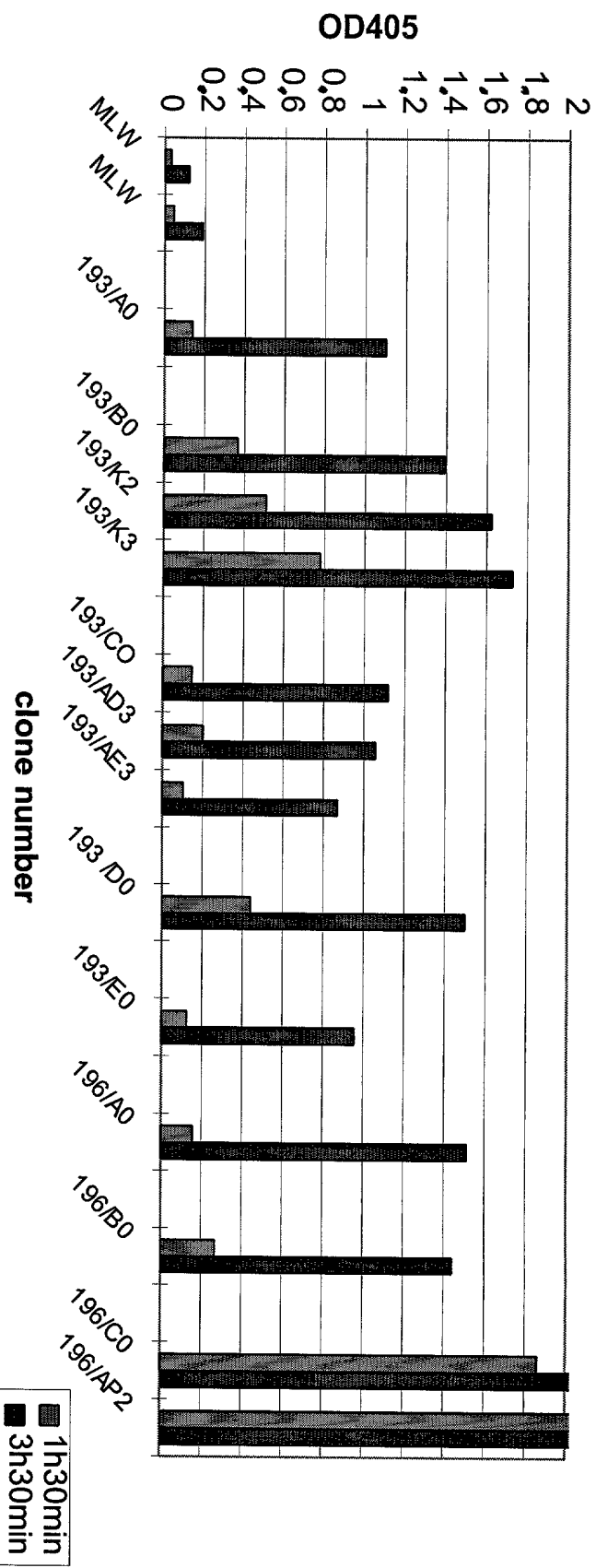


Fig. 5

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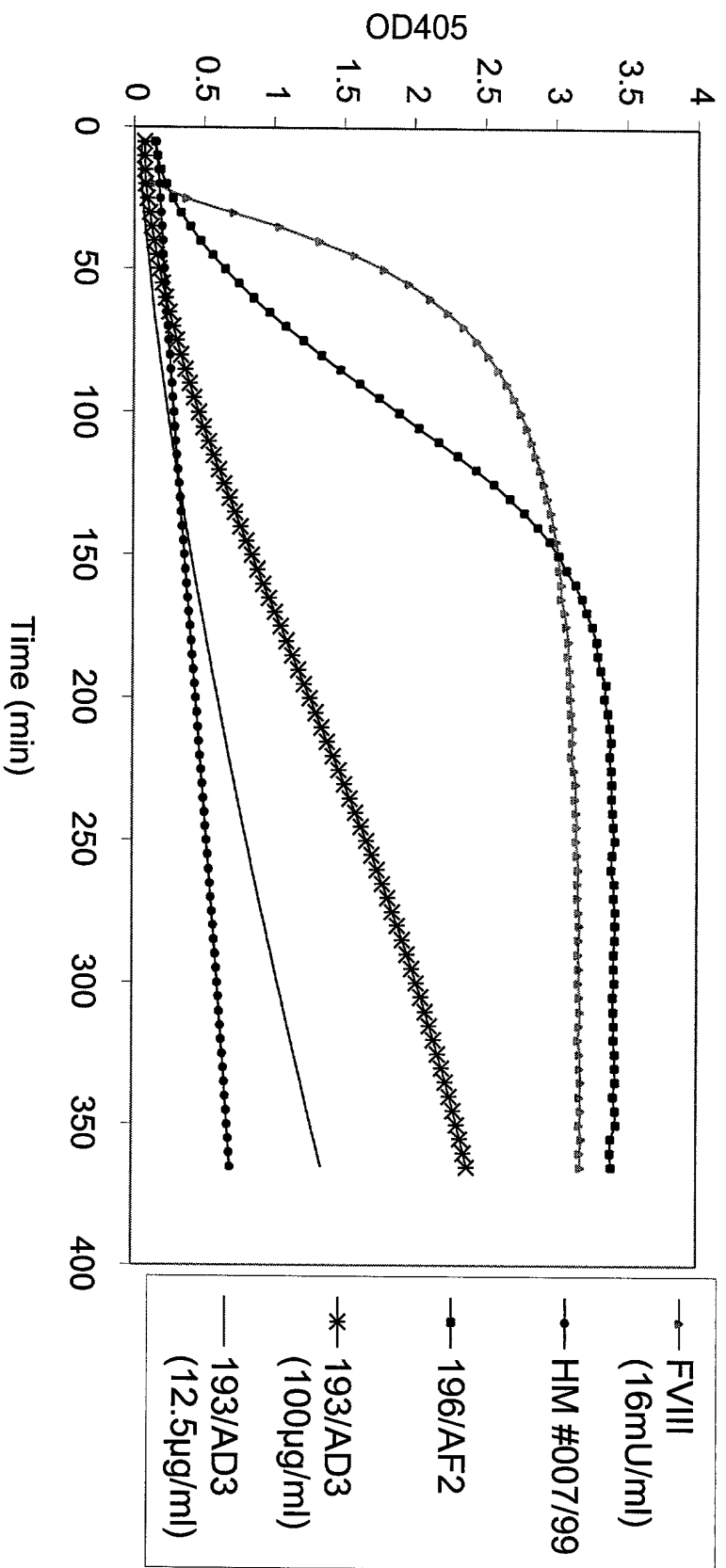


Fig. 6A

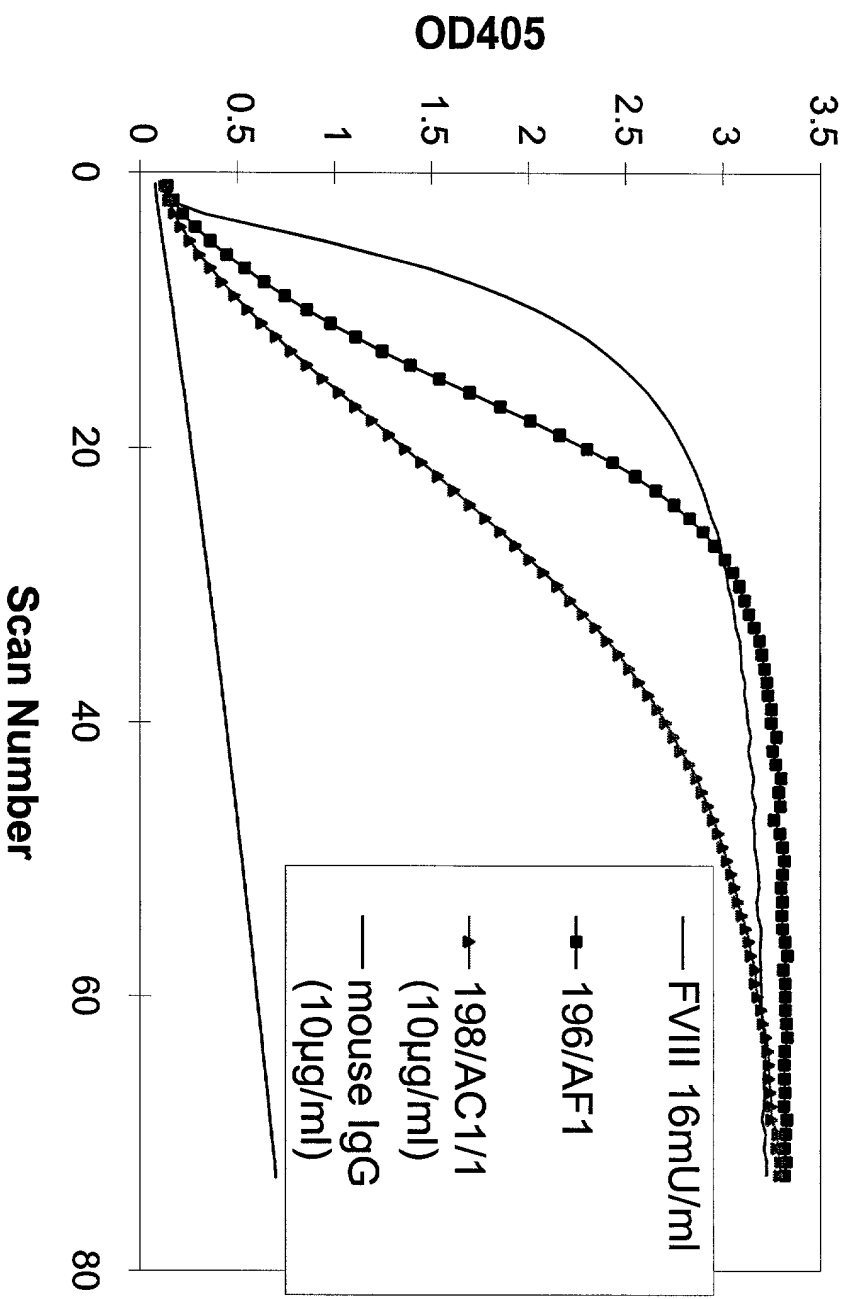


Fig. 6B

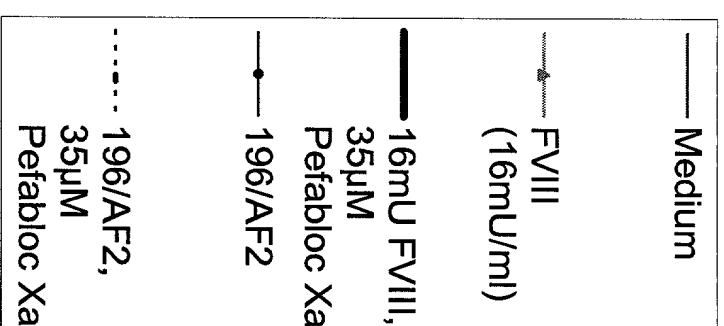
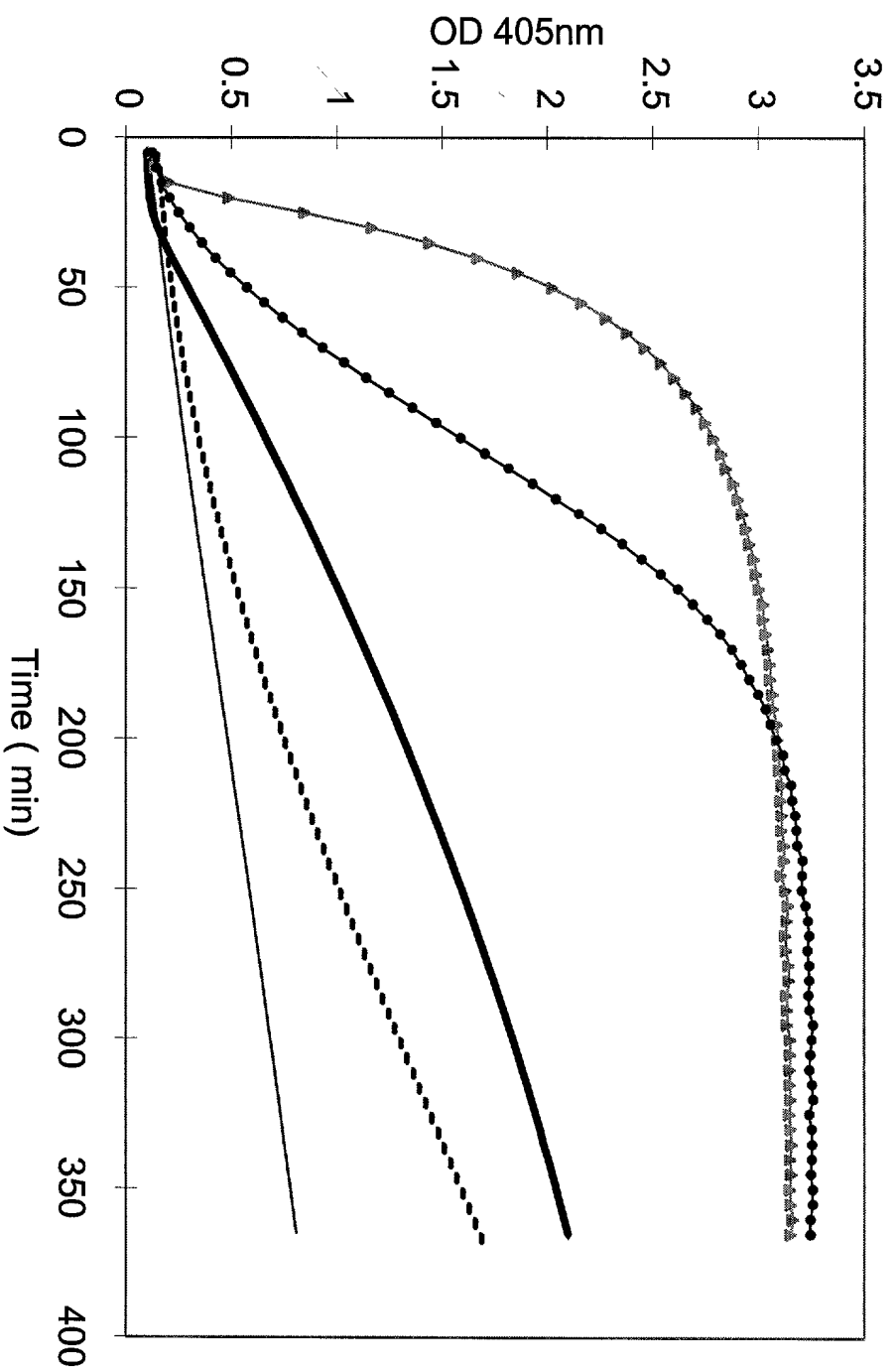


Fig. 7A

096192, 091400

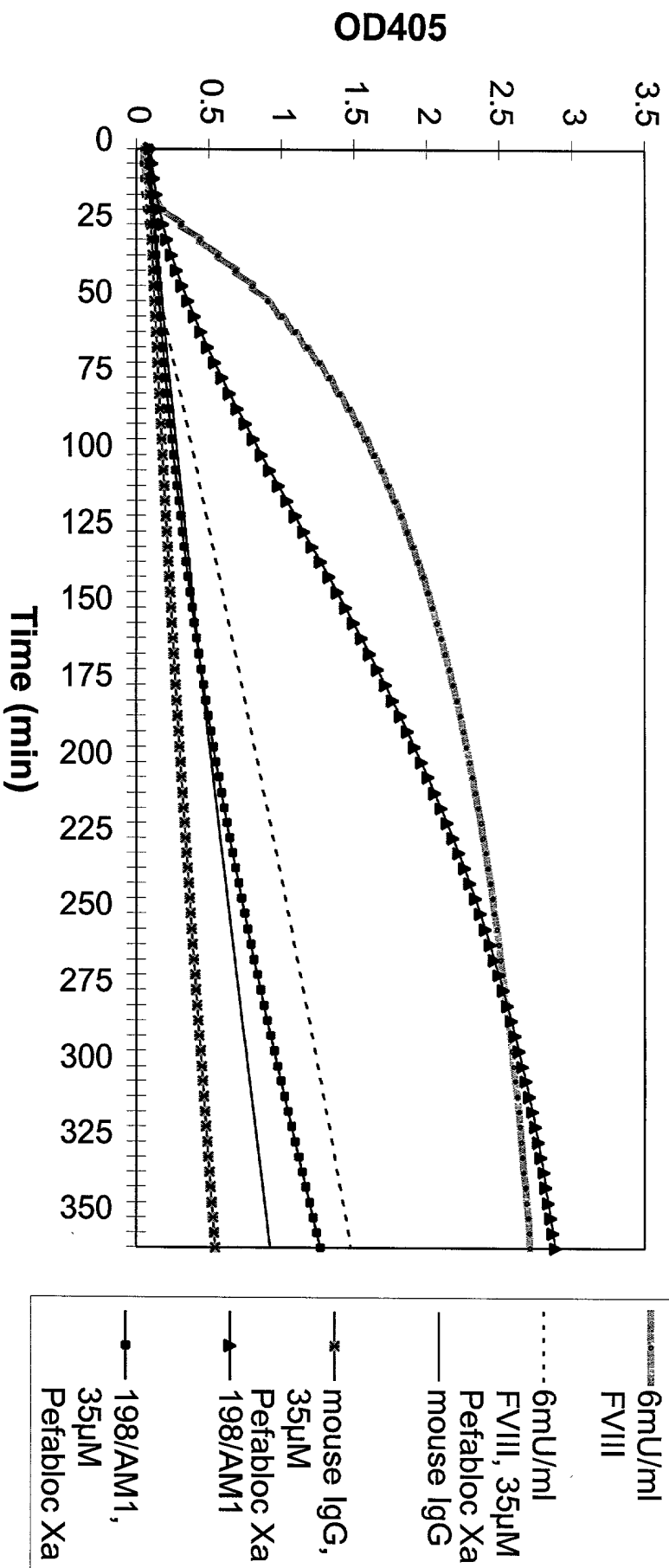


Fig. 7B

09661992.091400

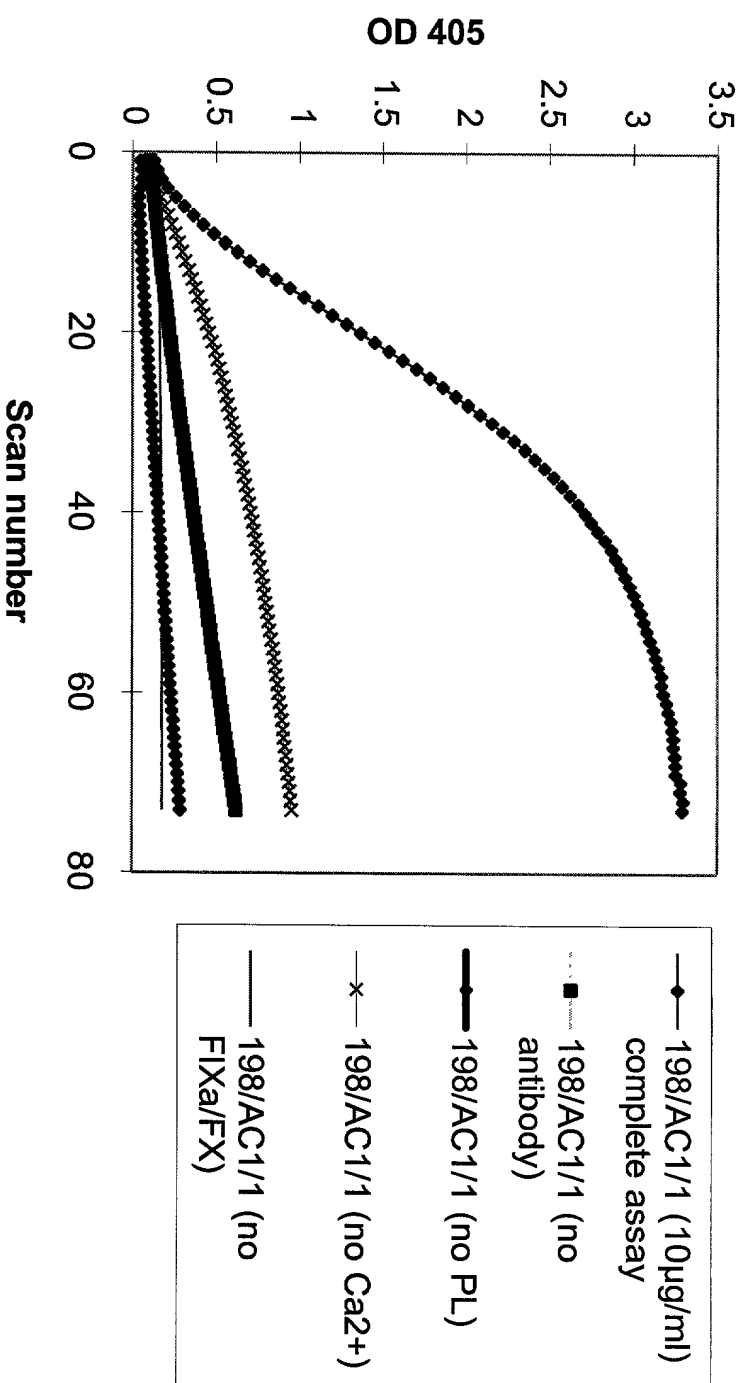


Fig. 8A

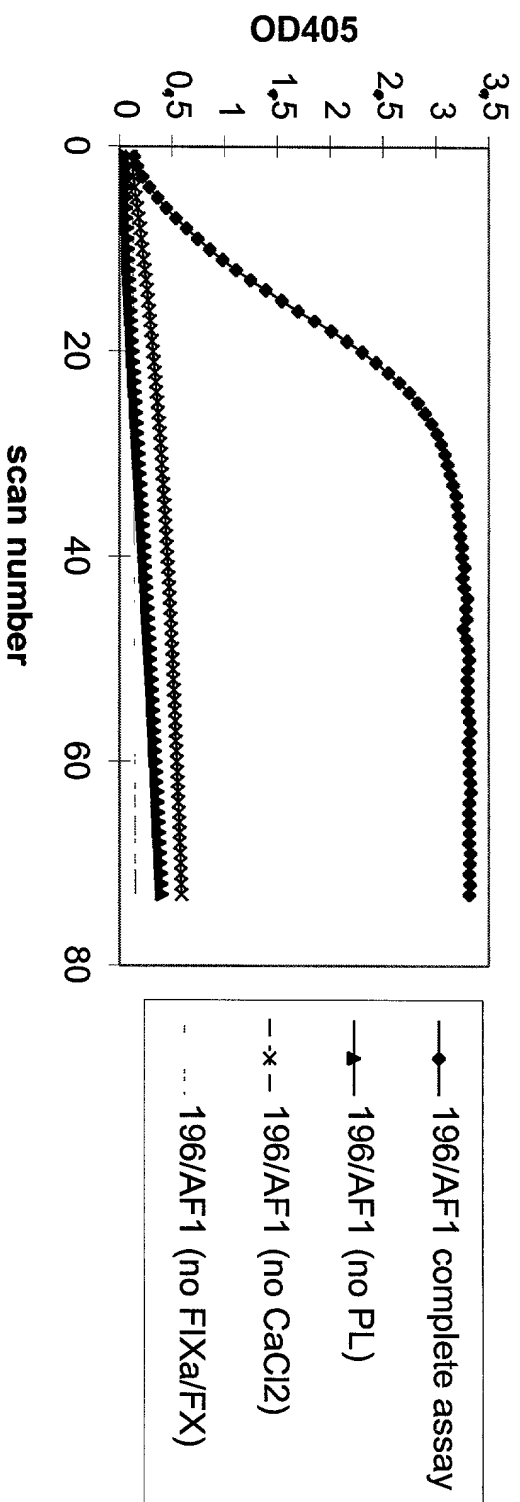


Fig. 8B

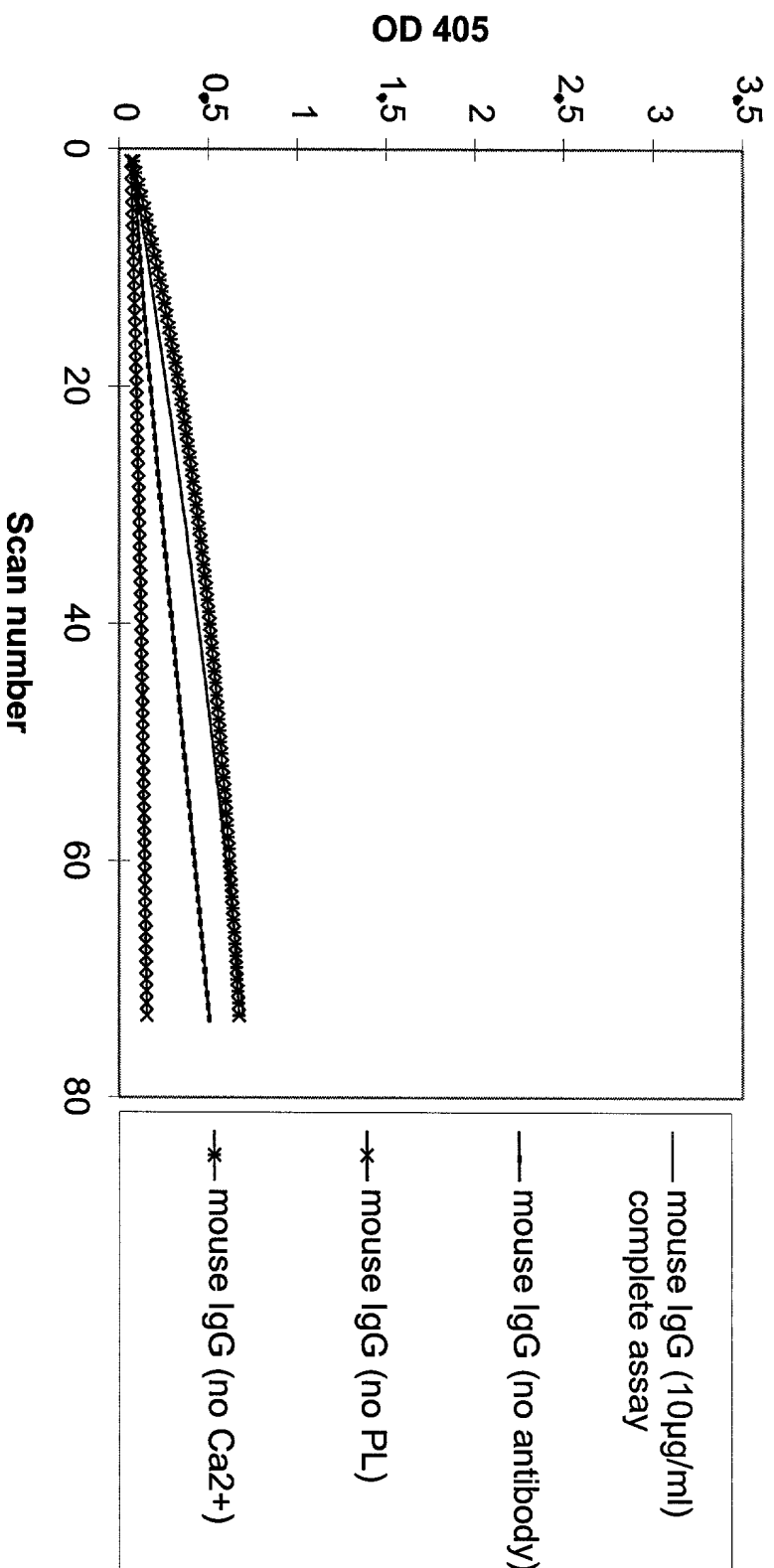


Fig. 8C

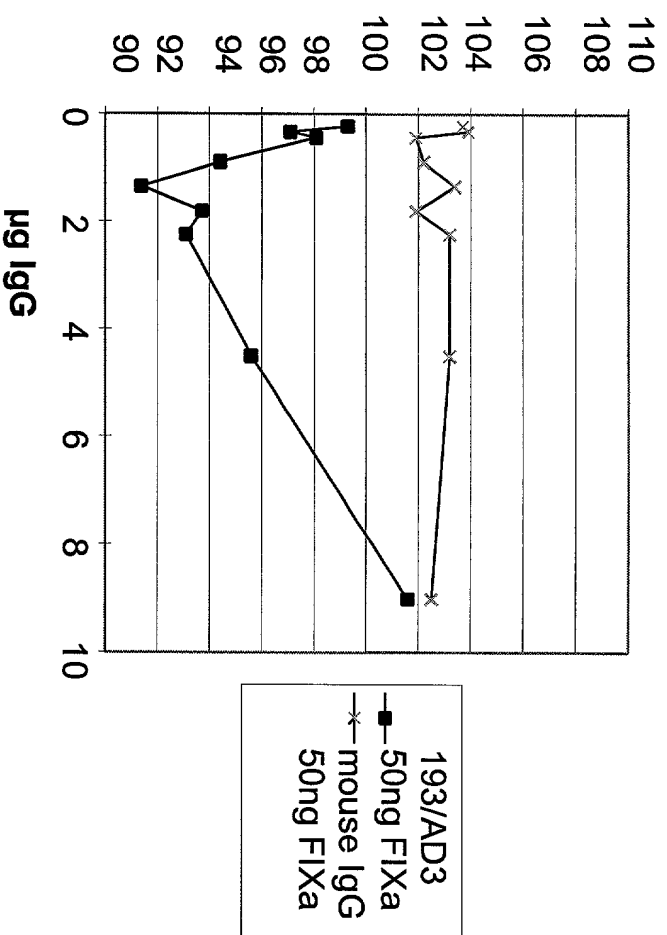


Fig.9

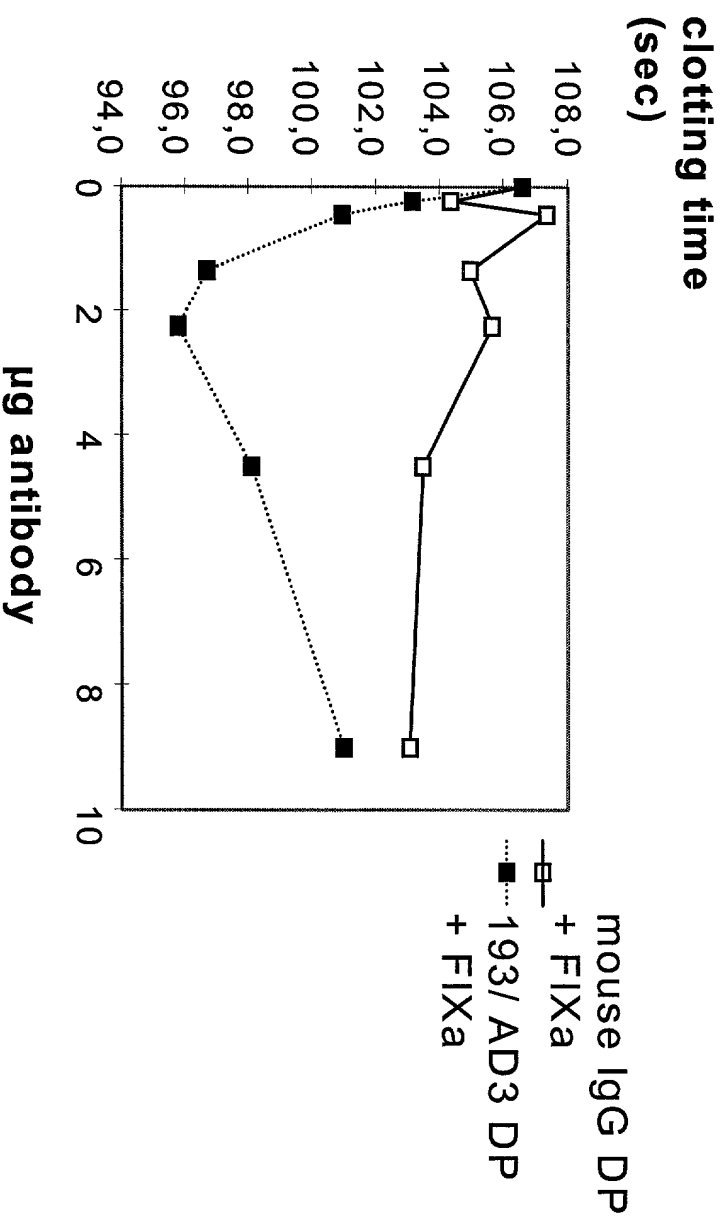


Fig. 10A

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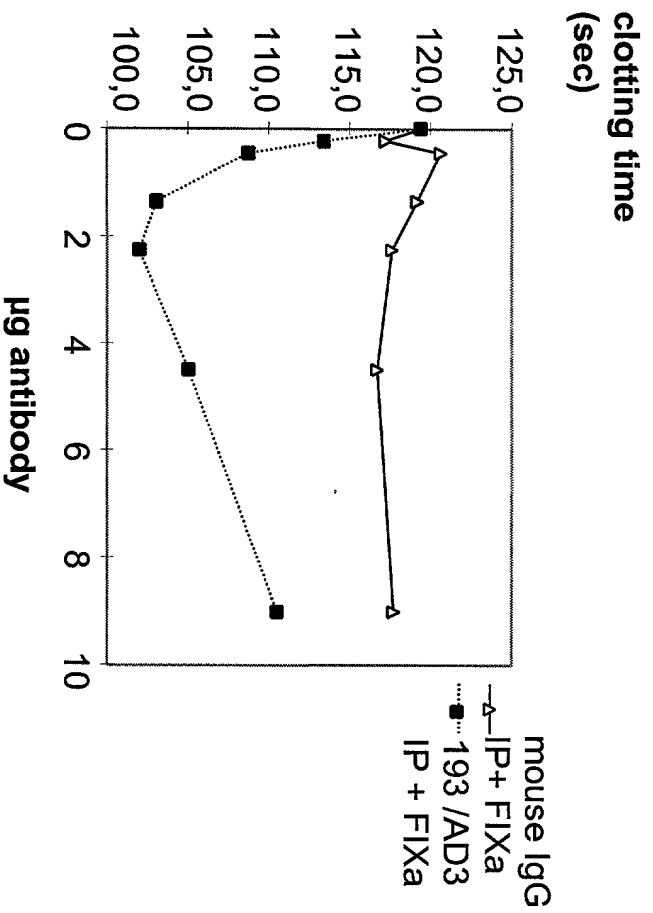


Fig. 10B

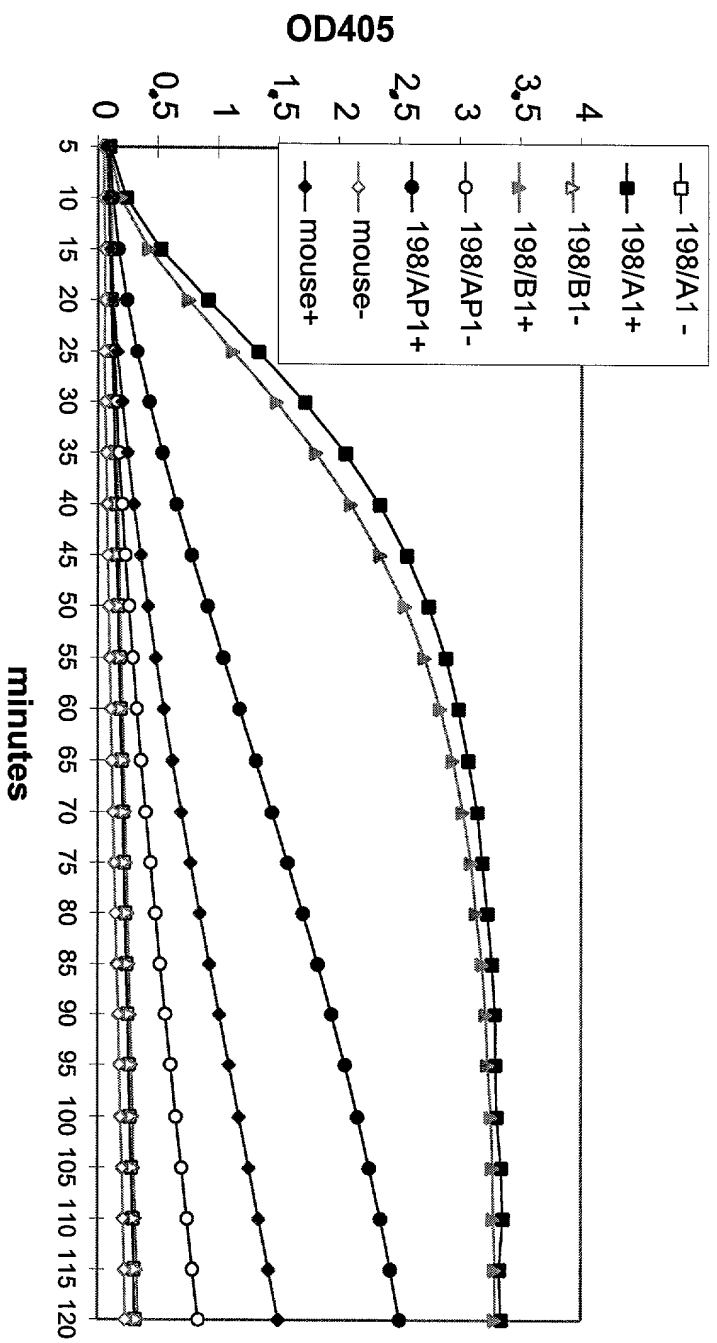


Fig.11

Mouse V_H back primers (containing SfiI-site):

VH1BACK-SfiI	5' C ATG CCA TGA CTC GCG GCC CAG CCG GCC ATG GCC SAG GTS MAR CTG CAG SAG TCM GG 3' (SEQ.ID.NO. 50)
VH1BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC GAG GTG CAG CTT CAG GAG TCA GG 3' (SEQ.ID.NO. 51)
VH2BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC GAT GTG CAG CTT CAG GAG TCR GG 3' (SEQ.ID.NO. 52)
VH3BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC CAG GTG CAG CTG AAG SAG TCA GG 3' (SEQ.ID.NO. 53)
VH4/6BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC GAG GTY CAG CTG CAR CAR TCT GG 3' (SEQ.ID.NO. 54)
VH5/9BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC CAG GTY CAR CTG CAG CAG YCT GG 3' (SEQ.ID.NO. 55)
VH7BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC GAR GTG AAG CTG GTG GAR TCT GG 3' (SEQ.ID.NO. 56)
VH8BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC GAG GTT CAG CTT CAG CAG TCT GG 3' (SEQ.ID.NO. 57)
VH10BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC GAA GTG CAG CTG KITG GAG WCT GG 3' (SEQ.ID.NO. 58)
VH11BACKSfi	5' GTC CTC GCA ACT GCG GCC CAG CCG GCC ATG GCC CAG ATC CAG TTG CTG CAG TCT GG 3' (SEQ.ID.NO. 59)

Mouse J_H forward primers (containing 1/2 linker-sequence and AscI-site):

VH1FOR2LiAsc	5' ACC GCC AGA GGC GCG CCC ACC TGA ACC GCC TCC ACC TGA GGA GAC GGT GAC CGT GGT CCC TTG GCC CC 3' (SEQ.ID.NO. 60)
JH1FORLiAsc	5' ACC GCC AGA GGC GCG CCC ACC TGA ACC GCC TCC ACC TGA GGA GAC GGT GAC CGT GGT CCC 3' (SEQ.ID.NO. 61)
JH2FORLiAsc	5' ACC GCC AGA GGC GCG CCC ACC TGA ACC GCC TCC ACC TGA GGA GAC TGT GAG AGT GGT GCC 3' (SEQ.ID.NO. 62)
JH3FORLiAsc	5' ACC GCC AGA GGC GCG CCC ACC TGA ACC GCC TCC ACC TGC AGA GAC AGT GAC CAG AGT CCC 3' (SEQ.ID.NO. 63)
JH4FORLiAsc	5' ACC GCC AGA GGC GCG CCC ACC TGA ACC GCC TCC ACC TGA GGA GAC GGT GAC TGA GGT TCC 3' (SEQ.ID.NO. 64)

IUPAC-Code: M=A/C, W=A/T, R=A/G, Y=C/T, S=C/G, K=G/T, H=A/C/T, D=A/G/T, V=A/C/G, B=T/C/G.

Fig. 12-2

09661992.091400

Primers for cloning mouse V_k genes

Mouse V_k back primers (containing *Asc*I-site and $\frac{1}{2}$ linker-sequence):

VK2BACK-LiAsci	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAC ATT GAG CTC ACC CAG TCT CCA 3' (SEQ.ID.NO. 65)
VK1BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAC ATT GTG ATG WCA CAG TCT CC 3' (SEQ.ID.NO. 66)
VK2BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAT GTT KITG ATG ACC CAA ACT CC 3' (SEQ.ID.NO. 67)
VK3BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAT ATT GTG ATR ACB CAG GCW GC 3' (SEQ.ID.NO. 68)
VK4BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAC ATT GTG CTG ACM CAR TCT CC 3' (SEQ.ID.NO. 69)
VK5BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG SAA AWT GTK CTC ACC CAG TCT CC 3' (SEQ.ID.NO. 70)
VK6BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAY ATY VWG ATG ACM CAG WCT CC 3' (SEQ.ID.NO. 71)
VK7BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG CAA ATT GTT CTC ACC CAG TCT CC 3' (SEQ.ID.NO. 72)
VK8BACKLi Asc	5' GGT TCA GAT GGG CGC GCC TCT GGC GGT GGC GGA TCG TCA TTA TTG CAG GTG CTT GTG GG 3' (SEQ.ID.NO. 73)

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Mouse J_K forward primers (containing NotI-site):

JK1NOT10	5'	GAG	TCA	TTC	TGC	GGC	CGC	CCG	TTT	GAT	TTC	CAG	CTT	GGT	GCC	3'
		(SEQ.ID.NO. 74)														
JK2NOT10	5'	GAG	TCA	TTC	TGC	GGC	CGC	CCG	TTT	TAT	TTC	CAG	CTT	GGT	CCC	3'
		(SEQ.ID.NO. 75)														
JK3NOT10	5'	GAG	TCA	TTC	TGC	GGC	CGC	CCG	TTT	TAT	TTC	CAG	TCT	GGT	CCC	3'
		(SEQ.ID.NO. 76)														
JK4NOT10	5'	GAG	TCA	TTC	TGC	GGC	CGC	CCG	TTT	TAT	TTC	CAA	CTT	TGT	CCC	3'
		(SEQ.ID.NO. 77)														
JK5NOT10	5'	GAG	TCA	TTC	TGC	GGC	CGC	CCG	TTT	CAG	CTC	CAG	CTT	GGT	CCC	3'
		(SEQ.ID.NO. 78)														

IUPAC-Code: K=G/T, M=A/C, W=A/T, R=A/G, Y=C/T, S=C/G, H=A/C/T, D=A/G/T, V=A/C/G, B=T/C/G.

Fig. 13-2

09561992 091400

VH

```

+1  E  V  K  L  V  E  S  G  P  E  L  K  K  P  G
1   GAG GTG AAG CTG GTG GAG TCT GGA CCT GAG CTG AAG AAG CCT GGA
+1  E  T  V  K  I  S  C  K  A  S  G  Y  I  F  T
46  GAG ACA GTC AAG ATC TCC TGC AAG GCT TCT GGG TAT ATC TTC ACA
+1  N  Y  G  M  N  W  V  K  Q  A  P  G  K  G  L
91  AAC TAT GGA ATG AAC TGG GTG AAG CAG GCT CCA GGA AAG GGT TTA
+1  K  W  M  G  W  I  N  T  Y  T  G  E  P  T  Y
136 AAG TGG ATG GGC TGG ATA AAC ACC TAC ACT GGA GAG CCA ACA TAT
+1  A  D  D  F  K  G  R  F  A  F  S  L  E  T  S
181 GCT GAT GAC TTC AAG GGA CGG TTT GCC TTC TCT TTG GAA ACC TCT
+1  A  S  T  A  Y  L  Q  I  N  N  L  K  N  E  D
226 GCC AGC ACT GCC TAT TTG CAG ATC AAC AAC CTC AAA AAT GAG GAC
+1  T  A  T  Y  F  C  A  L  Y  G  N  S  P  K  G
271 ACG GCT ACA TAT TTC TGT GCA TTA TAT GGT AAC TCC CCT AAG GGG

+1  F  A  Y  W  G  Q  G  T  L  V  T  V  S  A  G
316 TTT GCT TAC TGG GGC CAA GGG ACT CTG GTC ACT GTC TCT GCA GGT

                                linker
+1  G  G  G  S  G  G  R  A  S  G  G  G  S  D
361 GGA GGC GGT TCA GGT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAT

                                VL
+1  I  Q  M  T  Q  S  P  K  F  L  L  V  S  A  G
406 ATT CAG ATG ACA CAG TCT CCC AAA TTC CTG CTT GTA TCA GCA GGA

```

+1	D	R	V	T	I	T	C	K	A	S	Q	S	V	S	N
451	GAC	AGG	GTT	ACC	ATA	ACC	TGC	AAG	GCC	AGT	CAG	AGT	GTG	AGT	AAT
+1	D	V	A	W	Y	Q	Q	K	P	G	Q	S	P	K	L
496	GAT	GTA	GCT	TGG	TAC	CAA	CAG	AAG	CCG	GGG	CAG	TCT	CCT	AAA	CTA
+1	L	M	Y	Y	A	S	N	R	Y	T	G	V	P	D	R
541	CTG	ATG	TAC	TAT	GCA	TCC	AAT	CGC	TAC	ACT	GGA	GTC	CCT	GAT	CGC
+1	F	T	G	S	G	Y	G	T	D	F	T	F	T	I	S
586	TTC	ACT	GGC	AGT	GGA	TAT	GGG	ACG	GAT	TTC	ACT	TTC	ACC	ATC	AGC
+1	T	V	Q	A	E	D	L	A	V	Y	F	C	Q	Q	D
631	ACT	GTG	CAG	GCT	GAA	GAC	CTG	GCA	GTT	TAT	TTC	TGT	CAG	CAG	GAT
+1	Y	G	S	P	P	T	F	G	G	T	K	L	E	I	
676	TAT	GGC	TCT	CCT	CCC	ACG	TTC	GGA	GGG	GGC	ACC	AAG	CTG	GAA	ATT
+1	K	R													
721	AAA	CGG													

Fig. 14-2

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VH

+1 E V Q L V E S G G G L V K P G
 1 GAA GTG CAG CTG GTG GAG TCT GGG GGA GGC CTA GTG AAG CCT GGA

+1 G S L K L S C A A S G F T F S
 46 GGG TCC CTG AAA CTC TCC TGT GCA GCC TCT GGA TTC ACT TTC AGT

+1 T Y T M S W V R Q T P E K R L
 91 ACC TAT ACC ATG ATG TCT TGG GTT CGC CAG ACT CCG GAG AAG AGG CTG

+1 E W V A T I S S G G S Y T Y Y
 136 GAG TGG GTC GCA ACC ATT AGT AGT GGT GGT AGT TAC ACC TAC TAT

+1 P D S V R G R F T I S R D N A
 181 CCA GAC AGT GTG AGG GGC CGA TTC ACC ATC TCC AGA GAC AAT GCC

+1 K N T L Y L Q M S S L K S E D
 226 AAG AAC ACC CTG TAC CTG CAA ATG AGC AGT CTG AAG TCT GAG GAC

+1 T A M Y Y C T R D G G H G Y G
 271 ACA GCC ATG TAT TAC TGT ACA AGA GAT GGG GGA CAC GGG TAC GGT

+1 S S F D Y W G Q G T T L T V S
 316 AGT AGC TTT GAC TAC TGG GGC CAA GGC ACC ACT CTC ACA GTC TCC

linker

+1 S G G G G S G G R A S G G G
 361 TCA GGT GGA GGC GGT TCA GGT GGG CGC GCC TCT GGC GGT GGC GGA

VL

+1 S Q I V L T Q S P L S L P V S
 406 TCG CAA ATT GTG CTC ACC CAG TCT CCA CTC TCC CTG CCT GTC AGT

FIG 15-1

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+1	L	G	D	Q	A	S	I	S	C	R	S	S	Q	S	I
451	CTT	GGA	GAT	CAA	GCC	TCC	ATC	TCT	TGC	AGA	TCT	AGT	CAG	AGC	ATT
+1	V	H	S	N	G	N	T	Y	L	E	W	Y	L	Q	K
496	GTA	CAT	AGT	AAT	GGA	AAC	ACC	TAT	TTA	GAA	TGG	TAC	CTG	CAG	AAA
+1	P	G	Q	S	P	K	L	L	I	Y	K	V	S	N	R
541	CCA	GGC	CAG	TCT	CCA	AAG	CTC	CTG	ATC	TAC	AAA	GTT	TCC	AAC	CGA
+1	F	S	G	V	P	D	K	F	S	G	S	G	S	G	T
586	TTT	TCT	GGG	GTG	CCA	GAC	AAA	TTC	AGT	GGC	AGT	GGA	TCA	GGG	ACA
+1	D	F	T	L	K	I	S	R	V	E	A	E	D	L	G
631	GAT	TTC	ACA	CTC	AAG	ATC	AGC	AGA	GTG	GAG	GCT	GAG	GAT	CTG	GGA
+1	V	Y	Y	C	F	Q	G	S	H	V	P	W	T	F	G
676	GTT	TAT	TAC	TGC	TTT	CAA	GGT	TCA	CAT	GTT	CCG	TGG	ACG	TTC	GGT
1	G	G	T	K	L	E	I	K	R						
721	GGA	GGC	ACC	AAG	CTG	GAA	ATC	AAA	CGG						

Fig. 15 - 2

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+1 E V Q L Q E S G G L V K P G
 1 GAG GTG CAG CTT CAG GAG TCA GGG GGA GGC TTA GTG AAG CCT GGA
 +1 G S L K L S C A A S G F T F S
 46 GGG TCC CTG AAA CTC TCC TGT GCA GCC TCT GGA TTC ACT TTC AGT
 +1 S Y T M S W V R Q T P E K R L
 91 AGC TAT ACC ATG TCT TGG GTT CGC CAG ACT CCG GAG AAG AGG CTG
 +1 E W V A T I S S G G S S T Y Y
 136 GAG TGG GTC GCA ACC ATT AGT AGT GGT GGT AGT TCC ACC TAC TAT
 +1 P D S V K G R F T I S R D N A
 181 CCA GAC AGT GTG AAG GGC CGA TTC ACC ATC TCC AGA GAC AAT GCC
 +1 K N T L Y L Q M S S L R S E D
 226 AAG AAC ACC CTG TAC CTG CAA ATG AGC AGT CTG AGG TCT GAG GAC
 +1 T A M Y Y C T R E G G G F T V
 271 ACA GCC ATG TAT TAC TGT ACA AGA GAG GGG GGT GGT TTC ACC GTC
 +1 N W Y F D V W G A G T L V T V
 316 AAC TGG TAC TTC GAT GTC TGG GGC GCA GGG ACT CTG GTC ACT GTC
 linker
 +1 S A G G G G S G G R A S G G
 361 TCT TCA GGT GGA GGC GGT TCA GGT GGG CGC GCC TCT GGC GGT GGC
 VL
 +1 G S E N V L T Q S P A S L A V
 406 GGA TCG GAA AAT GTG CTC ACC CAG TCT CCA GCT TCT TTG GCT GTG

F1616-1

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+1 S L G Q R A T I S C R A S E S
 451 TCT CTA GGG CAG AGG GCC ACC ATA TCC TGC AGA GCC AGT GAA AGT
 +1 V D S Y G Y N F M H W Y Q Q I
 496 GTT GAT AGT TAT TAT GGC TAT AAT TTT ATG CAC TGG TAT CAG CAG ATA
 +1 P G Q P P K L L I Y R A S N L
 541 CCA GGA CAG CCA CCC AAA CTC CTC ATC TAT CGT GCA TCC AAC CTA
 +1 E S G I P A R F S G S G S R T
 586 GAG TCT GGG ATC CCT GCC AGG TTC AGT GGC AGT GGG TCT AGG ACA
 +1 D F T L T I N P V E A D D V A
 631 GAC TTC ACC CTC ACC AAT AAT CCT GTG GAG GCT GAT GAT GTT GCA
 +1 T Y Y C Q Q Q S N E D P L T F G
 676 ACC TAT TAC TGT CAG CAA AGT AAT GAG GAT CCG CTC ACG TTC GGT
 +1 T G T R L E I K R
 721 ACT GGG ACC AGA CTG GAA ATA AAA CGG

Fig.16 - 2

09661992.091400

VH

+1 E V Q L Q E S G G L V K P G G S L K L
 1 GAG GTG CAG CTT CAG GAG TCA GGG GGA GGC TTA GTG AAG CCT GGA GGG TCC CTG AAA CTC
 CTC CAC GTC GAA GTC CTC AGT CCC CCT CCG AAT CAC TTC GGA CCT CCC AGG GAC TTT GAG

+1 S C A A S G F I F S S Y T M S W V R Q T
 61 TCC TGT GCA GCC TCT GGA TTC ATT TTT AGT AGT TAT ACC ATG TCT TGG GTT CGC CAG ACT
 AGG ACA CGT CGG AGA CCT AAG TAA AAA TCA TCA ATA TGG TAC AGA ACC CAA GCG GTC TGA

+1 P E K R L E W V A T I S S G G S S T Y Y
 121 CCG GAG AAG AGG CTG GAG TGG GTC GCA ACC ATT AGT AGT AGT GGT GGT AGT TCC ACC TAC TAT
 GGC CTC TTC TCC GAC CTC ACC CAG CGT TGG TAA TCA TCA CCA CCA TCA AGG TGG ATG ATA

+1 P D S V K G R F T I S R D N A K N T L Y
 181 CCA GAC AGT GTG AAG GGC CGA TTC ACC ATC TCC AGA GAC AAT GCC AAG AAC ACC CTG TAC
 GGT CTG TCA CAC TTC CCG GCT AAG TGG TAG AAG TCT CTG TTA CCG TTC TTG TGG GAC ATG

+1 L Q M S S L K S E D T A M Y H C T R E G
 241 CTG CAA ATG AGC AGT CTG AAG TCT GAG GAC ACA GCC ATG TAT CAC TGT ACA AGA GAG GGG
 GAC GTT TAC TCG TCA GAC TTC AGA CTC CTG TGT CGG TAC ATA GTG ACA TGT TCT CTC CCC

+1 G G Y Y V N W Y F D V W G A G T T L T V
 301 GGT GGT TAT TAC GTC AAC TGG TAC TTC GAT GTC TGG GGC GCA GGC ACC ACT CTC ACA GTC
 CCA CCA ATA ATG CAG TTG ACC ATG AAG CTA CAG ACC CCG CGT CCG TGG TGA GAG TGT CAG

linker

VL

+1 S S G G G G S G G R A S G G G G S D I E
 361 TCC TCA GGT GGA GGC GGT TCA GGT GGG CGC GCC TCT GGC GGT GGC GGA TCG GAC ATT GAG
 AGG AGT CCA CCT CCG CCA AGT CCA CCC GCG CGG AGA CCG CCA CCG CCT AGC CTG TAA CTC

+1 L T Q S P A S L A V S L G Q R A T I S C
 421 CTC ACN CAG TCT CCA GCT TCT TTG GCT GTG TCT CTA GGG CAG AGG GCC ACC ATA TCC TGC
 GAG TGN GTC AGA GGT CGA AGA AAC CGA CAC AGA GAT CCC GTC TCC CGG TGG TAT AGG ACG

FIG 17-1

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+1	R	A	S	E	S	V	D	S	Y	G	K	S	F	M	H	W	Y	Q	Q	K
481	AGA	GCC	AGT	GAA	AGT	GTT	GAT	AGT	TAT	GGC	AAG	AGT	TTT	ATG	CAC	TGG	TAC	CAG	CAG	AAA
	TCT	CGG	TCA	CTT	TCA	CAA	CTA	TCA	ATA	CCG	TTC	TCA	AAA	TAC	GTG	ACC	ATG	GTC	GTC	TTT
+1	P	G	Q	P	P	K	L	L	I	Y	R	A	S	N	L	E	S	G	I	P
541	CCA	GGG	CAG	CCA	CCC	AAA	CTC	CTC	ATC	TAT	CGT	GCA	TCC	AAC	CTA	GAA	TCT	GGG	ATC	CCT
	GGT	CCC	GTC	GGT	GGG	TTT	GAG	GAG	TAG	ATA	GCA	CGT	AGG	TTG	GAT	CTT	AGA	CCC	TAG	GGA
+1	A	R	F	S	G	S	G	S	R	T	D	F	T	L	T	I	N	P	V	E
601	GCC	AGG	TTC	AGT	GGC	AGT	GGG	TCT	AGG	ACA	GAC	TTC	ACC	CTC	ACC	ATT	AAT	CCT	GTG	GAG
	CGG	TCC	AAG	TCA	CCG	TCA	CCC	AGA	TCC	TGT	CTG	AAG	TGG	GAG	TGG	TAA	TTA	GGA	CAC	CTC
+1	A	D	D	V	A	T	Y	Y	Y	C	Q	Q	S	N	E	D	P	L	T	F
661	GCT	GAT	GAT	GTT	GCN	ACC	TAT	TAC	TGT	CAG	CAA	AGT	AAT	GAG	GAT	CCC	CTC	ACG	TTC	GGT
	CGA	CTA	CTA	CAA	CGN	TGG	ATA	ATG	ACA	GTC	GTT	TCA	TTA	CTC	CTA	GGG	GAG	TGC	AAG	CCA
+1	A	G	T	R	L	E	I	K	R											
721	GCT	GGG	ACC	AGA	CTG	GAA	ATA	AAA	CGG											
	CGA	CCC	TGG	TCT	GAC	CTT	TAT	TTT	GCC											

Fig. 17-2

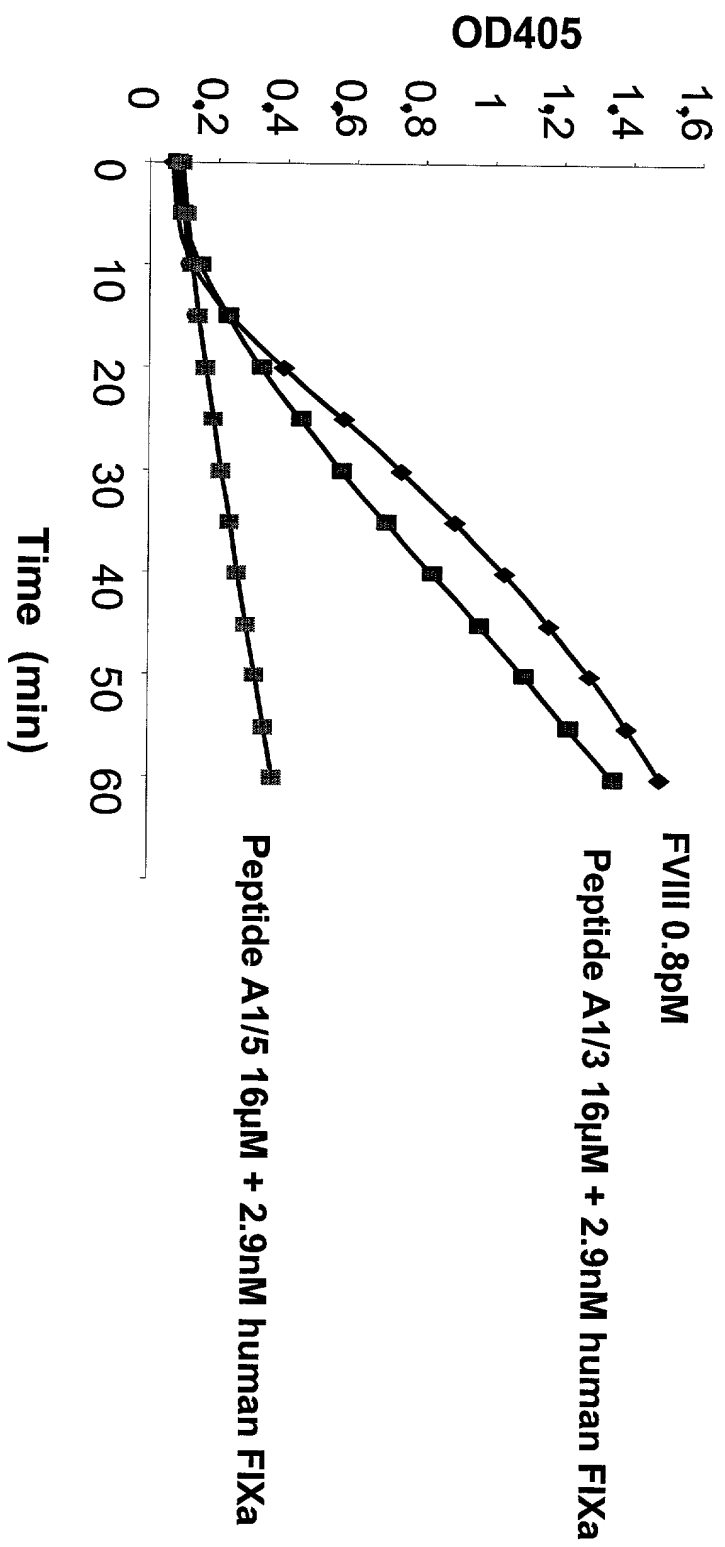


Fig. 18

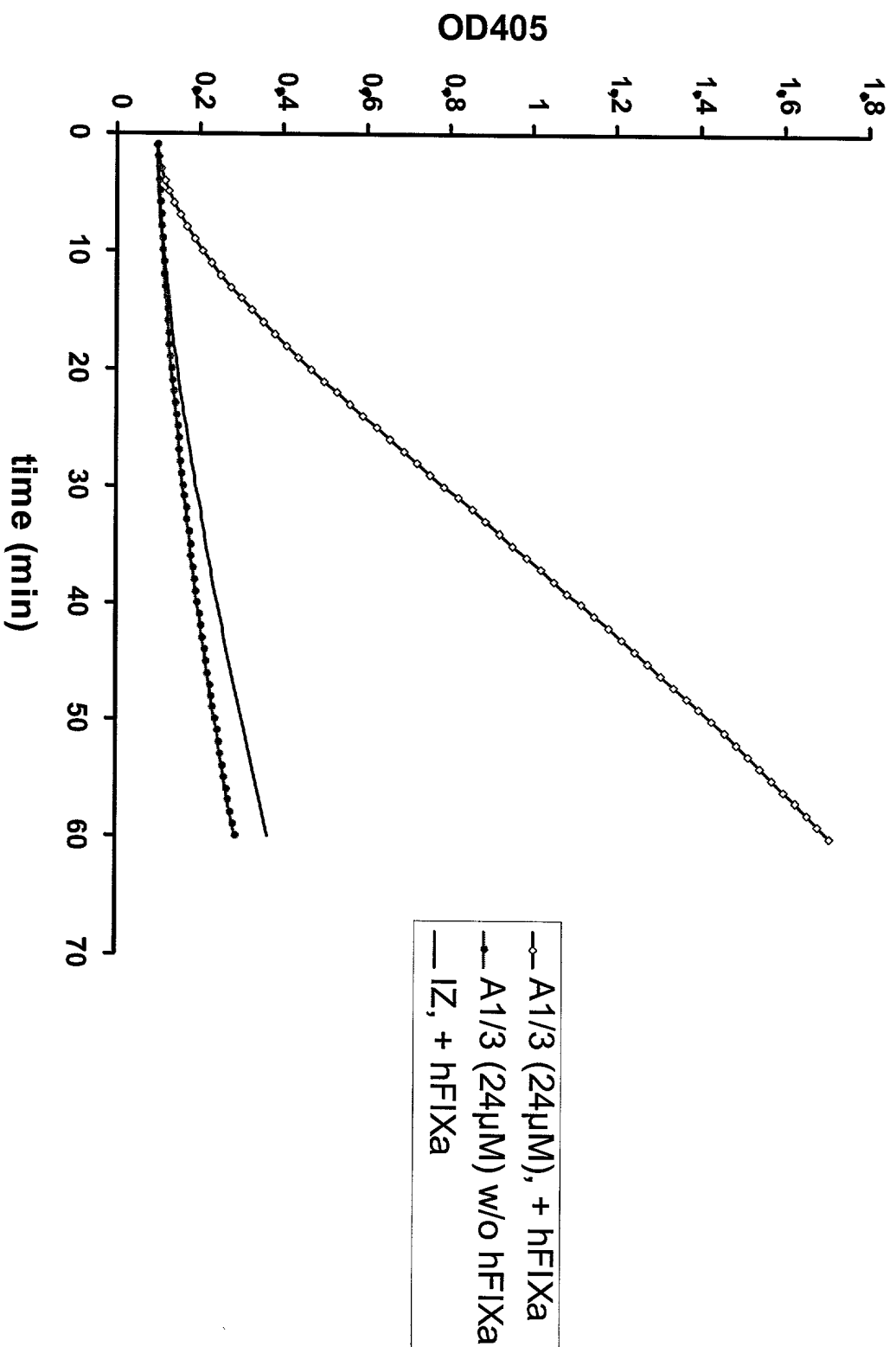


Fig. 19

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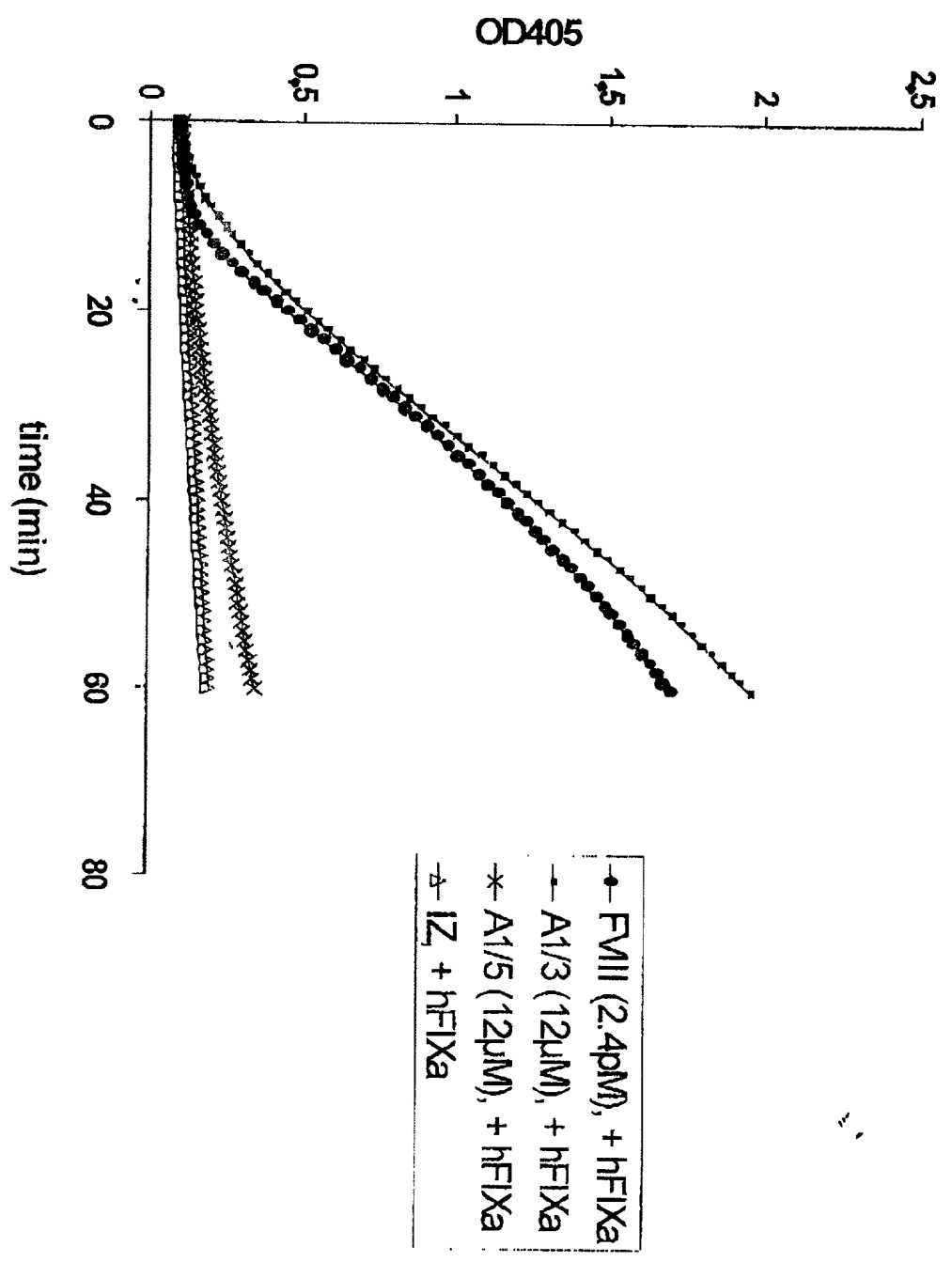


Fig. 20

09661992.091400

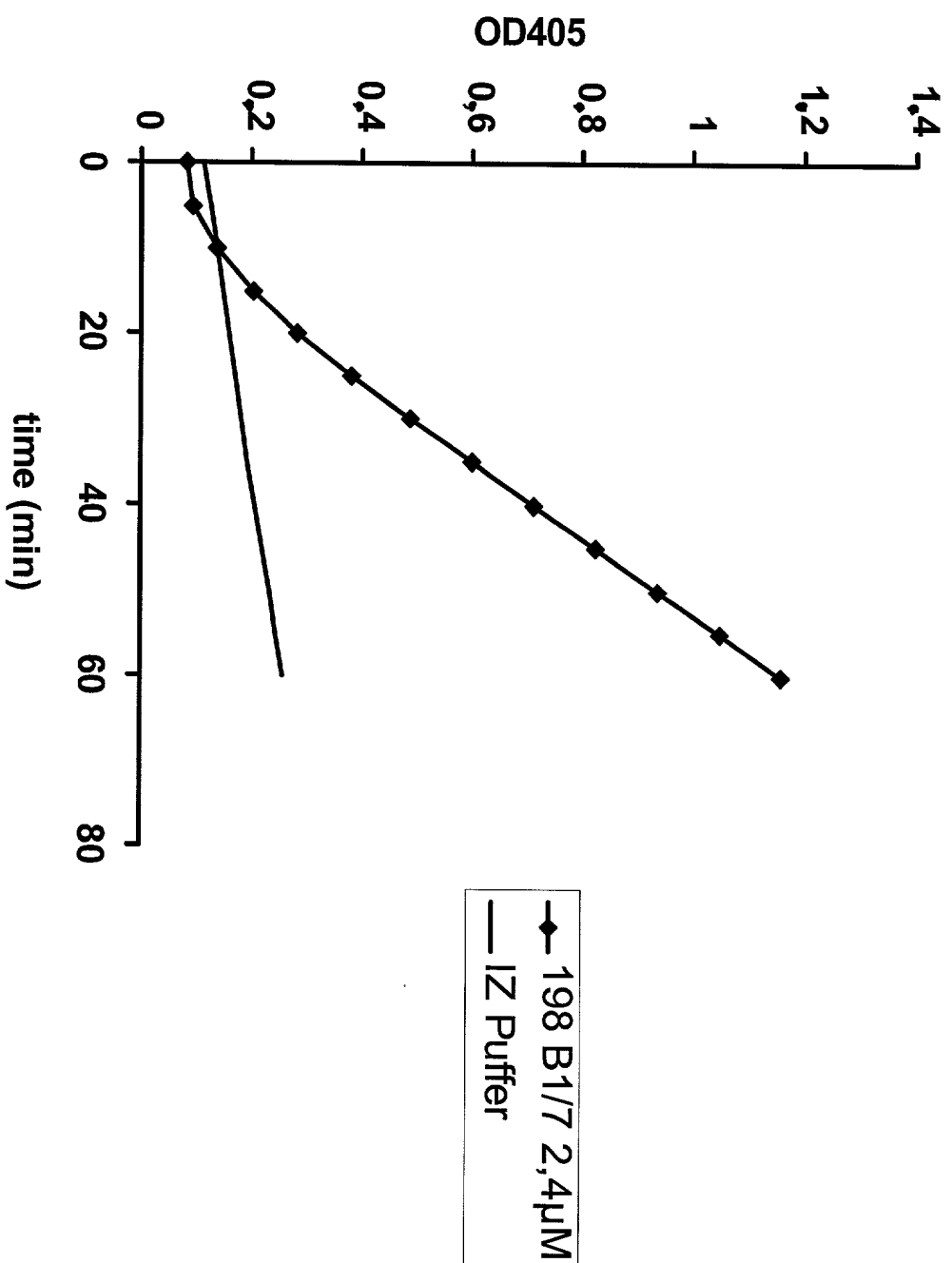


Fig. 21

09661992.091400

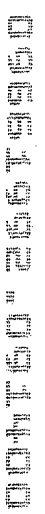


Fig. 22

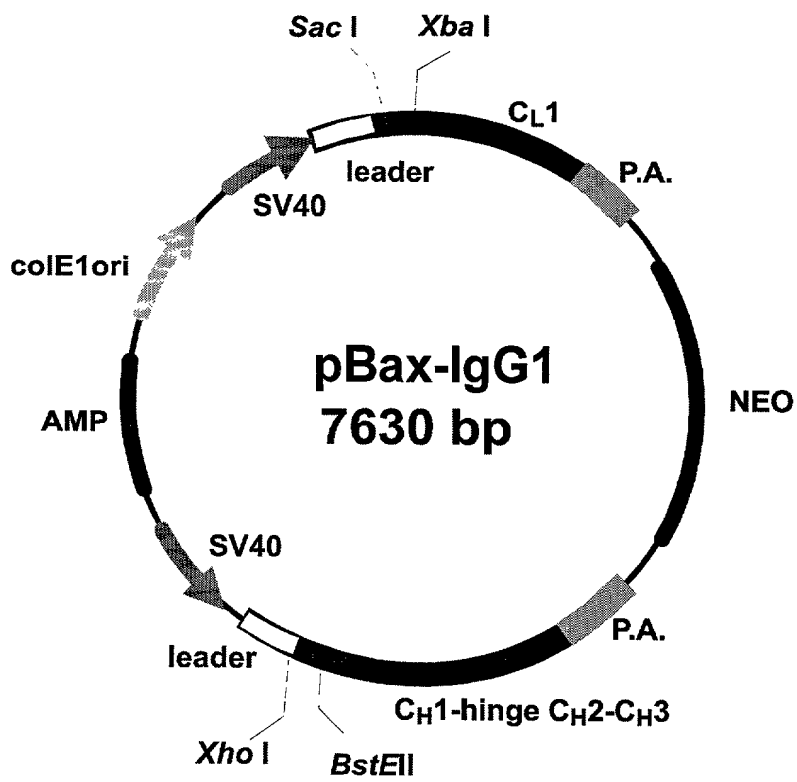


Figure 23

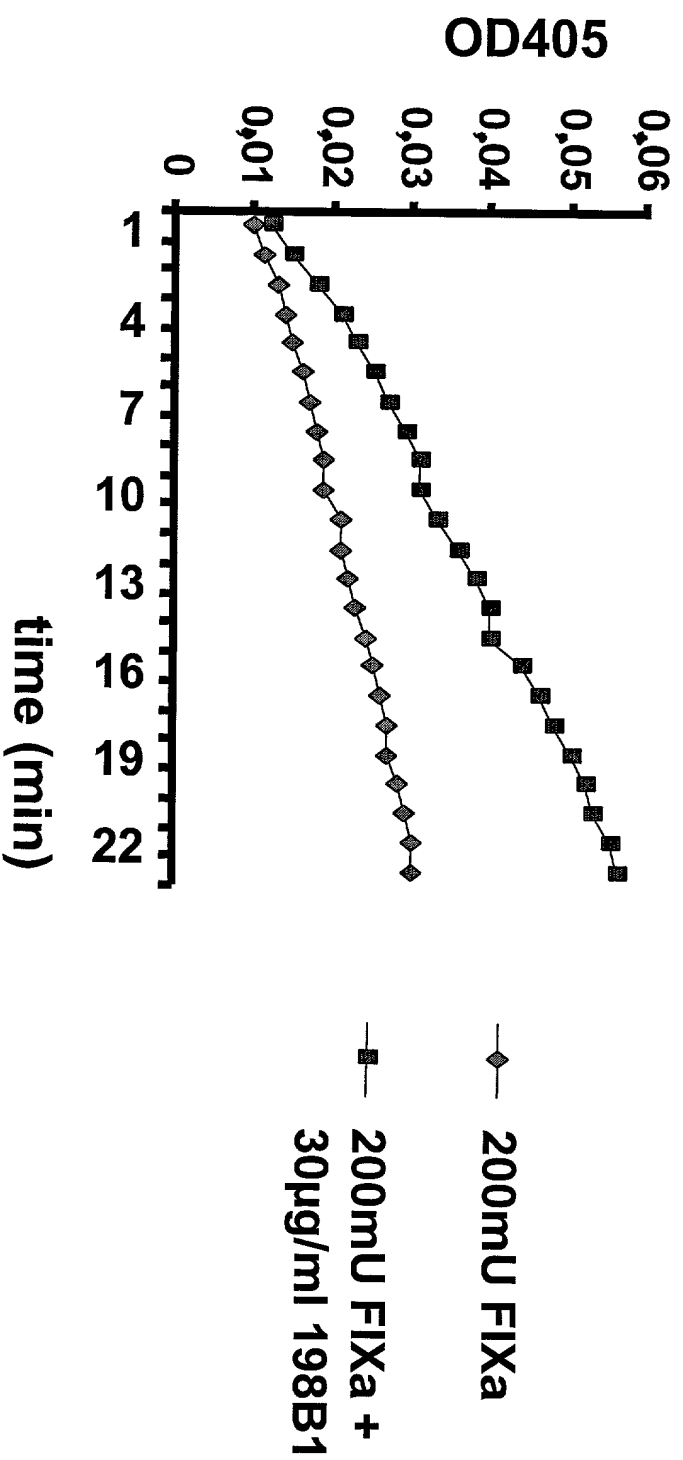


Fig. 24A

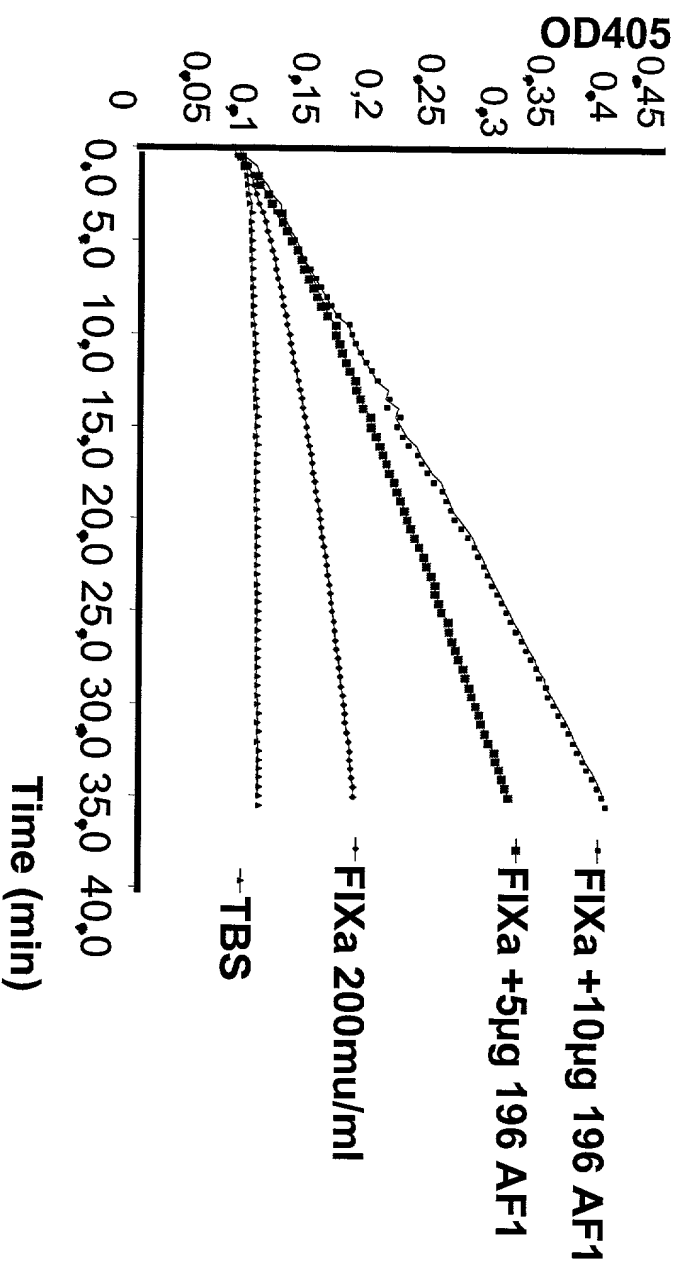


Fig. 24B

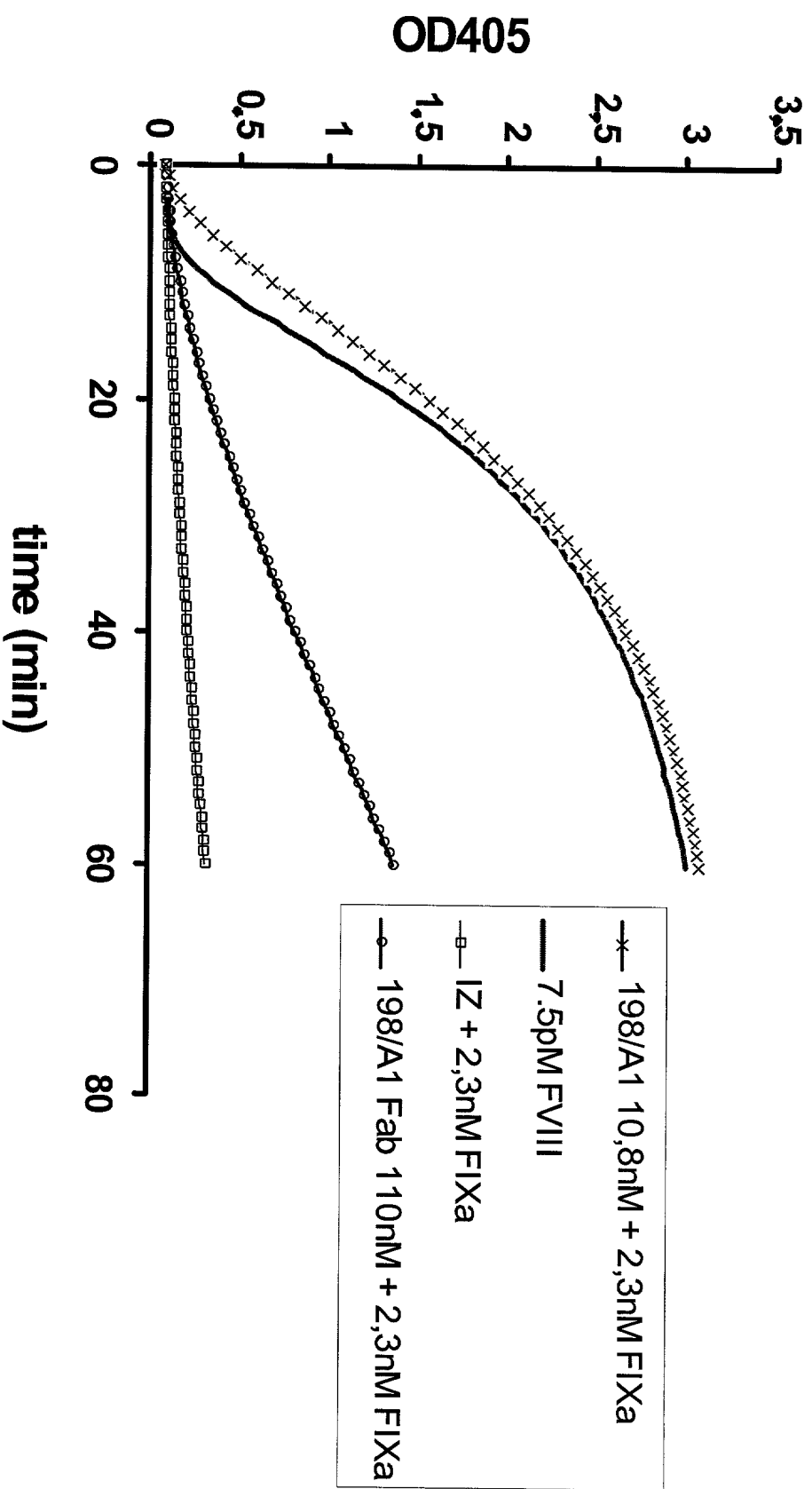


Fig. 25

09661992.091400

00161992-091400

	PelB-leader													
+1	M	K	Y	L	L	P	T	A	A	A	G	L	L	L
1	ATG	AAA	TAC	CTA	TTG	CCT	ACG	GCA	GCC	GCT	GGA	TTG	TTA	TTA
	TAC	TTT	ATG	GAT	AAC	GGA	TGC	CGT	CGG	CGA	CCT	AAC	AAT	AAT
VH														
+1	L	A	A	Q	P	A	M	A	E	V	K	L	V	E
43	CTC	GCG	GCC	CAG	CCG	GCC	ATG	GCG	GAG	GTG	AAG	CTG	GTG	GAG
	GAG	CGC	CGG	GTC	GGC	CGG	TAC	CGC	CTC	CAC	TTC	GAC	CAC	CTC
+1	S	G	G	G	L	V	K	P	G	G	S	L	K	L
85	TCT	GGG	GGA	GGC	TTA	GTG	AAG	CCT	GGA	GGG	TCC	CTG	AAA	CTC
	AGA	CCC	CCT	CCG	AAT	CAC	TTC	GGA	CCT	CCC	AGG	GAC	TTT	GAG
+1	S	C	A	A	S	G	F	T	F	S	S	Y	T	M
127	TCC	TGT	GCA	GCC	TCT	GGA	TTC	ACT	TTC	AGT	AGC	TAT	ACC	ATG
	AGG	ACA	CGT	CGG	AGA	CCT	AAG	TGA	AAG	TCA	TCG	ATA	TGG	TAC
+1	S	W	V	R	Q	T	P	E	K	R	L	E	W	V
169	TCT	TGG	GTT	CGC	CAG	ACT	CCG	GAG	AAG	AGG	CTG	GAG	TGG	GTC
	AGA	ACC	CAA	GCG	GTC	TGA	GGC	CTC	TTC	TCC	GAC	CTC	ACC	CAG
+1	A	T	I	S	S	G	G	S	S	T	Y	Y	P	D
211	GCA	ACC	ATT	AGT	AGT	GGN	GGT	AGT	TCC	ACC	TAC	TAT	CCA	GAC
	CGT	TGG	TAA	TCA	TCA	CCN	CCA	TCA	AGG	TGG	ATG	ATA	GGT	CTG
+1	S	V	K	G	R	F	T	I	S	R	D	N	A	K
253	AGT	GTG	AAG	GGC	CGA	TTC	ACC	ATC	TCC	AGA	GAC	AAT	GCC	AAG
	TCA	CAC	TTC	CCG	GCT	AAG	TGG	TAG	AGG	TCT	CTG	TTA	CGG	TTC
+1	N	T	L	Y	L	Q	M	S	S	L	R	S	E	D
295	AAC	ACC	CTG	TAC	CTG	CAA	ATG	AGC	AGT	CTG	AGG	TCT	GAG	GAC
	TTG	TGG	GAC	ATG	GAC	GTT	TAC	TCG	TCA	GAC	TCC	AGA	CTC	CTG
+1	T	A	M	Y	Y	C	T	R	E	G	G	G	F	T
337	ACA	GCC	ATG	TAT	TAC	TGT	ACA	AGA	GAG	GGG	GGT	GGT	TTC	ACC
	TGT	CGG	TAC	ATA	ATG	ACA	TGT	TCT	CTC	CCC	CCA	CCA	AAG	TGG
+1	V	N	W	Y	F	D	V	W	G	A	G	T	S	V
379	GTC	AAC	TGG	TAC	TTC	GAT	GTC	TGG	GGC	GCA	GGA	ACC	TCA	GTC
	CAG	TTG	ACC	ATG	AAG	CTA	CAG	ACC	CCG	CGT	CCT	TGG	AGT	CAG
Linker														
+1	T	V	S	S	G	G	G	G	S	G	G	R	A	S
421	ACC	GTC	TCC	TCA	GGT	GGA	GGC	GGT	TCA	GGT	GGG	CGC	GCC	TCT
	TGG	CAG	AGG	AGT	CCA	CCT	CCG	CCA	AGT	CCA	CCC	GCG	CGG	AGA

FIG 26-1

	+1	G	G	G	G	S	VL									
463		GGC	GGT	GGC	GGA	TCG	D	I	V	L	T	Q	S	P	A	
		CCG	CCA	CCG	CCT	AGC	GAC	ATT	GTG	CTG	ACA	CAG	TCT	CCA	GCT	
							CTG	TAA	CAC	GAC	TGT	GTC	AGA	GGT	CGA	
	+1	S	L	A	V	S	L	G	Q	R	A	T	I	S	C	
505		TCT	TTG	GCT	GTG	TCT	CTA	GGG	CAG	AGG	GCC	ACC	ATA	TCC	TGC	
		AGA	AAC	CGA	CAC	AGA	GAT	CCC	GTC	TCC	CGG	TGG	TAT	AGG	ACG	
	+1	R	A	S	E	S	V	D	S	Y	G	Y	N	F	M	
547		AGA	GCC	AGT	GAA	AGT	GTT	GAT	AGT	TAT	GGC	TAT	AAT	TTT	ATG	
		TCT	CGG	TCA	CTT	TCA	CAA	CTA	TCA	ATA	CCG	ATA	TTA	AAA	TAC	
	+1	H	W	Y	Q	Q	I	P	G	Q	P	P	K	L	L	
589		CAC	TGG	TAT	CAG	CAG	ATA	CCA	GGA	CAG	CCA	CCC	AAA	CTC	CTC	
		GTG	ACC	ATA	GTC	GTC	TAT	GGT	CCT	GTC	GGT	GGG	TTT	GAG	GAG	
	+1	I	Y	R	A	S	N	L	E	S	G	I	P	A	R	
631		ATC	TAT	CGT	GCA	TCC	AAC	CTA	GAG	TCT	GGG	ATC	CCT	GCC	AGG	
		TAG	ATA	GCA	CGT	AGG	TTG	GAT	CTC	AGA	CCC	TAG	GGA	CGG	TCC	
	+1	F	S	G	S	G	S	R	T	D	F	T	L	T	I	
673		TTC	AGT	GGC	AGT	GGG	TCT	AGG	ACA	GAC	TTC	ACC	CTC	ACC	ATT	
		AAG	TCA	CCG	TCA	CCC	AGA	TCC	TGT	CTG	AAG	TGG	GAG	TGG	TAA	
	+1	N	P	V	E	A	D	D	V	A	T	Y	Y	C	Q	
715		AAT	CCT	GTG	GAG	GCT	GAT	GAT	GTT	GCA	ACC	TAT	TAC	TGT	CAG	
		TTA	GGA	CAC	CTC	CGA	CTA	CTA	CAA	CGT	TGG	ATA	ATG	ACA	GTC	
	+1	Q	S	N	E	D	P	L	T	F	G	T	G	T	R	
757		CAA	AGT	AAT	GAG	GAT	CCG	CTC	ACG	TTC	GGT	ACT	GGG	ACC	AGA	
		GTT	TCA	TTA	CTC	CTA	GGC	GAG	TGC	AAG	CCA	TGA	CCC	TGG	TCT	
	+1	L	E	I	K	R	Spacer				Alkaline phosphatase					
799		CTG	GAA	ATA	AAA	CGG	A	A	A	A	R	A	P	E	M	
		GAC	CTT	TAT	TTT	GCC	GCG	GCC	GCA	GCC	CGG	GCA	CCA	GAA	ATG	
							CGC	CGG	CGT	CGG	GCC	CGT	GGT	CTT	TAC	
	+1	P	V	L	E	N	R	A	A	Q	G	D	I	T	A	
841		CCT	GTT	CTG	GAA	AAC	CGG	GCT	GCT	CAG	GGC	GAT	ATT	ACT	GCA	
		GGA	CAA	GAC	CTT	TTG	GCC	CGA	CGA	GTC	CCG	CTA	TAA	TGA	CGT	
	+1	P	G	G	A	R	R	L	T	G	D	Q	T	A	A	
883		CCC	GGC	GGT	GCT	CGC	CGT	TTA	ACG	GGT	GAT	CAG	ACT	GCC	GCT	
		GGG	CCG	CCA	CGA	GCG	GCA	AAT	TGC	CCA	CTA	GTC	TGA	CGG	CGA	
	+1	L	R	D	S	L	S	D	K	P	A	K	N	I	I	
925		CTG	CGT	GAT	TCT	CTT	AGC	GAT	AAA	CCT	GCA	AAA	AAT	ATT	ATT	
		GAC	GCA	CTA	AGA	GAA	TCG	CTA	TTT	GGA	CGT	TTT	TTA	TAA	TAA	

+1	L	L	I	G	D	G	M	G	D	S	E	I	T	A
967	TTG	CTG	ATT	GGC	GAT	GGG	ATG	GGG	GAC	TCG	GAA	ATT	ACT	GCC
	AAC	GAC	TAA	CCG	CTA	CCC	TAC	CCC	CTG	AGC	CTT	TAA	TGA	CGG
+1	A	R	N	Y	A	E	G	A	G	G	F	F	K	G
1009	GCA	CGT	AAT	TAT	GCC	GAA	GGT	GCG	GGC	GGC	TTT	TTT	AAA	GGT
	CGT	GCA	TTA	ATA	CGG	CTT	CCA	CGC	CCG	CCG	AAA	AAA	TTT	CCA
+1	I	D	A	L	P	L	T	G	Q	Y	T	H	Y	A
1051	ATA	GAT	GCC	TTA	CCG	CTT	ACC	GGG	CAA	TAC	ACT	CAC	TAT	GCG
	TAT	CTA	CGG	AAT	GGC	GAA	TGG	CCC	GTT	ATG	TGA	GTG	ATA	CGC
+1	L	N	K	K	T	G	K	P	D	Y	V	T	D	S
1093	CTG	AAT	AAA	AAA	ACC	GGC	AAA	CCG	GAC	TAC	GTC	ACC	GAC	TCG
	GAC	TTA	TTT	TTT	TGG	CCG	TTT	GGC	CTG	ATG	CAG	TGG	CTG	AGC
+1	A	A	S	A	T	A	W	S	T	G	V	K	T	Y
1135	GCT	GCA	TCA	GCA	ACC	GCC	TGG	TCA	ACC	GGT	GTC	AAA	ACC	TAT
	CGA	CGT	AGT	CGT	TGG	CGG	ACC	AGT	TGG	CCA	CAG	TTT	TGG	ATA
+1	N	G	A	L	G	V	D	I	H	E	K	D	H	P
1177	AAC	GGC	GCG	CTG	GGC	GTC	GAT	ATT	CAC	GAA	AAA	GAT	CAC	CCA
	TTG	CCG	CGC	GAC	CCG	CAG	CTA	TAA	GTG	CTT	TTT	CTA	GTG	GGT
+1	T	I	L	E	M	A	K	A	A	G	L	A	T	G
1219	ACG	ATT	CTG	GAA	ATG	GCA	AAA	GCC	GCA	GGT	CTG	GCG	ACC	GGT
	TGC	TAA	GAC	CTT	TAC	CGT	TTT	CGG	CGT	CCA	GAC	CGC	TGG	CCA
+1	N	V	S	T	A	E	L	Q	D	A	T	P	A	A
1261	AAC	GTT	TCT	ACC	GCA	GAG	TTG	CAG	GAT	GCC	ACG	CCC	GCT	GCG
	TTG	CAA	AGA	TGG	CGT	CTC	AAC	GTC	CTA	CGG	TGC	GGG	CGA	CGC
+1	L	V	A	H	V	T	S	R	K	C	Y	G	P	S
1303	CTG	GTG	GCA	CAT	GTG	ACC	TCG	CGC	AAA	TGC	TAC	GGT	CCG	AGC
	GAC	CAC	CGT	GTA	CAC	TGG	AGC	GCG	TTT	ACG	ATG	CCA	GGC	TCG
+1	A	T	S	E	K	C	P	G	N	A	L	E	K	G
1345	GCG	ACC	AGT	GAA	AAA	TGT	CCG	GGT	AAC	GCT	CTG	GAA	AAA	GGC
	CGC	TGG	TCA	CTT	TTT	ACA	GGC	CCA	TTG	CGA	GAC	CTT	TTT	CCG
+1	G	K	G	S	I	T	E	Q	L	L	N	A	R	A
1387	GGA	AAA	GGA	TCG	ATT	ACC	GAA	CAG	CTG	CTT	AAC	GCT	CGT	GCC
	CCT	TTT	CCT	AGC	TAA	TGG	CTT	GTC	GAC	GAA	TTG	CGA	GCA	CGG
+1	D	V	T	L	G	G	G	A	K	T	F	A	E	T
1429	GAC	GTT	ACG	CTT	GGC	GGC	GGC	GCA	AAA	ACC	TTT	GCT	GAA	ACG
	CTG	CAA	TGC	GAA	CCG	CCG	CCG	CGT	TTT	TGG	AAA	CGA	CTT	TGC

+1 A T A G E W Q G K T L R E Q
 1471 GCA ACC GCT GGT GAA TGG CAG GGA AAA ACG CTG CGT GAA CAG
 CGT TGG CGA CCA CTT ACC GTC CCT TTT TGC GAC GCA CTT GTC

+1 A Q A R G Y Q L V S D A A S
 1513 GCA CAG GCG CGT GGT TAT CAG TTG GTG AGC GAT GCT GCC TCA
 CGT GTC CGC GCA CCA ATA GTC AAC CAC TCG CTA CGA CGG AGT

+1 L N S V T E A N Q Q K P L L
 1555 CTG AAT TCG GTG ACG GAA GCG AAT CAG CAA AAA CCC CTG CTT
 GAC TTA AGC CAC TGC CTT CGC TTA GTC GTT TTT GGG GAC GAA

+1 G L F A D G N M P V R W L G
 1597 GGC CTG TTT GCT GAC GGC AAT ATG CCA GTG CGC TGG CTA GGA
 CCG GAC AAA CGA CTG CCG TTA TAC GGT CAC GCG ACC GAT CCT

+1 P K A T Y H G N I D K P A V
 1639 CCG AAA GCA ACG TAC CAT GGC AAT ATC GAT AAG CCC GCA GTC
 GGC TTT CGT TGC ATG GTA CCG TTA TAG CTA TTC GGG CGT CAG

+1 T C T P N P Q R N D S V P T
 1681 ACC TGT ACG CCA AAT CCG CAA CGT AAT GAC AGT GTA CCA ACC
 TGG ACA TGC GGT TTA GGC GTT GCA TTA CTG TCA CAT GGT TGG

+1 L A Q M T D K A I E L L S K
 1723 CTG GCG CAG ATG ACC GAC AAA GCC ATT GAA TTG TTG AGT AAA
 GAC CGC GTC TAC TGG CTG TTT CGG TAA CTT AAC AAC TCA TTT

+1 N E K G F F L Q V E G A S I
 1765 AAT GAG AAA GGC TTT TTC CTG CAA GTT GAA GGT GCG TCA ATC
 TTA CTC TTT CCG AAA AAG GAC GTT CAA CTT CCA CGC AGT TAG

+1 D K Q D H A A N P C G Q I G
 1807 GAT AAA CAG GAT CAT GCT GCG AAT CCT TGT GGG CAA ATT GGC
 CTA TTT GTC CTA GTA CGA CGC TTA GGA ACA CCC GTT TAA CCG

+1 E T V D L D E A V Q R A L E
 1849 GAG ACG GTC GAT CTC GAT GAA GCC GTA CAA CGG GCG CTG GAA
 CTC TGC CAG CTA GAG CTA CTT CGG CAT GTT GCC CGC GAC CTT

+1 F A K K E G N T L V I V T A
 1891 TTC GCT AAA AAG GAG GGT AAC ACG CTG GTC ATA GTC ACC GCT
 AAG CGA TTT TTC CTC CCA TTG TGC GAC CAG TAT CAG TGG CGA

+1 D H A H A S Q I V A P D T K
 1933 GAT CAC GCC CAC GCC AGC CAG ATT GTT GCG CCG GAT ACC AAA
 CTA GTG CGG GTG CGG TCG GTC TAA CAA CGC GGC CTA TGG TTT

+1 A P G L T Q A L N T K D G A
 1975 GCT CCG GGC CTC ACC CAG GCG CTA AAT ACC AAA GAT GGC GCA
 CGA GGC CCG GAG TGG GTC CGC GAT TTA TGG TTT CTA CCG CGT

FIG 26-4

	+1	V	M	V	M	S	Y	G	N	S	E	E	D	S	Q	
2017		GTG	ATG	GTG	ATG	AGT	TAC	GGG	AAC	TCC	GAA	GAG	GAT	TCA	CAA	
		CAC	TAC	CAC	TAC	TCA	ATG	CCC	TTG	AGG	CTT	CTC	CTA	AGT	GTT	
	+1	E	H	T	G	S	Q	L	R	I	A	A	Y	G	P	
2059		GAA	CAT	ACC	GGC	AGT	CAG	TTG	CGT	ATT	GCG	GCG	TAT	GGC	CCG	
		CTT	GTA	TGG	CCG	TCA	GTC	AAC	GCA	TAA	CGC	CGC	ATA	CCG	GGC	
	+1	H	A	A	N	V	V	G	L	T	D	Q	T	D	L	
2101		CAT	GCC	GCC	AAT	GTT	GTT	GGA	CTG	ACC	GAC	CAG	ACC	GAT	CTC	
		GTA	CGG	CGG	TTA	CAA	CAA	CCT	GAC	TGG	CTG	GTC	TGG	CTA	GAG	
	+1	F	Y	T	M	K	A	A	L	G	D	I	His tag			
2143		TTC	TAC	ACC	ATG	AAA	GCC	GCT	CTG	GGG	GAT	ATC	A	H	H	
		AAG	ATG	TGG	TAC	TTT	CGG	CGA	GAC	CCC	CTA	TAG	GCA	CAC	CAT	
													CGT	GTG	GTA	
	+1	H	H	H	H	*										
2185		CAC	CAT	CAC	CAT	TAA										
		GTG	GTA	GTG	GTA	ATT										

Fig. 26-5

004760-26579960

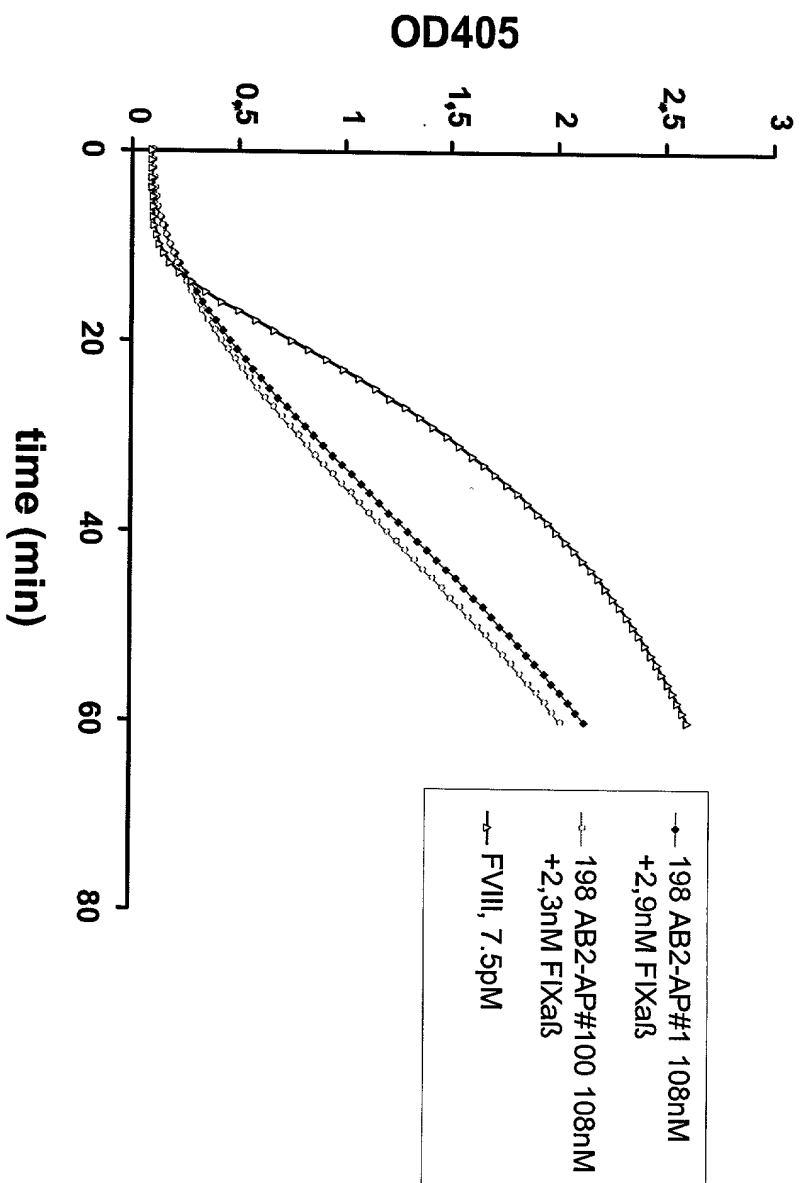


Fig. 27

001750" 255T9960

PelB-Leader

+1 M K Y L L P T A A A G L L L L
 1 ATG AAA TAC CTA TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC
 TAC TTT ATG GAT AAC GGA TGC CGT CGG CGA CCT AAC AAT AAT GAG

VH

+1 A A Q P A M A E V K L V E S G
 46 GCG GCC CAG CCG GCC ATG GCG GAG GTG AAG CTG GTG GAG TCT GGG
 CGC CGG GTC GGC CGG TAC CGC CTC CAC TTC GAC CAC CTC AGA CCC

+1 G G L V K P G G S L K L S C A
 91 GGA GGC TTA GTG AAG CCT GGA GGG TCC CTG AAA CTC TCC TGT GCA
 CCT CCG AAT CAC TTC GGA CCT CCC AGG GAC TTT GAG AGG ACA CGT

+1 A S G F T F S S Y T M S W V R
 136 GCC TCT GGA TTC ACT TTC AGT AGC TAT ACC ATG TCT TGG GTT CGC
 CGG AGA CCT AAG TGA AAG TCA TCG ATA TGG TAC AGA ACC CAA GCG

+1 Q T P E K R L E W V A T I S S
 181 CAG ACT CCG GAG AAG AGG CTG GAG TGG GTC GCA ACC ATT AGT AGT
 GTC TGA GGC CTC TTC TCC GAC CTC ACC CAG CGT TGG TAA TCA TCA

+1 G G S S T Y Y P D S V K G R F
 226 GGN GGT AGT TCC ACC TAC TAT CCA GAC AGT GTG AAG GGC CGA TTC
 CCN CCA TCA AGG TGG ATG ATA GGT CTG TCA CAC TTC CCG GCT AAG

+1 T I S R D N A K N T L Y L Q M
 271 ACC ATC TCC AGA GAC AAT GCC AAG AAC ACC CTG TAC CTG CAA ATG
 TGG TAG AGG TCT CTG TTA CGG TTC TTG TGG GAC ATG GAC GTT TAC

+1 S S L R S E D T A M Y Y C T R
 316 AGC AGT CTG AGG TCT GAG GAC ACA GCC ATG TAT TAC TGT ACA AGA
 TCG TCA GAC TCC AGA CTC CTG TGT CGG TAC ATA ATG ACA TGT TCT

+1 E G G G F T V N W Y F D V W G
 361 GAG GGG GGT GGT TTC ACC GTC AAC TGG TAC TTC GAT GTC TGG GGC
 CTC CCC CCA CCA AAG TGG CAG TTG ACC ATG AAG CTA CAG ACC CCG

Linker

+1 A G T S V T V S S G G G G S G
 406 GCA GGA ACC TCA GTC ACC GTC TCC TCA GGT GGA GGC GGT TCA GGT
 CGT CCT TGG AGT CAG TGG CAG AGG AGT CCA CCT CCG CCA AGT CCA

VL

+1 G R A S G G G S D I V L T Q
 451 GGG CGC GCC TCT GGC GGT GGC GGA TCG GAC ATT GTG CTG ACA CAG
 CCC GCG CGG AGA CCG CCA CCG CCT AGC CTG TAA CAC GAC TGT GTC

F1628-1

+1	X	P	A	S	L	A	V	S	L	G	Q	R	A	T	I
496	TNT	CCA	GCT	TCT	TTG	GCT	GTG	TCT	CTA	GGG	CAG	AGG	GCC	ACC	ATA
	ANA	GGT	CGA	AGA	AAC	CGA	CAC	AGA	GAT	CCC	GTC	TCC	CGG	TGG	TAT
+1	S	C	R	A	S	E	S	V	D	S	Y	G	Y	N	F
541	TCN	TGC	AGA	GCC	AGT	GAA	AGT	GTT	GAT	AGT	TAT	GGC	TAT	AAT	TTT
	AGN	ACG	TCT	CGG	TCA	CTT	TCA	CAA	CTA	TCA	ATA	CCG	ATA	TTA	AAA
+1	M	H	W	Y	Q	Q	I	P	G	Q	P	P	K	L	L
586	ATG	CAC	TGG	TAT	CAG	CAG	ATA	CCA	GGA	CAG	CCA	CCC	AAA	CTC	CTC
	TAC	GTG	ACC	ATA	GTC	GTC	TAT	GGT	CCT	GTC	GGT	GGG	TTT	GAG	GAG
+1	I	Y	R	A	S	N	L	E	S	G	I	P	A	R	F
631	ATC	TAT	CGT	GCA	TCC	AAC	CTA	GAG	TCT	GGG	ATC	CCT	GCC	AGG	TTC
	TAG	ATA	GCA	CGT	AGG	TTG	GAT	CTC	AGA	CCC	TAG	GGA	CGG	TCC	AAG
+1	S	G	S	G	S	R	T	D	F	T	L	T	I	N	P
676	AGT	GGC	AGT	GGG	TCT	AGG	ACA	GAC	TTC	ACC	CTC	ACC	ATT	AAT	CCT
	TCA	CCG	TCA	CCC	AGA	TCC	TGT	CTG	AAG	TGG	GAG	TGG	TAA	TTA	GGA
+1	V	E	A	D	D	V	A	T	Y	Y	C	Q	Q	S	N
721	GTG	GAG	GCT	GAT	GAT	GTT	GCA	ACC	TAT	TAC	TGT	CAG	CAA	AGT	AAT
	CAC	CTC	CGA	CTA	CTA	CAA	CGT	TGG	ATA	ATG	ACA	GTC	GTT	TCA	TTA
+1	E	D	P	L	T	F	G	T	G	T	R	L	E	I	K
766	GAG	GAT	CCG	CTC	ACG	TTC	GGT	ACT	GGG	ACC	AGA	CTG	GAA	ATA	AAA
	CTC	CTA	GGC	GAG	TGC	AAG	CCA	TGA	CCC	TGG	TCT	GAC	CTT	TAT	TTT
	Spacer			Hinge			Helix								
+1	R	A	A	A	P	K	P	S	T	P	P	G	S	S	R
811	CGG	GCG	GCC	GCA	CCG	AAG	CCT	TCC	ACT	CCG	CCC	GGG	TCT	TCC	CGT
	GCC	CGC	CGG	CGT	GGC	TTC	GGA	AGG	TGA	GGC	GGG	CCC	AGA	AGG	GCA
+1	M	K	Q	L	E	D	K	V	E	E	L	L	S	K	N
856	ATG	AAA	CAG	CTG	GAA	GAC	AAA	GTA	GAG	GAG	CTC	CTT	AGC	AAG	AAC
	TAC	TTT	GTC	GAC	CTT	CTG	TTT	CAT	CTC	CTC	GAG	GAA	TCG	TTC	TTG
+1	Y	H	L	E	N	E	V	A	R	L	K	K	L	V	G
901	TAC	CAT	CTA	GAA	AAC	GAG	GTA	GCT	CGT	CTG	AAA	AAG	CTT	GTT	GGT
	ATG	GTA	GAT	CTT	TTG	CTC	CAT	CGA	GCA	GAC	TTT	TTC	GAA	CAA	CCA
	Spacer			His-tag											
+1	E	R	G	G	H	H	H	H	H	H	*				
946	GAA	CGT	GGT	GGT	CAC	CAT	CAC	CAT	CAC	CAT	TAA				
	CTT	GCA	CCA	CCA	GTG	GTA	GTG	GTA	GTG	GTA	ATT				

Fig. 28-2

[illegible]

FIG 29-1

+1	V	S	L	G	Q	R	A	T	I	S	C	R	A	S
505	GTA	TCT	CTG	GGG	CAG	AGG	GCC	ACC	ATC	TCA	TGC	AGG	GCC	AGC
	CAT	AGA	GAC	CCC	GTC	TCC	CGG	TGG	TAG	AGT	ACG	TCC	CGG	TCG
+1	K	S	V	S	T	S	G	Y	S	Y	M	H	W	Y
547	AAA	AGT	GTC	AGT	ACA	TCT	GGC	TAT	AGT	TAT	ATG	CAC	TGG	TAC
	TTT	TCA	CAG	TCA	TGT	AGA	CCG	ATA	TCA	ATA	TAC	GTG	ACC	ATG
+1	Q	Q	K	P	G	Q	P	P	K	L	L	I	Y	L
589	CAA	CAG	AAA	CCA	GGA	CAG	CCA	CCC	AAA	CTC	CTC	ATC	TAT	CTT
	GTT	GTC	TTT	GGT	CCT	GTC	GGT	GGG	TTT	GAG	GAG	TAG	ATA	GAA
+1	A	S	N	L	E	S	G	V	P	A	R	F	S	G
631	GCA	TCC	AAC	CTA	GAA	TCT	GGG	GTC	CCT	GCC	AGG	TTC	AGT	GGC
	CGT	AGG	TTG	GAT	CTT	AGA	CCC	CAG	GGA	CGG	TCC	AAG	TCA	CCG
+1	S	G	S	G	T	D	F	T	L	N	I	H	P	V
673	AGT	GGG	TCT	GGG	ACA	GAC	TTC	ACC	CTC	AAC	ATC	CAT	CCT	GTG
	TCA	CCC	AGA	CCC	TGT	CTG	AAG	TGG	GAG	TTG	TAG	GTA	GGA	CAC
+1	E	E	E	D	A	A	T	Y	Y	C	Q	H	S	R
715	GAG	GAG	GAG	GAT	GCT	GCA	ACC	TAT	TAC	TGT	CAG	CAC	AGT	AGG
	CTC	CTC	CTC	CTA	CGA	CGT	TGG	ATA	ATG	ACA	GTC	GTG	TCA	TCC
+1	E	L	P	R	T	F	G	G	G	T	K	L	E	I
757	GAG	CTT	CCT	CGG	ACG	TTC	GGT	GGA	GGC	ACC	AAG	CTG	GAA	ATC
	CTC	GAA	GGA	GCC	TGC	AAG	CCA	CCT	CCG	TGG	TTC	GAC	CTT	TAG
+1	K	R	Spacer				Alkaline phosphatase							
799	AAA	CGG	A	A	A	A	R	A	P	E	M	P	V	L
	TTT	GCC	GCG	GCC	GCA	GCC	CGG	GCA	CCA	GAA	ATG	CCT	GTT	CTG
			CGC	CGG	CGT	CGG	GCC	CGT	GGT	CTT	TAC	GGA	CAA	GAC
+1	E	N	R	A	A	Q	G	D	I	T	A	P	G	G
841	GAA	AAC	CGG	GCT	GCT	CAG	GGC	GAT	ATT	ACT	GCA	CCC	GGC	GGT
	CTT	TTG	GCC	CGA	CGA	GTC	CCG	CTA	TAA	TGA	CGT	GGG	CCG	CCA
+1	A	R	R	L	T	G	D	Q	T	A	A	L	R	D
883	GCT	CGC	CGT	TTA	ACG	GGT	GAT	CAG	ACT	GCC	GCT	CTG	CGT	GAT
	CGA	GCG	GCA	AAT	TGC	CCA	CTA	GTC	TGA	CGG	CGA	GAC	GCA	CTA
+1	S	L	S	D	K	P	A	K	N	I	I	L	L	I
925	TCT	CTT	AGC	GAT	AAA	CCT	GCA	AAA	AAT	ATT	ATT	TTG	CTG	ATT
	AGA	GAA	TCG	CTA	TTT	GGA	CGT	TTT	TTA	TAA	TAA	AAC	GAC	TAA
+1	G	D	G	M	G	D	S	E	I	T	A	A	R	N
967	GGC	GAT	GGG	ATG	GGG	GAC	TCG	GAA	ATT	ACT	GCC	GCA	CGT	AAT
	CCG	CTA	CCC	TAC	CCC	CTG	AGC	CTT	TAA	TGA	CGG	CGT	GCA	TTA

+1 Y A E G A G G F F K G I D A
 1009 TAT GCC GAA GGT GCG GGC GGC TTT TTT AAA GGT ATA GAT GCC
 ATA CGG CTT CCA CGC CCG CCG AAA AAA TTT CCA TAT CTA CGG

+1 L P L T G Q Y T H Y A L N K
 1051 TTA CCG CTT ACC GGG CAA TAC ACT CAC TAT GCG CTG AAT AAA
 AAT GGC GAA TGG CCC GTT ATG TGA GTG ATA CGC GAC TTA TTT

+1 K T G K P D Y V T D S A A S
 1093 AAA ACC GGC AAA CCG GAC TAC GTC ACC GAC TCG GCT GCA TCA
 TTT TGG CCG TTT GGC CTG ATG CAG TGG CTG AGC CGA CGT AGT

+1 A T A W S T G V K T Y N G A
 1135 GCA ACC GCC TGG TCA ACC GGT GTC AAA ACC TAT AAC GGC GCG
 CGT TGG CCG ACC AGT TGG CCA CAG TTT TGG ATA TTG CCG CGC

+1 L G V D I H E K D H P T I L
 1177 CTG GGC GTC GAT ATT CAC GAA AAA GAT CAC CCA ACG ATT CTG
 GAC CCG CAG CTA TAA GTG CTT TTT CTA GTG GGT TGC TAA GAC

+1 E M A K A A G L A T G N V S
 1219 GAA ATG GCA AAA GCC GCA GGT CTG GCG ACC GGT AAC GTT TCT
 CTT TAC CGT TTT CGG CGT CCA GAC CGC TGG CCA TTG CAA AGA

+1 T A E L Q D A T P A A L V A
 1261 ACC GCA GAG TTG CAG GAT GCC ACG CCC GCT GCG CTG GTG GCA
 TGG CGT CTC AAC GTC CTA CGG TGC GGG CGA CGC GAC CAC CGT

+1 H V T S R K C Y G P S A T S
 1303 CAT GTG ACC TCG CGC AAA TGC TAC GGT CCG AGC GCG ACC AGT
 GTA CAC TGG AGC GCG TTT ACG ATG CCA GGC TCG CGC TGG TCA

+1 E K C P G N A L E K G G K G
 1345 GAA AAA TGT CCG GGT AAC GCT CTG GAA AAA GGC GGA AAA GGA
 CTT TTT ACA GGC CCA TTG CGA GAC CTT TTT CCG CCT TTT CCT

+1 S I T E Q L L N A R A D V T
 1387 TCG ATT ACC GAA CAG CTG CTT AAC GCT CGT GCC GAC GTT ACG
 AGC TAA TGG CTT GTC GAC GAA TTG CGA GCA CGG CTG CAA TGC

+1 L G G G A K T F A E T A T A
 1429 CTT GGC GGC GGC GCA AAA ACC TTT GCT GAA ACG GCA ACC GCT
 GAA CCG CCG CCG CGT TTT TGG AAA CGA CTT TGC CGT TGG CGA

+1 G E W Q G K T L R E Q A Q A
 1471 GGT GAA TGG CAG GGA AAA ACG CTG CGT GAA CAG GCA CAG GCG
 CCA CTT ACC GTC CCT TTT TGC GAC GCA CTT GTC CGT GTC CGC

F16 29-3

004760 26579360

+1	R	G	Y	Q	L	V	S	D	A	A	S	L	N	S
1513	CGT	GGT	TAT	CAG	TTG	GTG	AGC	GAT	GCT	GCC	TCA	CTG	AAT	TCG
	GCA	CCA	ATA	GTC	AAC	CAC	TCG	CTA	CGA	CGG	AGT	GAC	TTA	AGC
+1	V	T	E	A	N	Q	Q	K	P	L	L	G	L	F
1555	GTG	ACG	GAA	GCG	AAT	CAG	CAA	AAA	CCC	CTG	CTT	GGC	CTG	TTT
	CAC	TGC	CTT	CGC	TTA	GTC	GTT	TTT	GGG	GAC	GAA	CCG	GAC	AAA
+1	A	D	G	N	M	P	V	R	W	L	G	P	K	A
1597	GCT	GAC	GGC	AAT	ATG	CCA	GTG	CGC	TGG	CTA	GGA	CCG	AAA	GCA
	CGA	CTG	CCG	TTA	TAC	GGT	CAC	GCG	ACC	GAT	CCT	GGC	TTT	CGT
+1	T	Y	H	G	N	I	D	K	P	A	V	T	C	T
1639	ACG	TAC	CAT	GGC	AAT	ATC	GAT	AAG	CCC	GCA	GTC	ACC	TGT	ACG
	TGC	ATG	GTA	CCG	TTA	TAG	CTA	TTC	GGG	CGT	CAG	TGG	ACA	TGC
+1	P	N	P	Q	R	N	D	S	V	P	T	L	A	Q
1681	CCA	AAT	CCG	CAA	CGT	AAT	GAC	AGT	GTA	CCA	ACC	CTG	GCG	CAG
	GGT	TTA	GGC	GTT	GCA	TTA	CTG	TCA	CAT	GGT	TGG	GAC	CGC	GTC
+1	M	T	D	K	A	I	E	L	L	S	K	N	E	K
1723	ATG	ACC	GAC	AAA	GCC	ATT	GAA	TTG	TTG	AGT	AAA	AAT	GAG	AAA
	TAC	TGG	CTG	TTT	CGG	TAA	CTT	AAC	AAC	TCA	TTT	TTA	CTC	TTT
+1	G	F	F	L	Q	V	E	G	A	S	I	D	K	Q
1765	GGC	TTT	TTC	CTG	CAA	GTT	GAA	GGT	GCG	TCA	ATC	GAT	AAA	CAG
	CCG	AAA	AAG	GAC	GTT	CAA	CTT	CCA	CGC	AGT	TAG	CTA	TTT	GTC
+1	D	H	A	A	N	P	C	G	Q	I	G	E	T	V
1807	GAT	CAT	GCT	GCG	AAT	CCT	TGT	GGG	CAA	ATT	GGC	GAG	ACG	GTC
	CTA	GTA	CGA	CGC	TTA	GGA	ACA	CCC	GTT	TAA	CCG	CTC	TGC	CAG
+1	D	L	D	E	A	V	Q	R	A	L	E	F	A	K
1849	GAT	CTC	GAT	GAA	GCC	GTA	CAA	CGG	GCG	CTG	GAA	TTC	GCT	AAA
	CTA	GAG	CTA	CTT	CGG	CAT	GTT	GCC	CGC	GAC	CTT	AAG	CGA	TTT
+1	K	E	G	N	T	L	V	I	V	T	A	D	H	A
1891	AAG	GAG	GGT	AAC	ACG	CTG	GTC	ATA	GTC	ACC	GCT	GAT	CAC	GCC
	TTC	CTC	CCA	TTG	TGC	GAC	CAG	TAT	CAG	TGG	CGA	CTA	GTG	CGG
+1	H	A	S	Q	I	V	A	P	D	T	K	A	P	G
1933	CAC	GCC	AGC	CAG	ATT	GTT	GCG	CCG	GAT	ACC	AAA	GCT	CCG	GGC
	GTG	CGG	TCG	GTC	TAA	CAA	CGC	GGC	CTA	TGG	TTT	CGA	GGC	CCG
+1	L	T	Q	A	L	N	T	K	D	G	A	V	M	V
1975	CTC	ACC	CAG	GCG	CTA	AAT	ACC	AAA	GAT	GGC	GCA	GTG	ATG	GTG
	GAG	TGG	GTC	CGC	GAT	TTA	TGG	TTT	CTA	CCG	CGT	CAC	TAC	CAC

F16 29-4

+1 M S Y G N S E E D S Q E H T
 2017 ATG AGT TAC GGG AAC TCC GAA GAG GAT TCA CAA GAA CAT ACC
 TAC TCA ATG CCC TTG AGG CTT CTC CTA AGT GTT CTT GTA TGG

+1 G S Q L R I A A Y G P H A A
 2059 GGC AGT CAG TTG CGT ATT GCG GCG TAT GGC CCG CAT GCC GCC
 CCG TCA GTC AAC GCA TAA CGC CGC ATA CCG GGC GTA CGG CGG

+1 N V V G L T D Q T D L F Y T
 2101 AAT GTT GTT GGA CTG ACC GAC CAG ACC GAT CTC TTC TAC ACC
 TTA CAA CAA CCT GAC TGG CTG GTC TGG CTA GAG AAG ATG TGG

	+1	M	K	A	A	L	G	D	I	A		His tag			
2143		ATG	AAA	GCC	GCT	CTG	GGG	GAT	ATC	GCA	CAC	CAT	CAC	CAT	CAC
		TAC	TTT	CGG	CGA	GAC	CCC	CTA	TAG	CGT	GTG	GTA	GTG	GTA	GTG

+1 H *
 2185 CAT TAA
 GTA ATT

Fig. 29-5

004760" 26579360

PelB-Leader

+1 M K Y L L P T A A A G L L L L
 1 ATG AAA TAC CTA TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC
 TAC TTT ATG GAT AAC GGA TGC CGT CGG CGA CCT AAC AAT AAT GAG

VH

+1 A A Q P A M A E V Q L Q Q S G
 46 GCG GCC CAG CCG GCC ATG GCG GAG GTT CAG CTT CAG CAG TCT GGA
 CGC CGG GTC GGC CGG TAC CGC CTC CAA GTC GAA GTC GTC AGA CCT

+1 P E L V K P G A S V K I S C K
 91 CCT GAG CTG GTG AAG CCC GGG GCC TCA GTG AAG ATT TCC TGC AAA
 GGA CTC GAC CAC TTC GGG CCC CGG AGT CAC TTC TAA AGG ACG TTT

+1 A S G Y A F S S S W M N W V K
 136 GCT TCT GGC TAC GCA TTC AGT AGC TCT TGG ATG AAC TGG GTG AAG
 CGA AGA CCG ATG CGT AAG TCA TCG AGA ACC TAC TTG ACC CAC TTC

+1 Q R P G Q G L E W I G R I Y P
 181 CAG AGG CCT GGA CAG GGT CTT GAG TGG ATT GGA CGG ATT TAT CCT
 GTC TCC GGA CCT GTC CCA GAA CTC ACC TAA CCT GCC TAA ATA GGA

+1 G N G D T N Y N G K F K G K A
 226 GGA AAT GGA GAT ACT AAC TAC AAT GGG AAG TTC AAG GGC AAG GCC
 CCT TTA CCT CTA TGA TTG ATG TTA CCC TTC AAG TTC CCG TTC CGG

+1 T L T A D K S S S T A Y M Q L
 271 ACA CTG ACT GCA GAC AAA TCC TCC AGC ACA GCC TAC ATG CAG CTC
 TGT GAC TGA CGT CTG TTT AGG AGG TCG TGT CGG ATG TAC GTC GAG

+1 S S L T S V D S A V Y F C A D
 316 AGC AGC CTG ACC TCT GTG GAC TCT GCG GTC TAT TTC TGT GCA GAT
 TCG TCG GAC TGG AGA CAC CTG AGA CGC CAG ATA AAG ACA CGT CTA

+1 G N V Y Y Y A M D Y W G Q G T
 361 GGT AAC GTA TAT TAC TAT GCT ATG GAC TAC TGG GGT CAA GGA ACC
 CCA TTG CAT ATA ATG ATA CGA TAC CTG ATG ACC CCA GTT CCT TGG

Linker

+1 S V T V S S G G G G S G G R A
 406 TCA GTC ACC GTC TCC TCA GGT GGA GGC GGT TCA GGT GGG CGC GCC
 AGT CAG TGG CAG AGG AGT CCA CCT CCG CCA AGT CCA CCC GCG CGG

FIG 30-1

	+1	S	G	G	G	G	S	VL								Q	I	V	L	T	Q	S	P	A							
451		TCT	GGC	GGT	GGC	GGA	TCG	CAA	ATT	GTT	CTC	ACC	CAG	TCT	CCT	GCT	AGA	CCG	CCA	CCG	CCT	AGC	GTT	TAA	CAA	GAG	TGG	GTC	AGA	GGA	CGA
	+1	S	L	A	V	S	L	G	Q	R	A	T	I	S	C	R															
496		TCC	TTA	GCT	GTA	TCT	CTG	GGG	CAG	AGG	GCC	ACC	ATC	TCA	TGC	AGG	AGG	AAT	CGA	CAT	AGA	GAC	CCC	GTC	TCC	CGG	TGG	TAG	AGT	ACG	TCC
	+1	A	S	K	S	V	S	T	S	G	Y	S	Y	M	H	W															
541		GCC	AGC	AAA	AGT	GTC	AGT	ACA	TCT	GGC	TAT	AGT	TAT	ATG	CAC	TGG	CGG	TCG	TTT	TCA	CAG	TCA	TGT	AGA	CCG	ATA	TCA	ATA	TAC	GTG	ACC
	+1	Y	Q	Q	K	P	G	Q	P	P	K	L	L	I	Y	L															
586		TAC	CAA	CAG	AAA	CCA	GGA	CAG	CCA	CCC	AAA	CTC	CTC	ATC	TAT	CTT	ATG	GTT	GTC	TTT	GGT	CCT	GTC	GGT	GGG	TTT	GAG	GAG	TAG	ATA	GAA
	+1	A	S	N	L	E	S	G	V	P	A	R	F	S	G	S															
631		GCA	TCC	AAC	CTA	GAA	TCT	GGG	GTC	CCT	GCC	AGG	TTC	AGT	GGC	AGT	CGT	AGG	TTG	GAT	CTT	AGA	CCC	CAG	GGA	CGG	TCC	AAG	TCA	CCG	TCA
	+1	G	S	G	T	D	F	T	L	N	I	H	P	V	E	E															
676		GGG	TCT	GGG	ACA	GAC	TTC	ACC	CTC	AAC	ATC	CAT	CCT	GTG	GAG	GAG	CCC	AGA	CCC	TGT	CTG	AAG	TGG	GAG	TTG	TAG	GTA	GGA	CAC	CTC	CTC
	+1	E	D	A	A	T	Y	Y	C	Q	H	S	R	E	L	P															
721		GAG	GAT	GCT	GCA	ACC	TAT	TAC	TGT	CAG	CAC	AGT	AGG	GAG	CTT	CCT	CTC	CTA	CGA	CGT	TGG	ATA	ATG	ACA	GTC	GTG	TCA	TCC	CTC	GAA	GGA
	+1	R	T	F	G	G	G	T	K	L	E	I	K	R	Spacer																
766		CGG	ACG	TTC	GGT	GGA	GGC	ACC	AAG	CTG	GAA	ATC	AAA	CGG	GCG	GCC	GCC	TGC	AAG	CCA	CCT	CCG	TGG	TTC	GAC	CTT	TAG	TTT	GCC	CGC	CGG
	+1	A	Hinge										Helix																		
811		GCA	CCG	AAG	CCT	TCC	ACT	CCG	CCC	GGG	TCT	TCC	CGT	ATG	AAA	CAG	CGT	GGC	TTC	GGA	AGG	TGA	GGC	GGG	CCC	AGA	AGG	GCA	TAC	TTT	GTC
	+1	L	E	D	K	V	E	E	L	L	S	K	N	Y	H	L															
856		CTG	GAA	GAC	AAA	GTA	GAG	GAG	CTC	CTT	AGC	AAG	AAC	TAC	CAT	CTA	GAC	CTT	CTG	TTT	CAT	CTC	CTC	GAG	GAA	TCG	TTC	TTG	ATG	GTA	GAT
	+1	E	N	E	V	A	R	L	K	K	L	V	G	E	R	G															
901		GAA	AAC	GAG	GTA	GCT	CGT	CTG	AAA	AAG	CTT	GTT	GGT	GAA	CGT	GGT	CTT	TTG	CTC	CAT	CGA	GCA	GAC	TTT	TTC	GAA	CAA	CCA	CTT	GCA	CCA

F1630-2

	Spacer	His-tag						
+1	G	H	H	H	H	H	H	*
946	GGT	CAC	CAT	CAC	CAT	CAC	CAT	TAA
	CCA	GTG	GTA	GTG	GTA	GTG	GTA	ATT

Fig. 30 - 3

004T60"26T9960

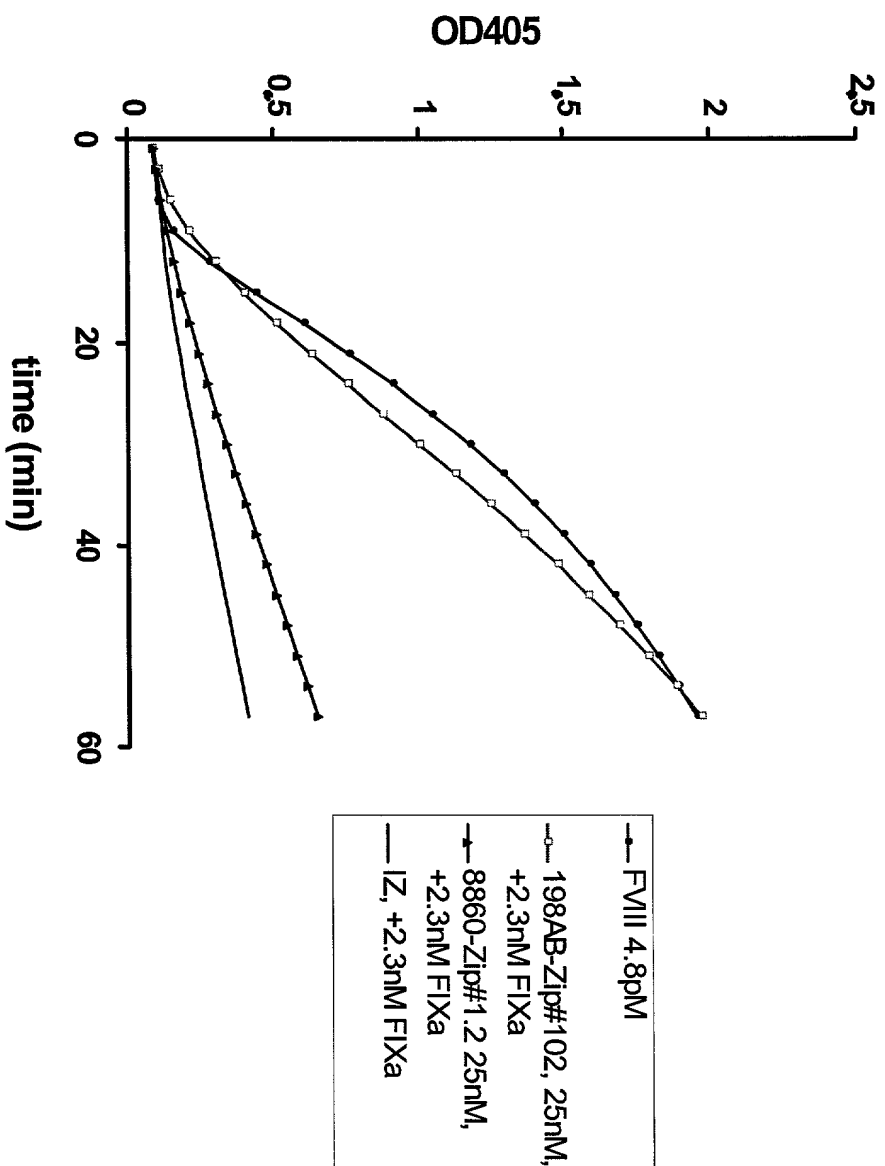


Fig. 31

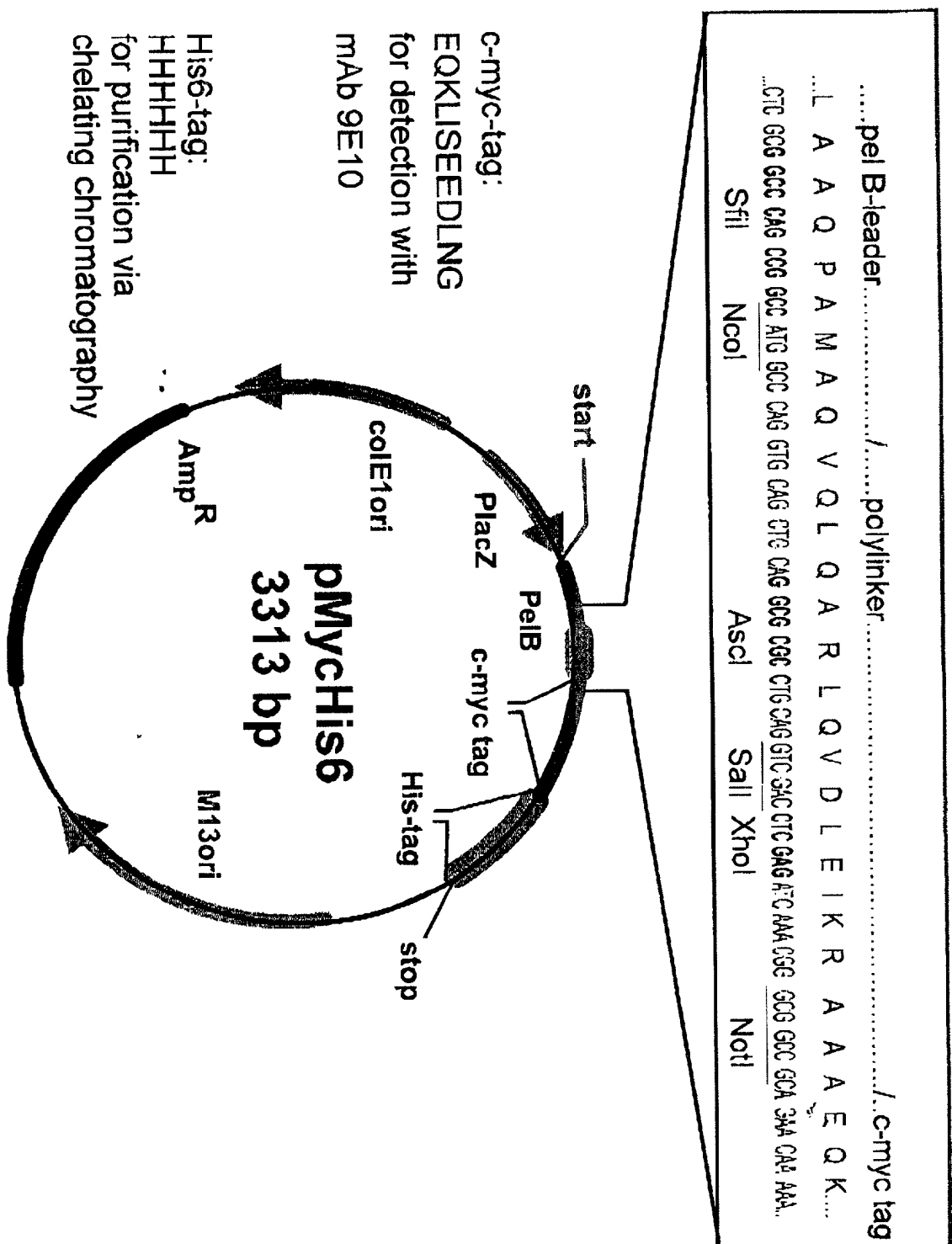


Fig. 32

09661992.091400

004760"265T9960

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      PelB-leader
+1  M   K   Y   L   L   P   T   A   A   A   G   L   L   L   L
1  ATG AAA TAC CTA TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC
   TAC TTT ATG GAT AAC GGA TGC CGT CGG CGA CCT AAC AAT AAT GAG

      VH
+1  A   A   Q   P   A   M   A   E   V   K   L   V   E   S   G
46 GCG GCC CAG CCG GCC ATG GCC GAG GTG AAG CTG GTG GAG TCT GGG
   CGC CGG GTC GGC CGG TAC CGG CTC CAC TTC GAC CAC CTC AGA CCC

+1  G   G   L   V   K   P   G   G   S   L   K   L   S   C   A
91 GGA GGC TTA GTG AAG CCT GGA GGG TCC CTG AAA CTC TCC TGT GCA
   CCT CCG AAT CAC TTC GGA CCT CCC AGG GAC TTT GAG AGG ACA CGT

+1  A   S   G   F   T   F   S   S   Y   T   M   S   W   V   R
136 GCC TCT GGA TTC ACT TTC AGT AGC TAT ACC ATG TCT TGG GTT CGC
   CGG AGA CCT AAG TGA AAG TCA TCG ATA TGG TAC AGA ACC CAA GCG

+1  Q   T   P   E   K   R   L   E   W   V   A   T   I   S   S
181 CAG ACT CCG GAG AAG AGG CTG GAG TGG GTC GCA ACC ATT AGT AGT
   GTC TGA GGC CTC TTC TCC GAC CTC ACC CAG CGT TGG TAA TCA TCA

+1  G   G   S   S   T   Y   Y   P   D   S   V   K   G   R   F
226 GGN GGT AGT TCC ACC TAC TAT CCA GAC AGT GTG AAG GGC CGA TTC
   CCN CCA TCA AGG TGG ATG ATA GGT CTG TCA CAC TTC CCG GCT AAG

+1  T   I   S   R   D   N   A   K   N   T   L   Y   L   Q   M
271 ACC ATC TCC AGA GAC AAT GCC AAG AAC ACC CTG TAC CTG CAA ATG
   TGG TAG AGG TCT CTG TTA CGG TTC TTG TGG GAC ATG GAC GTT TAC

+1  S   S   L   R   S   E   D   T   A   M   Y   Y   C   T   R
316 AGC AGT CTG AGG TCT GAG GAC ACA GCC ATG TAT TAC TGT ACA AGA
   TCG TCA GAC TCC AGA CTC CTG TGT CGG TAC ATA ATG ACA TGT TCT

+1  E   G   G   G   F   T   V   N   W   Y   F   D   V   W   G
361 GAG GGG GGT GGT TTC ACC GTC AAC TGG TAC TTC GAT GTC TGG GGC
   CTC CCC CCA CCA AAG TGG CAG TTG ACC ATG AAG CTA CAG ACC CCG

      Leader
+1  A   G   T   S   V   T   V   S   S   G   G   G   G   S   G
406 GCA GGA ACC TCA GTC ACC GTC TCC TCA GGT GGA GGC GGT TCA GGT
   CGT CCT TGG AGT CAG TGG CAG AGG AGT CCA CCT CCG CCA AGT CCA

      VK
+1  G   R   A   S   G   G   G   G   S   D   I   V   L   T   Q
451 GGG CGC GCC TCT GGC GGT GGC GGA TCG GAC ATT GTG CTG ACA CAG

```

FIG 34-1

CCC GCG CGG AGA CCG CCA CCG CCT AGC CTG TAA CAC GAC TGT GTC

+1 S P A S L A V S L G Q R A T I
 496 TCT CCA GCT TCT TTG GCT GTG TCT CTA GGG CAG AGG GCC ACC ATA
 AGA GGT CGA AGA AAC CGA CAC AGA GAT CCC GTC TCC CGG TGG TAT

+1 S C R A S E S V D S Y G Y N F
 541 TCC TGC AGA GCC AGT GAA AGT GTT GAT AGT TAT GGC TAT AAT TTT
 AGG ACG TCT CGG TCA CTT TCA CAA CTA TCA ATA CCG ATA TTA AAA

+1 M H W Y Q Q I P G Q P P K L L
 586 ATG CAC TGG TAT CAG CAG ATA CCA GGA CAG CCA CCC AAA CTC CTC
 TAC GTG ACC ATA GTC GTC TAT GGT CCT GTC GGT GGG TTT GAG GAG

+1 I Y R A S N L E S G I P A R F
 631 ATC TAT CGT GCA TCC AAC CTA GAG TCT GGG ATC CCT GCC AGG TTC
 TAG ATA GCA CGT AGG TTG GAT CTC AGA CCC TAG GGA CGG TCC AAG

+1 S G S G S R T D F T L T I N P
 676 AGT GGC AGT GGG TCT AGG ACA GAC TTC ACC CTC ACC ATT AAT CCT
 TCA CCG TCA CCC AGA TCC TGT CTG AAG TGG GAG TGG TAA TTA GGA

+1 V E A D D V A T Y Y C Q Q S N
 721 GTG GAG GCT GAT GAT GTT GCA ACC TAT TAC TGT CAG CAA AGT AAT
 CAC CTC CGA CTA CTA CAA CGT TGG ATA ATG ACA GTC GTT TCA TTA

+1 E D P L T F G T G T R L E I K
 766 GAG GAT CCG CTC ACG TTC GGT ACT GGG ACC AGA CTG GAA ATA AAA
 CTC CTA GGC GAG TGC AAG CCA TGA CCC TGG TCT GAC CTT TAT TTT

		Spacer			Myc-tag										
+1	R	A	A	A	E	Q	K	L	I	S	E	E	D	L	N
811	CGG	GCG	GCC	GCA	GAA	CAA	AAA	CTC	ATC	TCA	GAA	GAG	GAT	CTG	AAT
	GCC	CGC	CGG	CGT	CTT	GTT	TTT	GAG	TAG	AGT	CTT	CTC	CTA	GAC	TTA

		Spacer		His tag							
+1	G	A	A	H	H	H	H	H	H	*	*
856	GGG	GCG	GCA	CAT	CAC	CAT	CAC	CAT	CAC	TAA	TAA
	CCC	CGC	CGT	GTA	GTG	GTA	GTG	GTA	GTG	ATT	ATT

Fig. 34-2

PelB-leader

+1 M K Y L L P T A A A G L L L L
 1 ATG AAA TAC CTA TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC
 TAC TTT ATG GAT AAC GGA TGC CGT CGG CGA CCT AAC AAT AAT GAG

+1 A A Q P A M A | VH
 46 GCG GCC CAG CCG GCC ATG GCC | E V Q L Q Q S G
 CGC CGG GTC GGC CGG TAC CGG | CTC CAA GTC GAA GTC GTC AGA CCT

+1 P E L V K P G A S V K I S C K
 91 CCT GAG CTG GTG AAG CCC GGG GCC TCA GTG AAG ATT TCC TGC AAA
 GGA CTC GAC CAC TTC GGG CCC CGG AGT CAC TTC TAA AGG ACG TTT

+1 A S G Y A F S S S W M N W V K
 136 GCT TCT GGC TAC GCA TTC AGT AGC TCT TGG ATG AAC TGG GTG AAG
 CGA AGA CCG ATG CGT AAG TCA TCG AGA ACC TAC TTG ACC CAC TTC

+1 Q R P G Q G L E W I G R I Y P
 181 CAG AGG CCT GGA CAG GGT CTT GAG TGG ATT GGA CGG ATT TAT CCT
 GTC TCC GGA CCT GTC CCA GAA CTC ACC TAA CCT GCC TAA ATA GGA

+1 G N G D T N Y N G K F K G K A
 226 GGA AAT GGA GAT ACT AAC TAC AAT GGG AAG TTC AAG GGC AAG GCC
 CCT TTA CCT CTA TGA TTG ATG TTA CCC TTC AAG TTC CCG TTC CGG

+1 T L T A D K S S S T A Y M Q L
 271 ACA CTG ACT GCA GAC AAA TCC TCC AGC ACA GCC TAC ATG CAG CTC
 TGT GAC TGA CGT CTG TTT AGG AGG TCG TGT CGG ATG TAC GTC GAG

+1 S S L T S V D S A V Y F C A D
 316 AGC AGC CTG ACC TCT GTG GAC TCT GCG GTC TAT TTC TGT GCA GAT
 TCG TCG GAC TGG AGA CAC CTG AGA CGC CAG ATA AAG ACA CGT CTA

+1 G N V Y Y Y A M D Y W G Q G T
 361 GGT AAC GTA TAT TAC TAT GCT ATG GAC TAC TGG GGT CAA GGA ACC
 CCA TTG CAT ATA ATG ATA CGA TAC CTG ATG ACC CCA GTT CCT TGG

+1 S V T V S S | Leader
 406 TCA GTC ACC GTC TCC TCA | G G G G S G G R A
 AGT CAG TGG CAG AGG AGT | GGT GGA GGC GGT TCA GGT GGG CGC GCC
 CCA CCT CCG CCA AGT CCA CCC GCG CGG

+1 S G G G G S | VL
 451 TCT GGC GGT GGC GGA TCG | Q I V L T Q S P A
 AGA CCG CCA CCG CCT AGC | CAA ATT GTT CTC ACC CAG TCT CCT GCT
 GTT TAA CAA GAG TGG GTC AGA GGA CGA

F1635-1

+1 S L A V S L G Q R A T I S C R
 496 TCC TTA GCT GTA TCT CTG GGG CAG AGG GCC ACC ATC TCA TGC AGG
 AGG AAT CGA CAT AGA GAC CCC GTC TCC CGG TGG TAG AGT ACG TCC

+1 A S K S V S T S G Y S Y M H W
 541 GCC AGC AAA AGT GTC AGT ACA TCT GGC TAT AGT TAT ATG CAC TGG
 CGG TCG TTT TCA CAG TCA TGT AGA CCG ATA TCA ATA TAC GTG ACC

+1 Y Q Q K P G Q P P K L L I Y L
 586 TAC CAA CAG AAA CCA GGA CAG CCA CCC AAA CTC CTC ATC TAT CTT
 ATG GTT GTC TTT GGT CCT GTC GGT GGG TTT GAG GAG TAG ATA GAA

+1 A S N L E S G V P A R F S G S
 631 GCA TCC AAC CTA GAA TCT GGG GTC CCT GCC AGG TTC AGT GGC AGT
 CGT AGG TTG GAT CTT AGA CCC CAG GGA CGG TCC AAG TCA CCG TCA

+1 G S G T D F T L N I H P V E E
 676 GGG TCT GGG ACA GAC TTC ACC CTC AAC ATC CAT CCT GTG GAG GAG
 CCC AGA CCC TGT CTG AAG TGG GAG TTG TAG GTA GGA CAC CTC CTC

+1 E D A A T Y Y C Q H S R E L P
 721 GAG GAT GCT GCA ACC TAT TAC TGT CAG CAC AGT AGG GAG CTT CCT
 CTC CTA CGA CGT TGG ATA ATG ACA GTC GTG TCA TCC CTC GAA GGA

	+1	R	T	F	G	G	G	T	K	L	E	I	K	R		Spacer
766		CGG	ACG	TTC	GGT	GGA	GGC	ACC	AAG	CTG	GAA	ATC	AAA	CGG		A A
		GCC	TGC	AAG	CCA	CCT	CCG	TGG	TTC	GAC	CTT	TAG	TTT	GCC		GCG GCC
																CGC CGG

	+1	A	Myc-tag											Spacer		
811		GCA	E	Q	K	L	I	S	E	E	D	L	N	G		A A
		CGT	GAA	CAA	AAA	CTC	ATC	TCA	GAA	GAG	GAT	CTG	AAT	GGG		GCG GCA
			CTT	GTT	TTT	GAG	TAG	AGT	CTT	CTC	CTA	GAC	TTA	CCC		CGC CGT

His tag
 +1 H H H H H H *
 856 CAT CAC CAT CAC CAT CAC TAA
 GTA GTG GTA GTG GTA GTG ATT

Fig. 35-2

004750" 25579960

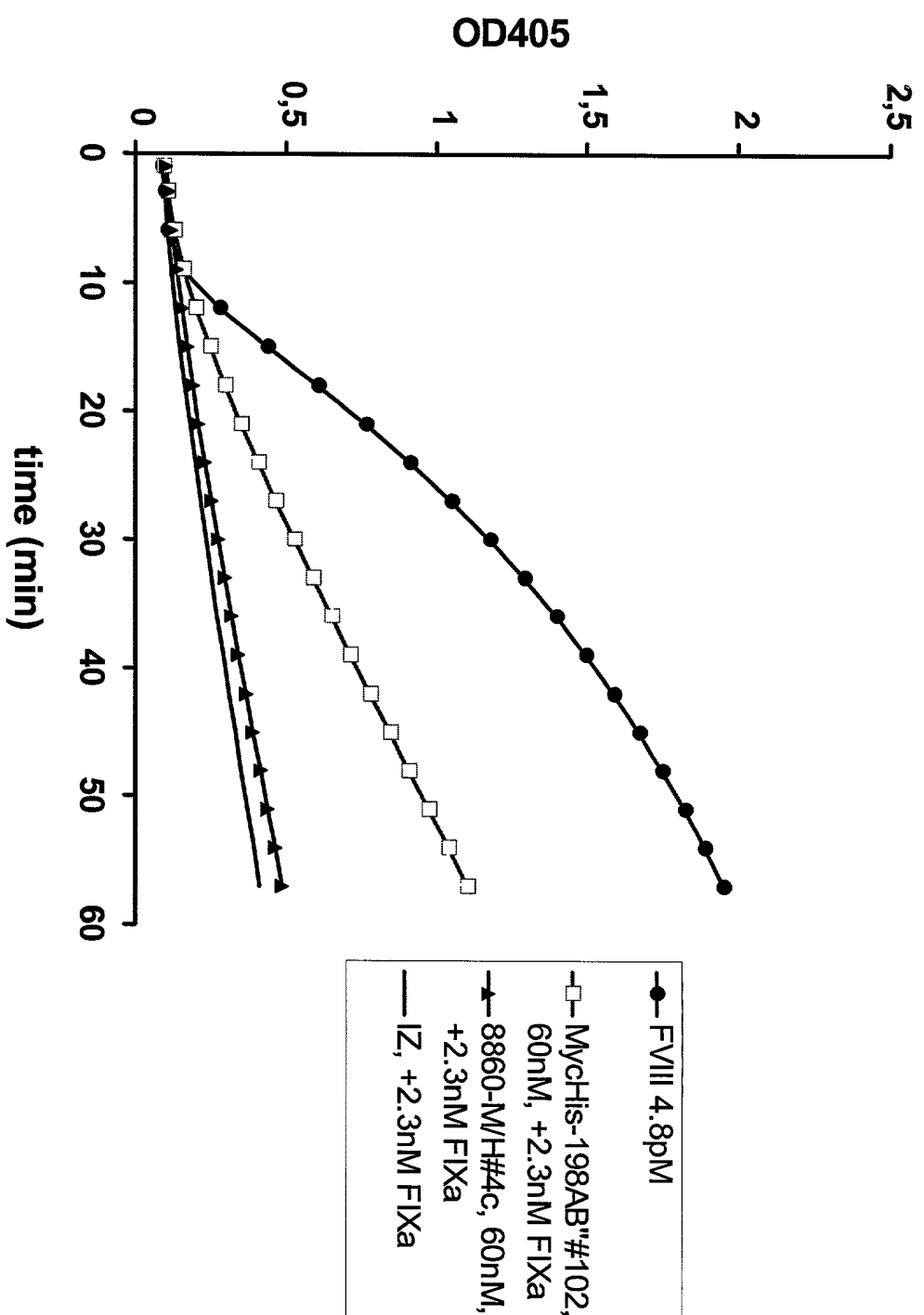


Fig. 36